Prevention and control of multi-drug-resistant Gram-negative bacteria: recommendations from a Joint Working Party

Appendices A–E

Contents

Prevention and control of multi-drug-resistant Gram-negative bacteria: recommendations from a
Joint Working Party
Appendices A–E
Appendix A. Scope
1 Guideline title
1.1 Short title
2. The remit
2.1 Population
2.2 Healthcare setting
2.3 Clinical management4
2.4 Main outcomes
2.5 Economic aspects
2.6 Status5
3. Related NICE guidance5
4. Further information5
Appendix B. Declarations of interest6
B.1 Introduction6
B.2 Peter Wilson (Secretary)6
B.3 David Livermore
B.4 Beryl Oppenheim:6
B.5 David Enoch:7
B.6 Cliodna McNulty7
B.7 Jon Otter
B.8 Maria Cann7
B.9 Peter Jenks
Appendix C. Clinical evidence tables9
Antibiotic stewardship
Other infection control measures
-

Selective decontamination
Systematic reviews
Treatment
Appendix D. Excluded clinical studies
Study design
Case-control study42
Case series/report51
Cross-sectional
In-vitro studies
Prospective cohort
Surveillance
Narrative reviews, commentaries or editorials63
Retrospective cohort
Study design not relevant72
Controlled before-after studies without a minimum of two intervention and control sites73
Interrupted time series studies without at least three data points before and after the intervention
Participants not relevant73
Antibiotics used not relevant for the review74
Not multi-drug-resistant infections74
Appendix E. Peer review

Appendix A. Scope

1 Guideline title

Prevention and control of multi-drug-resistant Gram-negative bacteria:

recommendations from a Joint Working Party

1.1 Short title

Control of multi-drug-resistant Gram-negative bacteria

2 The remit

To examine and make recommendations both for treatment and prevention of transmission of multi-drug-resistant (MDR) Gram-negative infections, resulting in the publication of guidelines on:

- current epidemiology and infection control issues; and
- therapeutic issues and antibiotic guidance for treating infections caused by MDR Gram-negative bacteria.

For the purposes of this Working Group, the remit will mainly include infections in critical and non-critical care patients in secondary care. However, the same general principles would apply in community settings, particularly in areas where inappropriate treatment is encouraging selection. The remit does not include management of cystic fibrosis or community outbreaks. Multi-drug resistance among Gram-negative bacteria will be defined as resistance to three or more of the following antimicrobials: ceftazidime, ciprofloxacin, meropenem, gentamicin or piperacillin/tazobactam. Consideration will be given to laboratory testing and susceptibility testing, although only screening and confirmatory tests available in a general microbiology laboratory and not those limited to reference laboratories. The use of antibiotic combinations in the therapy of infections will be considered, particularly oral combinations that can be used in the outpatient setting.

2.1 Population

2.1.1 Groups that will be covered a) Adults

b) Children

c) Infections with the following organisms to evaluate the efficacy of antibiotics to treat communityacquired infections, and infections acquired in secondary or tertiary care that are caused by MDR Gram-negative bacteria.

Specific antibiotics: Whenever possible, antibiotics were separated as follows:

'Standard' antibiotics currently in use for which there is not much question on efficacy used as comparator: most cephalosporins, coamoxiclav, piperacillin/tazobactam quinolones, temocillin (pivmecillinam is the oral formulation of mecillinam).

Old antibiotics that have been re-introduced: aminoglycosides (including gentamicin and amikacin), colistin, fosfomycin, nitrofurantoin.

Recently developed antibiotics: tigecycline, cefepime, few very new cephalosporins (e.g. ceftobiprole), the newest carbapenems or those in testing (e.g. doripenem).

Specific pathogens: Escherichia coli, Klebsiella spp. including Klebsiella pneumoniae, Enterobacter spp., Pseudomonas aeruginosa, Acinetobacter spp., Proteus spp., Serratia spp., Citrobacter freundii, Morganella morgagnii.

2.1.2 Groups that will not be covered

Gonococci are Gram-negative and are increasingly resistant, but were excluded because relevant public health control actions are substantially different.

2.2 Healthcare setting All settings in which National Health Service care is received.

2.3 Clinical management

2.3.1 Key clinical issues that will be covered

a) Antimicrobial treatment of MDR Gram-negative infections

b) Antimicrobial stewardship

c) Epidemiology

d) Surveillance

e) Infection prevention: standards, hand and environmental hygiene, organizational structures

2.3.2 Clinical issues that will not be covered a) Sexually transmitted diseases

b) Cystic fibrosis

2.4 Main outcomes Recommendations for practice

- a) Surveillance
- b) Screening
- c) Prevention of transmission
- d) Cleaning and environment

2.5 Economic aspects

In most areas, there are no anticipated additional costs unless existing practice falls well below currently accepted best practice. Failure to implement the recommendations would result in greater costs in terms of both economics and quality of life. Screening and isolation will result in significant cost pressures where this is not currently practised, but these costs are set against reduced transmission and fewer cases needing antibiotic treatment. Prolonged isolation can have adverse effects on a patient's psychological health, so may have additional unexpected costs.

2.6 Status

2.6.1 Scope This is the final scope.

2.6.2 *Timing* The development of the guideline recommendation will begin in July 2011.

3. Related NICE guidance

National Institute for Health and Care Excellence. *Infection: prevention and control of healthcare-associated infections in primary and community care.* NICE Clinical Guideline 139. London: NICE; 2012. Available at: http://www.nice.org.uk/guidance/cg139 [last accessed August 2014].

4. Further information

Guideline development process

Scottish Intercollegiate Guidelines Network. *SIGN 50: a guideline developer's handbook.* Revised edition. Edinburgh: Healthcare Improvement Scotland; 2014. Available at: <u>http://www.sign.ac.uk</u> [last accessed December 2014].

Appendix B. Declarations of interest

B.1 Introduction

All members of the Guideline Development Team, expert co-optees and all members of the National Clinical Guidance Centre staff were required to make formal declarations of interest at the outset, and these were updated throughout the development process. With one exception, no interests were declared that required any actions.

B.2 Peter Wilson (Secretary)

First meeting 24/11/11

Consultant on Drug Safety Monitoring Boards for Roche and Genentech, Advisory Panel for 3M

Second meeting 22/10/13, third meeting 12/12/13: no change

No action required

B.3 David Livermore First meeting 24/11/11

Advisory boards or consultancy: Achaogen, Adenium, Alere, Allecra, AstraZeneca, Basilea, Bayer, BioVersys, Cubist, Curetis, Cycle, Discuva, Forest, GSK, Longitude, Meiji, Pfizer, Roche, Shionogi, Tetraphase, VenatoRx, Wockhardt. Paid lectures: AOP Orphan, AstraZeneca, Bruker, Curetis, Merck, Pfizer, Leo. Relevant shareholdings in Dechra, GSK, Merck, Perkin Elmer, Pfizer, collectively amounting to <10% of portfolio value

Second meeting 22/10/13, third meeting 12/12/13: did not attend

No action required

B.4 Beryl Oppenheim First meeting 24/11/11

Advisory board: Astellas, Forrest. Lecture: Alere

Second meeting 22/10/13 did not attend, third meeting 12/12/13: no change

No action required

B.5 David Enoch First meeting 24/11/11

ECCMID conference attendance: funded by Astellas and Eumedica

Second meeting 22/10/13 did not attend, third meeting 12/12/13: no change

No action required

B.6 Cliodna McNulty First meeting 24/11/11

Travel expenses: Merieux Diagnostics

Second meeting 22/10/13, third meeting 12/12/13: no change

No action required

B.7 Jon Otter First meeting 24/11/11 did not attend

Second meeting 22/10/13

Part-time employment at Bioquell. Paid lectures: 3M. Research funding: Pfizer and the Guy's & St Thomas' Charity

Third meeting 12/12/13: no change

Dr Otter did not take part in the section related to the environment, and restricted his advice to that on behalf of the Infection Prevention Society

B.8 Maria Cann First meeting 24/11/11 did not attend

IPS conference attendance: funded by corporate sponsorship from Mölnlycke Healthcare

Second meeting 22/10/13, third meeting 12/12/13: no change

No action required

B.9 Peter Jenks First meeting 24/11/11

Advisory Board: Baxter

Second meeting 22/10/13, third meeting 12/12/13: did not attend

No action required

No declared conflict of interests for the other participants

Appendix C. Clinical evidence tables

Antibiotic	stewardship
------------	-------------

	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
Ben-David 2010 ITS Setting Tertiary (one hospital)	To assess the effect of an intensified intervention, that included active surveillance, on the incidence of infection with carbapenem-resistant <i>K. pneumoniae</i> Participants <i>N</i> =390	Bacteria: K. pneumoniae Resistant to: carbapenems, cephalosporins, fluoroquinolones, trimethoprim-	Intervention 1. Enhanced national infection control programme: contact precautions were used for the care of all patients with CRKP colonization or infection; the prevalence of colonization or infection was reported daily, and	Infection control Before the intervention, the incidence of clinical infection with CRKP had increased 6.42-fold to 6.93 cases per 10,000 patient-days After an enhanced infection control and active surveillance programme	ITS Protection against secular changes (high quality) Protection against
Israel January 2006– December 2008	Age: not reported Male: not reported, female: not reported Inclusion criteria: data from medical records of all patients who acquired CRKP infection Exclusion criteria: not reported	sulfamethoxazole Mechanism of resistance: not reported	this information was mailed to the hospital management and the national coordinator; and patients infected with CRKP had their names entered into a database so that they could be identified at hospital re- admission 2. Active surveillence programme: obtaining rectal culture samples from patients hospitalized in ICUs and in step- down units, at admission to the unit and once weekly until the patient was discharged Length of pre-intervention: 17	was introduced, the incidence of clinical infection reduced to 1.8 cases per 10,000 patient-days (<i>P</i> <0.001). The slope significantly changed with the introduction of the intervention from 0.12 to -0.07 (<i>P</i> <0.001)	detection bias (acceptable quality)
			months prior Length of post-intervention: 19 months following		
Borer 2011	To devise a local strategy for	Bacteria: <i>K.</i>	Intervention	Bacterial colonization and	ITS
ITS	eradication of a hospital-wide outbreak caused by CRKP	pneumoniae Resistant to:	 Emergency department flagging system Building of a cohort space or 	infection During the intervention, the CRKP undetected ratio showed a significant	Protection against secular changes (high
Setting	Participants	carbapenems	ward	increase from 55.7% for June-	quality)

	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
Tertiary (one hospital) Israel May 2006– May 2010	N=803 Adolescents 13–18 years, adults 19–45 years, middle aged 46–64 years, aged 65–79 years, elderly 80+years Male: 410, female: 393 Inclusion criteria: data from medical records of patients with CRKP infection Exclusion criteria: not reported	Mechanism of resistance: not reported	 Intensive active surveillance in high-risk wards Epidemiological investigations Carbapenem-restriction policy Length of pre-intervention: 11 months prior Length of post-intervention: 36 months following 	December 2007 to 71.2% in 2008, 78.9% in 2009 and 92.5% for February– May 2010 ($P \le 0.001$). From May 2006 through April 2007 (pre-intervention), the CRKP-IN incidence density per 10,000 patient- days was 5.26. After the intervention programme was introduced, the incidence of clinical CRPK infection reduced to 2.91 cases per 10,000 patient-days ($P < 0.001$) in 12/2007, 1.91 in 12/2008 and 1.28 in 12/2009. The slope changed significantly with the introduction of the intervention ($P=0.004$). Antibiotic use Meropenem use showed a statistically significant decrease from 2007 to 2010 ($P \le 0.001$); colistin use increased significantly during the same period ($P \le 0.001$)	Protection against detection bias (acceptable to low quality)
Church 2011 ITS Setting Secondary (one hospital) USA January	To assess the possible effects of varying usage of levofloxacin, gatifloxacin and moxifloxacin on <i>P.</i> <i>aeruginosa</i> susceptibility to piperacillin-tazobactam, cefepime and tobramycin Participants <i>N</i> : not reported Age: not reported Male: not reported, female: not reported	Bacteria: <i>P. aeruginosa</i> Resistant to: aminoglycosides (tobramycin), cephalosporins (cefepime), piperacillin/tazobactam Mechanism of resistance: not reported	 Intervention Levofloxacin replaced with gatifloxacin in 2001 Gatifloxacin replaced with moxifloxacin in 2006 Ciprofloxacin available throughout study period Length of pre-intervention: 15 months prior Length of post-intervention 1: 60 months 	Antibiotic resistance and susceptibility No association between the susceptibility of <i>P. aeruginosa</i> isolates to tobramycin and formulary changes was noted. With cefepime, a significant change in susceptibility was detected after the introduction of gatifloxacin (<i>P</i> =0.0099) and moxifloxacin (<i>P</i> =0.0571). In the case of piperacillin/tazobactam, a positive change in susceptibility over time	ITS Protection against secular changes (low quality) Protection against detection bias (low quality)

	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
2000- December 2008	Inclusion criteria: data from clinical microbiology and pharmacy databases of the Medical University of South Carolina Medical Centre Exclusion criteria: not reported		Length of post-intervention 2: 30 months following	was detected after introduction of moxifloxacin (<i>P</i> =0.0589). In each analysis, the effect of total fluoroquinolone usage was not significant	
Cohen 2011 ITS Setting Tertiary (one hospital) Israel March 2006– August 2010	To describe the implementation of an institution-wide, multiple-step intervention to curtail the epidemic spread of CRKP Participants <i>N</i> =33,570 Age: not reported Male: not reported, female: not reported Inclusion criteria: all patients affected by CRKP Exclusion criteria: not reported	Bacteria: <i>K. pneumoniae</i> Resistant to: carbapenems Mechanism of resistance: not reported	 Intervention Single-room isolation and contact precautions Cohorting of patients and nursing staff, screening of patients in the same room as newly identified carriers of CRKP, and local protocol for continued cohorting of returning patients Weekly active surveillance in the ICU Active surveillance of patients on admission to the emergency department Length of pre-intervention: not reported Length of post-intervention 2: 39 months Length of post-intervention 3: 2 years Length of post-intervention 4: 15 months	Bacterial colonization and infectionThe incidence (total number of cases of in-hospital CRKP acquisition detected by clinical cultures) and weekly point prevalence were reported as the number of cases per 1000 hospital bedsIncidence was found to change significantly after intervention 2 (06/2007) and 3 (10/2008). Prevalence was found to change significantly only in September 2009 (after intervention 4)In the emergency department, the mean rate of compliance with the active surveillance protocol (\pm SD) was 43% \pm 10%	ITS Protection against secular changes (high quality) Protection against detection bias (acceptable to low quality)
Dortch 2011	To examine the effect of the	Bacteria: P.	Intervention	Antibiotic use	ITS

	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
ITS Setting Tertiary (one TICU, one SICU) USA January 2001– December 2008	antibiotic stewardship programme on the incidence of resistant Gram- negative HAIs Participants SICU <i>N</i> =6044, TICU <i>N</i> =14,802 Adults 19–45 years, middle aged 46–64 years, aged 65–79 years Male: 14,277, female: 6569 Inclusion criteria: all patients admitted to the SICU or TICU during the study period who contracted an HAI with microbiological confirmation of at least one Gram- negative pathogen, at least 18 years of age Exclusion criteria: not reported	aeruginosa, Acinetobacter spp. Resistant to: aminoglycosides, carbapenems, cephalosporins (third- and fourth-generation), fluoroquinolones Mechanism of resistance: not reported	 Antibiotic stewardship: April 2002, guidelines for prophylactic antibiotics were devised for select procedures Antibiotic rotation: January 2005, institution-wide initiative for surgical prophylaxis based on the Surgical Care Improvement Project Length of pre-intervention: 15 months Length of post-intervention 1: 11 months Length of post-intervention 2: 16 months 	Both in the SICU and TICU and there was a significant decrease in the utilization of total broad-spectrum antibiotics (BLIC, carbapenems, fluoroquinolones, third- and fourth- generation cephalosporins) targeting Gram-negative pathogens over the observation period (<i>P</i> <0.001) Infection During the 8-year observation period, the proportion of healthcare- associated infections caused by MDR Gram-negative pathogens decreased from 37.4% (2001) to 8.5% (2008), whereas the proportion of healthcare-associated infections caused by pan-sensitive pathogens increased from 34.1% to 53.2%	Protection against secular changes (high quality) Protection against detection bias (acceptable to low quality)
Lewis 2012	To examine the effect of restricting ciprofloxacin use on the resistance of nosocomial Gram-negative bacilli, including <i>P. aeruginosa</i> , to group 2	Bacteria: E. aerogenes, E. cloacae, P. aeruginosa, A. baumannii	Intervention Restriction of ciprofloxacin: ciprofloxacin use was restricted hospital wide in July 2007; after	Antibiotic use Following the restriction of ciprofloxacin, there was a significant decreasing trend (<i>P</i> =0.0027) in its	ITS Protection against secular changes (high
Setting Tertiary (11 ICUs and	carbapenems in a hospital's ICUs and intermediate care units	Resistant to: carbapenems	this restriction, pre-approval by the on-call infectious diseases fellow was required for its use	use, from 87.09 DDD/1000 patient- days in 2004 to 8.04 DDD/1000 patient-days in 2010. Use of the	quality) Protection
immediate care units) USA January 2004–	Participants N: not reported Age: not reported Male: not reported, female: not reported	(imipenem, meropenem, doripenem), cephalosporins (cefepime), piperacillin/tazobactam,	Length of pre-intervention: 42 months Length of post-intervention: 42 months	group 2 carbapenems increased significantly (P =0.0134) from 11.96 DDD/1000 patient-days in 2004 to 28.19 DDD/1000 patient-days in 2010. Overall, there was a hospital- wide decrease of 18.4% (P <0.0001)	against detection bias (acceptable quality)
December 2010	Inclusion criteria: all clinical ICU and intermediate care unit specimens	fluoroquinolones (ciprofloxacin)		in the use of antibacterials during the study time	

	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
	(blood, sterile fluid, sputum, urine, wounds and anaerobic specimens) with test results that were positive for <i>P. aeruginosa, E. aerogenes, E. cloacae, A. baumannii</i> and <i>S. maltophilia</i> . Only nosocomial cases, defined as involving patients who had a hospital length of stay exceeding two days Exclusion criteria: results of surveillance and environmental sample cultures.	Mechanism of resistance: not reported		Infection There were no changes observed in the number of nosocomial <i>S.</i> <i>maltophilia</i> isolates per 10,000 patient-days following the restriction of ciprofloxacin Antibiotic resistance Over the seven-year time period, there was a decrease of 13.7% in the percentage of ciprofloxacin- resistant <i>P. aeruginosa</i> isolates that were collected, which equates to a decrease of 3.9% per year (<i>P</i> =0.0017). No significant changes was observed in the susceptibilities to the group II carbapenems of nosocomial Enterobacteriaceae or <i>A.</i> <i>baumannii</i> isolates	
Meyer 2009 ITS Setting Tertiary (one ICU) Germany January 2002– December 2006	To test whether reduction of third- generation cephalosporin use has a sustainable positive impact on the high endemic prevalence of third generation cephalosporin-resistant <i>K. pneumoniae</i> and <i>E. coli</i> in an ICU Participants <i>N</i> =3758 Age: not reported Male: not reported, female: not reported Inclusion criteria: not reported	Bacteria: E. coli, K. pneumoniae, P. aeruginosa Resistant to: cephalosporins (third- generation), piperacillin Mechanism of resistance: ESBL	 Intervention Education programmes for professionals and/or patients in July 2004 Education sessions on antibiotic guidelines were held in the departments of surgery and anaesthesiology Empiric standard therapy for peritonitis and other intra- abominal infections was switched from third- generation cephalosporins to piperacillin in combination with a beta-lactamase 	Antibiotic use Following the implementation of guidelines in a surgical ICU, a significant and sustainable decrease in the use of third-generation cephalosporins of -110.2 DDD/1000 patient-days (95% CI -140.0 to -80.4, R ² =0.468) was observed. There was a significant reduction in the use of ampicillins (-167.4 DDD/1000, 95% CI -223.8 to -110.9, R ² =0.378) and in the use of imidazoles (-94.5 DDD/1000, 95% CI -121.2 to -67.7, R ² =0.463)	ITS Protection against secular changes (high quality) Protection against detection bias (high quality)

	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
	Exclusion criteria: not reported		 inhibitor. The duration of antibiotic therapy for open fractures was shortened to single-shot pre-operative prophylaxis Length of pre-intervention: 30 months Length of post-intervention: 30 months 	The use of aminoglycosides decreased steadily before and after the intervention (slope -1.4 DDD/1000 patient-days per month, 95% CI -1.8 to -1.0, R ² =0.430); piperacillin and piperacillin/tazobactam showed a significant increase in level of 64.4 DDD/1000 patient-days (95% CI 38.5–90.3) and continued to increase by 2.3 DDD/1000 patient- days (95% CI 1.0–3.6) per month after the intervention (R ² =0.745)	
Meyer 2010 ITS Setting Tertiary (one ICU) Germany January 2002– December 2006	To evaluate the impact of a reduced duration of antibiotic prophylaxis for cerebrospinal shunts on total antibiotic use in the ICU and key resistant pathogens Participants <i>N</i> =11,887 Age: not reported Male: not reported, female: not reported Inclusion criteria: monthly data on antimicrobial use obtained from the computerized pharmacy database. Monthly resistance data collected from the microbiology laboratory. Only samples taken in the ICU were considered Exclusion criteria: copy strains – defined as an isolate of the same	Bacteria: E. coli, K. pneumoniae, P. aeruginosa Resistant to: carbapenems (imipenem), cephalosporins (third- generation) Mechanism of resistance: not reported	InterventionChange in antibiotic prophylaxis:Revised recommendation ofsingle-shot prophylaxis withcefuroxime for shunt catheters,beginning in January 2004Length of pre-intervention: 24months priorLength of post-intervention:36 months following	Antibiotic use Following the implementation of a comprehensive teaching session on antibiotic prophylaxis in cerebrospinal shunts in a surgical ICU, pre-operative prophylaxis for shunt catheters was changed into single-shot prophylaxis, and total antibiotic use decreased (–147.3 DDD/1000 patient-days, <i>P</i> =0.052). This corresponded to a decrease of 15% in the use of cefuroxime. The reduction in total antibiotic consumption was sustainable and did not increase over the next 36 months.	ITS Protection against secular changes (high quality) Protection against detection bias (acceptable quality)

Objective and par	rticipants MDR Gram-negative bacteria	ve Intervention, control and follow-up	Results	Quality assessment
species showing th susceptibility patte month period in the matter what the sit	rn throughout a 1- e same patient, no			
Yong 2010To perform an eval in antibiotic suscept common Gram-neg isolated from an IC whether an observ broad-spectrum ar the resistance path bacteriaSetting Tertiary (one ICU) AustraliaParticipants N=13,295 Age: not reported male: not reportedJanuary 2000– December 	ActinetobacterSpp.,ActinetobacterSpp.,BetterEnterobacterSubstrateEnterobacterSubstrateActinetobacterSubstrateActinetobacterSubstrateActinetobacterSubstrateResistant to:ActinetobacterSpp.Resistant to:aminoglycosides,Carbapenems(imipenem),Cephalosporins(ceftazidime),SubstrateSubstrateActinetobacterSpp.Resistant to:ActinetobacterActinetobacterSpp.Resistant to:ActinetobacterActinetobacterSpp.Resistant to:ActinetobacterActinetobacterSpp.Resistant to:ActinetobacterActinetobacterSpp.Resistant to:ActinetobacterActinetobacterSpp.Resistant to:ActinetobacterActinetobacterSpp.Spp.Spp.Resistant to:ActinetobacterActinetobacterSpp.Spp.Spp.Resistance:NetobacterResistance:NetobacterResistance:NetobacterResistance:NetobacterResistance:NetobacterResistance:NetobacterResistance:NetobacterResistance:ResistanceResistance:ResistanceResistance:ResistanceResistance:ResistanceResistance:ResistanceResistance:<	P. Intervention National guidelines on antimicrobial prescribing; antibiotic stewardship via computerized decision suppor systems. In 2001, one system guiding antibiotic use outside ICU – a web-based antimicrol approval system for third- generation cephalosporins (cefotaxime and ceftriaxone). 2002, targeting the ICU specifically – computerized decision support system for antibiotic prescribing Length of pre-intervention: months Length of post-intervention 54 months	reduction in the number of imipenem-resistant <i>E. coli</i> and <i>Klebsiella</i> spp. isolates observed in the ICU. A small but significant improvement in the number of imipenem-resistant <i>Acinetobacter</i> spp. isolates was also observed. For Enterobacteriaceae with potentially inducible beta- lactamases, no significant changes was observed in imipenem susceptibility, although gentamicin	ITS Protection against secular changes (high quality) Protection against detection bias (acceptable to low quality)

	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
				aminoglycosides and fluoroquinolones remained stable during the study period	
Xue 2009	To determine the relation of carbapenem restriction with the	Bacteria: A. baumanniii	Intervention Carbapenem restriction policy	Mortality Mortality rates did not differ	RCT Low
RCT	incidence of MDR <i>A. baumannii</i> in VAP	Resistant to: carbapenems	limiting the use of third- generation carbapenems. Only	significantly between the treatment groups (RR 0.78; 95% CI 0.29–	methodological quality (0)
Setting			used when severe sepsis and	2.12).	
Tertiary (one	Participants	Mechanism of	after consultation with a		Small sample
ICU)	<i>N</i> =26	resistance: ESBL	physician from the Department	Antibiotic resistance	size
China	Adults 19–45 years, middle aged 46–64 years, aged 65–79 years		of Infectious Diseases. N=12	More patients in the conventional group developed a carbapenem-	
June 2007–			Control group	resistant strain of A. baumannii,	
December 2007	Male: 15, female: 11		Conventional treatment: no restrictions of carbapenem	although the difference was not statistically significant (RR 0.63; 95%	
	Inclusion criteria: Patients receiving		(doctors were able to prescribe if	CI 0.38–1.04)	
	mechanical ventilation for more than		necessary). N=15	,	
	five days and diagnosed with VAP				
			Length of follow-up: duration of		
	Exclusion criteria: not reported		treatment		

K. pneumoniae, Klebsiella pneumonia; P.aeruginosa, Pseudomonas aeruginosa; A. baumanniii, Acinetobacter baumanniii; E. coli, Escherichia coli; E. aerogenes; Enterobacter aerogenes; E. cloacae, Enterobacter cloacae; S. maltophilia, Stenotrophomonas maltophilia; CRKP, carbapenem-resistant K. pneumoniae; SICU, surgical intensive care unit; TICU, trauma intensive care unit; VAP, ventilator-associated pneumonia; MDR, multi-drug resistant; ESBL, extended-spectrum beta-lactamase; BLIC, beta-lactam/beta-lactamase inhibitor combinations; ITS, interrupted time series; RCT, randomized controlled trial; ICU, intensive care unit; FQ, fluoroquinolones; 3/4CEPH, third- and fourth-generation cephalosporins; HAI, healthcare-associated infection; CI, confidence interval; RR, risk ratio; DDD, defined daily dose; SD, standard deviation.

Other infection control measures

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
Levin 2010	To analyse whether single patient	Bacteria: Acinetobacter	Intervention	Infection control	СВА
	rooms in the ICU decreased	spp., other Gram-	ICU A converted to single patient	The single-room ICU A had a	Low
CBA	bacterial transmission between ICU	negative bacteria	rooms. Old ICU A <i>N</i> =64, new	significantly lower ICU acquisition of	methodological
	patients		ICU A <i>N</i> =62	resistant organisms when compared	quality (0)
Setting		Resistant to:		with ICU B during the same period	
Tertiary (two	Participants	carbapenems	Control group	[3/62 (5%) vs 7/39 (18%),	
ICUs)	N=207		ICU B remained open plan. Old	respectively, <i>P</i> =0.043], which was	
Israel	Age: not reported	Mechanism of	ICU B <i>N</i> =44, new ICU B <i>N</i> =39	confirmed using survival analysis	
	Male: not reported, female: not	resistance: ESBL		(<i>P</i> =0.011). ICU B showed no	
Dates not	reported		Length of follow-up: not	changes over the study	
reported			reported		
	Inclusion criteria: not reported				
	Exclusion criteria: not reported				

ICU, intensive care unit; ESBL, extended-spectrum beta-lactamase; CBA, controlled before-after study.

Selective decontamination

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
Agusti 2002	To determine the efficacy of SDD in	Bacteria: A. baumannii	Intervention	Bacterial colonization	Quasi-
_	patients with multi-drug-resistant A.		SDD: a combination of	Rates of faecal, pharyngeal and	randomized
Quasi-	baumannii intestinal colonization	Resistant to:	polymyxin E (colistin) (150 mg)	axillary colonization did not	Low
randomized		aminoglycosides	and tobramycine (80 mg)	significantly reduce during ICU stay	methodological
	Participants	(tobramycine)	administered in 20-mL liquid	in the control group (P value not	quality (0)

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
Setting Tertiary (one ICU) Spain October 1998–June 1999	 <i>N</i>=54 Adults 19–45 years, middle aged 46–64 years, aged 65–79 years Male: 16, female: 5 Inclusion criteria: Intervention group 1. All patients with <i>A. baumannii</i> fecal colonization 2. An expected ICU stay exceeding five days Control group 1. All patients admitted 1 October–30 Novembe 1998 with <i>A. baumannii</i> faecal colonization 2. At least one series of axillary-pharyngeal-rectal swab performed Exclusion criteria: not reported 	Mechanism of resistance: not reported	form x 4/day (orally or through nasogastric tube), and 0.5 g of gel containing 2% of colistin and tobramycine applied round the gum margins and oropharynx x 4/day. Duration of treatment from detection of <i>A. baumannii</i> to discharge from ICU. <i>N</i> =21 Control group No intervention. <i>N</i> =33 Length of follow-up : duration of treatment	reported). In the SDD group, the rate of faecal and pharyngeal carriage was reduced significantly (<i>P</i> <0.001 and <i>P</i> =0.003, respectively), but not the rate of cutaneous carriage Antibiotic resistance MDR <i>A. baumannii</i> had not been detected at the time of faecal carriage in 21 of 33 (63.6%) of the control group and 11 of 21 (52.3%) of the SDD group. In the SDD group, all <i>A. baumannii</i> strains were tobramycin resistant and susceptible to colistin at the beginning of the study. No resistance to colistin developed during the study	Small sample size
Brun- Buisson 1989 Quasi- randomized	To study the efficacy of intestinal decontamination by oral non- absorbable antibiotic agents to control a nosocomial outbreak of intestinal colonization and infection with MDR Enterobacteriaceae, and to examine its effects on endemic	Bacteria: Enterobacter spp., P. aeruginosa Resistant to: aminoglycosides (amikacin), third- generation conholosporing	Intervention SDD: a combination of polymyxin E (colistin), 50 mg; neomycin, 1 g; and nalidixic acid (quinolone), 1 g administered in liquid form x 4/day either orally or through a nasogastric tube, starting within 24 h of admission	Mortality All-cause mortality and mortality from nosocomial infections did not differ significantly between patients receiving SDD or no prophylaxis Clinical success/improvement	Quasi- randomized Low methodological quality (0)
Setting Tertiary (one ICU) France January 1987-May 1987	nosocomial infection rates. Participants <i>N</i> =86 Adults 19–45 years, middle aged 46–64 years, aged 65–79 years Male: not reported, female: not reported	cephalosporins Mechanism of resistance: ESBL	starting within 24 h of admission and continuing until discharge from the unit. <i>N</i> =36 Control group No prophylaxis. <i>N</i> =50 Length of follow-up : not	 There was no significant difference between patients receiving SDD or no prophylaxis in: the incidence of any nosocomial infection the infections caused by Gram- negative bacteria 	

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
	Inclusion criteria: 1. Consecutive patients with unit stay exceeding two days 2. Severity score at admission >2 Exclusion criteria: 1. Severe neutropenia routinely receiving oral antibiotic prophylaxis		reported	 the number of nosocomial infections that needed antibiotic treatment There was no significant difference in the number of patients staying on ICU longer than seven or 15 days Bacterial colonization One SDD patient and 12 no prophylaxis patients were positive for MDR strains (RR 0.12; 95% CI 0.02– 0.85). No new cases of MDR strains of Enterobacteriacae were detected during the first four months after the trial Adverse events Three no prophylaxis patients needed therapy for a septic episode caused by Enterobacteriacae; however, this was not significantly different from the intervention group 	
Saidel-Odes 2012 RCT	To assess the effectiveness of SDD for eradicating CRKP oropharyngeal and gastrointestinal carriage Participants	Bacteria: <i>K.</i> pneumoniae Resistant to: carbapenems	Intervention SDD: topical application in the oropharynx of colistin sulfomethate sodium 100,000 U per g and gentamicin sulfate 1.6	Mortality The rate of mortality did not differ significantly between the SDD group and the placebo group. The causes of mortality were not reported. No	RCT High methodological quality (++)
Setting Tertiary (one internal medicine ward) Israel	N=40 Middle aged 46–64 years, aged 65– 79 years, elderly 80+ years Male: 26, female: 14 Inclusion criteria: 1. Hospitalized patients with CRKP	Mechanism of resistance: not reported	mg per g incorporated into the gel. Dose of 0.5 g x 4/day for seven days. Plus an oral solution of 80 mg of gentamicin and 1x10 U of polymyxin E (colistin), given orally or through a nasogastric tube X 4/day for seven days.	adverse events were reported Antibiotic susceptibility CRKP isolates from patients in the SDD arm remained susceptible to gentamicin and polymyxin E throughout the study (MIC ≤2 mg/mL	Small sample size
November	colonization with or without infection		N=20	and ≤0.094 mg/mL, respectively)	

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
2008–June 2010	 2. >18 years of age 3. Available for a follow-up period (while hospitalized or as outpatients) of at least seven weeks Exclusion criteria: <18 years of age, pregnancy, lactation, a known allergy to one of the study drugs, renal failure with creatinine clearance less than 50 mL/min, treatment with intravenous gentamicin and/or intravenous, polymyxin E at the time of randomization 		Control group Placebo: topical application in the oropharynx of the placebo gel, which was compounded from carboxymethyl cellulose. Dose of 0.5 g x 4/day for seven days. Plus two oral solutions, one containing sodium chloride 0.45% and the other containing pulverized sacarin, given orally or through a nasogastric tube X 4/day for seven days. <i>N</i> =20 Length of follow-up: six weeks	Bacterial colonization At the end of treatment, the number of participants in the SDD group that had a throat culture that was CRKP positive reduced from 30% to 0%, whereas in the placebo group, this reduced from 35% to 30% (<i>P</i> <0.0001)	

A. baumannii, Acinetobacter baumannii; K. pneumoniae, Klebsiella pneumoniae; MDR, multi-drug resistant; SDD, selective digestive decontamination; RR, risk ratio, CI, confidence interval; CRKP, carbapenem-resistant K. pneumonia; MIC, minimum inhibitory concentration; RCT, randomized controlled trial; ICU, intensive care unit.

Systematic reviews

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
Falagas 2009 ¹ Setting International Search up to January 2009	To assess the clinical and microbiological effectiveness of fosfomycin in the treatment of MDR, XDR or PDR non-fermenting Gram- negative bacterial infections Participants <i>N</i> =33 Studies: 23 microbiological, one animal and three cohort studies and three case reports Inclusion criteria: microbiological, animal experimental or clinical data on the effect of fosfomycin against MDR non-fermenting Gram-negative pathogens such as <i>Pseudomonas</i> spp., <i>Acinetobacter</i> spp., <i>Stenotrophomonas</i> spp. and <i>Burkholderia</i> spp. MDR, XDR or PDR non-fermenting Gram-negative bacilli or to Gram-negative bacilli with resistance to two or more classes of potentially effective antimicrobial agents Exclusion criteria: studies written in languages other than English, French, German, Italian or Spanish.	Bacteria: Pseudomonas spp., Acinetobacter spp., Stenotrophomonas spp. and Burkholderia spp. See Table II in the paper for details of clinical studies	Intervention Fosfomycin Control group Combination of fosfomycin with other antimicrobial agents	 Microbiological: a total of 1859 MDR non-fermenting Gram-negative isolates. Susceptibility rate to fosfomycin of MDR <i>P. aeruginosa</i> isolates was ≥90% and 50–90% in 7/19 and 4/19 relevant studies, respectively. 30.2% isolates of MDR <i>P. aeruginosa</i>, 3.5% MDR <i>A.</i> <i>baumannii</i> isolates were found to be susceptible to fosfomycin Clinical: 91% of the patients clinically improved (treatment of infections caused by MDR <i>P. aeruginosa</i>) 	Low methodological quality (0) This review was included because it is on the topic; however, the conclusions reached are not supported by the study design

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
	Studies representing abstracts in scientific conferences				
Falagas 2009 ² Setting Not reported Searches performed: 9 July 2008, 16 July 2008 and 11 September 2008	To evaluate the available clinical evidence regarding the effectiveness and safety of systemic colistin in children without cystic fibrosis Participants <i>N</i> =370 Studies: 10 case series and 15 case reports Inclusion criteria: studies with data regarding the use of intravenous, intrathecal, intramuscular or intraventricular colistin in paediatric patients for the treatment of infections caused by colistin- susceptible pathogens or for prophylaxis. All or the majority of patients involved in each individual study should not have cystic fibrosis Exclusion criteria: studies that focused on colistin use in paediatric patients with cystic fibrosis, or reporting the use of oral colistin or the use of colistin for topical treatment in paediatric patients. Abstracts in scientific conferences or studies published in languages other than English, Spanish, French, German, Italian or Greek	Bacteria: P. aeruginosa, A. baumannii, K. aerogenes, H. influenza, P. pyocyanin, P. aeruginosa, K. pneumoniae and A. aerogenes See Table I in the paper for details of studies	Intervention Colistin for the treatment of infections (<i>N</i> =326) Control group Colistin for surgical prophylaxis or prophylaxis of infections in burns patients (<i>N</i> =44)	Case series treatment:271 evaluable subjectsCure: 235/271Improvement: 10/271Deterioration: 6/271Death: 20/271Adverse effects (included in safety assessment N=311)1. Nephrotoxicity: 33/311 had cylindruria or haematuria, 8/311 had a blood urea nitrogen elevation of >10% (in one child owing to an overdosage of colistin), 5/311 had renal tubular cells in the urine, 3/311 had proteinuria and 2/311 had a significant increase in serum creatinine levels during intravenous colistin treatment. Data regarding adverse events not provided for two children2. Neurotoxicity: 0/3113. Other: 8/311Case series prophylaxis: Incidence of infection: 0/44 Death: 9/44 attributed to the underlying pathologies. No signs of colistin-related toxicity were found Adverse effects: 1. Tubular epithelial cells in urine, persistent for up	Acceptable methodological quality (+) This review was included because it is on the topic; however, the conclusions reached are not supported by the study design

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
				colistin: 16/44 2. Proteinuria, disappearing right after colistin withdrawal: 14/44 3. Oliguria during the initial stages of colistin treatment: 1/44 4. No adverse events: 13/44	
Falagas 2010 ³ Setting International Searches up to January 2009	To the evidence on fosfomycin as a treatment option for infections caused by members of the family Enterobacteriaceae with advanced resistance to antimicrobial drugs, including producers of ESBL Participants <i>N</i> =119 Studies: 17 in-vitro microbiological studies, two prospective studies, one retrospective study and two case reports Inclusion criteria: studies on Enterobacteriaceae isolates with an advanced drug resistance (MDR, carbapenem resistance, or production of ESBLs, AmpC β-lactamases, serine carbapenemases or metallo-β-lactamases) profile and their susceptibility to fosfomycin, and the clinical effectiveness of treatment with fosfomycin for infections with these pathogens Exclusion criteria: abstracts in	Bacteria: Microbiological studies K. pneumoniae isolates, E. coli Clinical studies E. coli, S. typhimurium, S. typhi See Table III in the paper for details of studies	Intervention Amoxicillin-clavulanate potassium Control group Fosfomycin–trometamol in two of the <i>E. coli</i> studies	Microbiological success11 of the 17 studies reported that at least 90% of the isolates were susceptible to fosfomycinClinical efficacyMeasured in four studies.Two studies oral treatment for lower UTI with ESBL-producing <i>E. coli</i> (one prospective) resulted in the treatment group with clinical cure in 75 of the 80 (93.8%) patients included in these studies.Two case reports of infection due to MDR Salmonella spp. Reported treatment was effective with fosfomycin	Low methodological quality (0) This review was included because it is on the topic; however, the conclusions reached are not supported by the study design

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
	scientific conferences or studies published in languages other than English, Spanish, French, German, Italian or Greek				
Falagas 2012 ⁴ Setting Not reported Searches from 2000 to 2010	To identify and evaluate the available data regarding the susceptibility of recent Gram- negative bacteria to isepamicin, including that of MDR strains of bacteria Participants <i>N</i> =512 Studies=11 microbiological, one RCT, one prospective study, one restrospective study Inclusion criteria: either a microbiological (in-vitro) study that evaluated the susceptibility of Gram- negative bacterial isolates (including MDR ones) to isepamicin or a clinical study that evaluated the use of isepamicin, given for the treatment of infections by the aforementioned pathogens or for prophylaxis for this type of infection. In addition, studies deemed relevant should have been published between 2000 and 2010 Exclusion criteria: studies that examined a sample of fewer than 10 isolates or patients, studies referring	Bacteria: Clinical studies S. epidermidis, E. coli, S. pneumoniae, P. aeruginosa See Table II in the paper for details of studies	Intervention Isepamicin Control group Two clinical studies – amikacin one clinical study – isepamicin + levofloxacin for prophylaxis	Microbiological: isepamicin was more effective in four studies than amikacin, six studies reported as effective, one study both groups ineffective. In studies including MDR bacteria, 2/4 reported more effective than amikacin; 1/4 as effective as amikacin; 1/4 both isepamicin and amikacin ineffective Clinical: 1. Paediatric infection treatment studies: 100% clinical and bacteriological response for both the isepamicin and the amikacin arms. Definition of clinical response not stated (e.g. cure, improvement) 2. Prophylactic study: acute bacterial prostatitis 1.3%	Low methodological quality (0) This review was included because it is on the topic; however, the conclusions reached are not supported by the study design

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
	to synergistic or pharmacodynamic/ pharmacokinetic parameters of isepamicin, studies that provided data regarding the susceptibility of isepamicin to micro-organisms other than Gram-negative bacteria or the susceptibility of other aminoglycosides only to Gram- negative bacteria. Abstracts in scientific conferences or studies published in languages other than English, Spanish, French, German or Italian				
Kaki 2011 ⁵ Setting International Search January 1996 to	To evaluate the current state of evidence for antimicrobial stewardship interventions in the critical care unit Participants <i>N</i> =not available/not reported for all included studies	Bacteria: P. aeruginosa, A. baumannii, E. coli, Klebsiella spp., ESBL See Table I in the paper for details of studies.	Intervention Antimicrobial stewardship: 1. Antibiotic restriction/ pre- approval 2. Computer-assisted decision support 3. Infectious diseases consultant 4. Re-assessment on pre-	Overall stewardship intervention: 1. Reductions in antimicrobial utilization (11–38% defined daily dose/1000 patient-days) 2. Lower total antimicrobial costs (US\$ 5–10/ patient-day) 3. Shorter average duration of	High methodological quality (++)
December 2010	Studies: three RCTs, three ITSs, and 18 uncontrolled before–after studies Inclusion criteria: application of any intervention; to improve antimicrobial utilization; and within an intensive		 specified date 5. Antibiotic de-escalation protocols 6. Antibiotic prophylaxis guideline 7. Antibiotic treatment guideline 	 antibiotic therapy 4. Less inappropriate use 5. Fewer antibiotic adverse events. stewardship intervention beyond six months: 1. Reductions in antimicrobial resistance rates 	
	care setting Exclusion criteria: if no intervention was applied, non-human or non- patient based, non-hospital based, or they did not involve intensive care patients. Additionally, antibiotic		Control group Not reported, presumably no stewardship	Antibiotic stewardship was not associated with increases in nosocomial infection rates, length of stay or mortality	

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
	cycling. Conference abstracts				
Siempos 2007 ⁶ Setting Not reported	To clarify whether carbapenems are more effective and/or safer than other broad-spectrum antibiotics for the empirical treatment of patients with HAP	Bacteria: <i>P. aeruginosa</i> See Table I in the paper for details of studies	Intervention Carbapenems: 1. Imipenem/ cilastatin (eight studies) 2. Meropenem (four studies)	1. All-cause mortality: lower mortality in the carbapenems group (OR 0.72, 95% CI 0.55–0.95) 2. Treatment success (clinical): no difference between groups (OR 1.08, 95% CI 0.91–1.29)	High methodological quality (++)
Search January 1950 to March 2006	Participants <i>N</i> =2731 Studies: 12 RCTs Inclusion criteria: randomized controlled clinical trial; studied the role of carbapenems in comparison with other broad- spectrum antibiotics or a combination of antibiotics for the empirical treatment of patients with HAP; assessed the effectiveness, toxicity and/or mortality of both therapeutic regimens. Included both patients with HAP and patients with community-acquired pneumonia; however, only data regarding patients with HAP were extracted. Trials with both blind and unblind design were included, and only RCTs written in English, French and German Exclusion criteria: RCTs conducted primarily in neutropenic patients with solid organ tumours or		Control group Imipenem/ cilastatin compared with: 1. Fluoroquinolones: levofloxacin, ciprofloxacin (three studies) 2. Other beta-lactams: piperacillin/tazobactam, aztreonam, cefepime, ceftazidime (five studies) Meropenem compared with: combination of a cephalosporin (ceftazidime, cefuroxime) with an aminoglycoside (amikacin, gentamicin, tobramycin)	 3. Treatment success (microbiological): no difference between groups (OR 1.04, 95% CI 0.72–1.50) 4. Adverse effects: no difference (0.81, 0.46–1.43) <i>P. aeruginosa</i> pneumonia subgroup: lower treatment success (OR 0.42, 95% CI 0.22–0.82) and lower eradication of <i>Pseudomonas</i> spp. strains (OR 0.50, 95% CI 0.24–0.89) in the carbamenems group. Late onset of HAP subgroup: no difference between groups (OR 1.34, 95% CI 0.91–1.97) 	

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
	haematological malignancies and trials that included fewer than 10 patients with pneumonia who received a carbapenem. Experimental trials and trials focusing on pharmacokinetic and/or pharmacodynamics parameters. Finally, RCTs comparing the effectiveness and safety of two different carbapenems				

P. aeruginosa, Pseudomonas aeruginosa; A. baumannii, Acinetobacter baumannii; K. aerogenes, Klebsiella aerogenes; H. influenza, Haemophilus influenza; P. pyocyanin, Pseudomonas pyocyanin; K. pneumoniae, Klebsiella pneumoniae; A. aerogenes, Aerobacter aerogenes; E. coli; Escherichia coli; S. typhimurium, Salmonella typhimurium; S.typhi, Salmonella typhi; S. pneumoniae, Streptococcus pneumoniae; S. epidermidis, Staphylococcus epidermidis; MDR, multi-drug resistant; XDR, extensively drug resistant; PDR, pan-drug resistant; RCT, randomized controlled trial; ESBL, extended-spectrum beta-lactamase; HAP, hospital-acquired pneumonia; OR, odds ratio; CI, confidence interval.

- Falagas ME, Kastoris AC, Karageorgopoulos DE, Rafailidis PI. Fosfomycin for the treatment of infections caused by multidrug-resistant nonfermenting Gram-negative bacilli: a systematic review of microbiological, animal and clinical studies. *Int J Antimicrob Agents* 2009;34:111–120.
- 2. Falagas ME, Vouloumanou EK, Rafailidis PI. Systemic colistin use in children without cystic fibrosis: a systematic review of the literature. *Int J Antimicrob Agents* 2009;**33**:503.e1–e13.
- 3. Falagas ME, Kastoris AC, Kapaskelis AM, Karageorgopoulos DE. Fosfomycin for the treatment of multidrug-resistant, including extendedspectrum beta-lactamase producing, Enterobacteriaceae infections: a systematic review. *Lancet Infect Dis* 2010;**10**:43–50.
- 4. Falagas ME, Karageorgopoulos DE, Georgantzi GG, Sun C, Wang R, Rafailidis PI. Susceptibility of Gram-negative bacteria to isepamicin: a systematic review. *Expert Rev Anti-Infect Ther* 2012;**10**:207–218.
- 5. Kaki R, Elligsen M, Walker S, Simor A, Palmay L, Daneman N. Impact of antimicrobial stewardship in critical care: a systematic review. *J* Antimicrob Chemother 2011;66:1223–1230.

6. Siempos II, Vardakas KZ, Manta KG, Falagas ME. Carbapenems for the treatment of immunocompetent adult patients with nosocomial pneumonia. *Eur Respir J* 2007;**29**:548–560.

Treatment

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
Betrosian 2007 RCT Setting Tertiary (1 ICU) Greece October 2004– February 2006	To evaluate the clinical efficacy and safety of high-dose regimen ampicillin sulbactam for the treatment of VAP from MDR <i>A.</i> <i>baumannii</i> Participants <i>N</i> =27 Age: not reported Male: 15, female: <i>N</i> =12 Inclusion criteria: all patients mechanically ventilated for more than 72 h with positive tracheal aspirates for <i>A. baumannii</i> Exclusion criteria: episodes of VAP in which <i>A. baumannii</i> was isolated in conjunction with another micro- organism	Bacteria: A. baumannii Resistant to: ampicillin/sulbactam and susceptible exclusively to colistin (polymyxin E) Mechanism of resistance: not reported	 Intervention Ampicillin/sulbactam at a rate 2: 1 every 8 h. 24 g/12 g daily for seven to 10 days. <i>N</i>=13 Control group Ampicillin/sulbactam at a rate 2: 1 every 8 h. 18 g/9 g daily for seven to 10 days. <i>N</i>=14 Length of follow-up: one month 	 Mortality 14-day VAP mortality and 30-day all-cause mortality were not significantly different between treatment groups Clinical success/improvement The number of patients with clinical success and clinical failure was not significantly different between treatment groups Bacterial colonization The two treatment groups showed no difference in the eradication of <i>A. baumannii</i> isolates (bacteriological success), bacteriological failure or superinfection Adverse events There was no difference in the adverse effects experienced by participants 	RCT Low methodological quality (0) Very small sample size
Betrosian 2008	To compare the clinical efficacy and safety of high-dose ampicillin/sulbactam vs colistin as	Bacteria: <i>A. baumannii</i> Resistant to:	Intervention Colistin, intravenous 3 MIU every 8 h for eight to 10 days.	Mortality 14-day VAP mortality and 28-day all- cause mortality were not significantly	RCT Low methodological

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
RCT	monotherapy for the treatment of Acinetobacter spp. VAP	Aminoglycosides, carbapenems,	N=15	different between treatment groups	quality (0)
Setting		cephalosporins,	Control group	Clinical success/improvement	Small sample
Tertiary (2 ICUs)	Participants <i>N</i> =28	fluoroquinolones	Ampicillin/sulbactam, 9 g (at a rate 2:1) every 8 h for eight to 10	The number of patients with clinical success and clinical failure was not	size
Greece	Middle aged 46–64 years, aged 65– 79 years	Mechanism of resistance: not	days, administered as follows: three vials (20 mL each)	significantly different between treatment groups	
Dates not reported	Male: 14, female: 14	reported	containing 3.0 g of ampicillin/sulbactam diluted in	Bacterial colonization	
	Inclusion criteria: ventilated patients for >72 h who developed MDR <i>A.</i> <i>baumannii</i> VAP		200 mL of 5% dextrose provided within 1-h duration infusion. <i>N</i> =13	The two treatment groups showed no difference in the eradication of <i>A.</i> <i>baumannii</i> isolates (bacteriological success) or bacteriological failure	
	Exclusion criteria: cases of VAP with mixed isolated micro-organisms, combination antibiotic therapy,		Length of follow-up: two-week- and one-month mortalities	(persistence of <i>A. baumannii</i> isolates (>104 CFU/mL)	
	allergy to beta-lactamase or penicillin, or previous enrolment in similar studies			Adverse events There was no difference in the adverse effects experienced by participants	
Chastre 2003	To compare the efficacy of eight days vs 15 days of antibiotic	Bacteria: <i>E. coli,</i> <i>Klebsiella</i> spp.,	Intervention Antibiotics for eight days:	Mortality 28-day and 60-day all-cause	RCT High
RCT	treatment of patients with microbiologically proven VAP	Enterobacter spp., P. aeruginosa,	specific antibiotics, doses and schedules are not reported.	mortality and in-hospital mortality did not significantly differ between the	methodological quality (++)
		Acinetobacter spp.,	Antibiotics were selected by the	eight- and 15-day regimes	quanty (11)
Setting	Participants	Proteus spp., Serratia	treating physicians. As per		
Tertiary (51	N=401	spp., C. freundii, M.	protocol, the initial regimen	Clinical success/improvement	
ICUs) France	Middle aged 46–64 years, aged 65– 79 years	morgagnii	should have preferably combined at least an	Risk differences (90% CIs) to develop an unfavourable outcome	
TAILE	Male: 141, female: 46	Resistant to:	aminoglycoside, or a	(defined as death, pulmonary	
May 1999-		ticarcillin, methicillin	fluoroquinolone and a broad-	infection recurrence, or prescription	
June 2002	Inclusion criteria:		spectrum beta-lactam	of a new antibiotic for any reason	
	1. >18 years of age	Mechanism of	antimicribial agent. N=197	provided for ≥48 h) were not	
	2. Clinical suspicion of VAP	resistance: ESBL		significantly different between the	
	3. Positive quantitative cultures of			eight- and 15-day regimes for all	

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
	 distal pulmonary secretion samples 4. Instigation within the 24 h following of appropriate empirical antibiotic therapy directed against the micro-organism/s responsible for the infection Exclusion criteria: Pregnant Enrolled in another trial Little chance of survival Neutropenia Concomitant acquired immunodiffeciency syndrome Immunosuppressants or long- term corticosteroid therapy Concomitant extrapulmonary infection that required prolonged antimicrobial treatment Attending physical declined full- life support. Early-onset pneumonia (within the first five days of mechanical ventilation) and no antimicrobial therapy during the 15 days preceding infection. 		Control group Antibiotics for 15 days: specific antibiotics, doses and schedules are not reported. Antibiotics were selected by the treating physicians. As per protocol, the initial regimen should have preferably combined at least an aminoglycoside or a fluoroquinolone and a broad- spectrum beta-lactam antimicribial agent. <i>N</i> =204 Length of follow-up: three months	 patients (RR 2.6, 90% CI -5.6 to 10.7) and for those patients with non-fermenting Gram-negative bacteria (RR 8.6, 90% CI -5.9 to 23.1) The rate of and time to (Kaplan- Meier method, log-rank test) pulmonary infection considered to be recurrence, relapses or superinfection was not significantly different between treatment regimes. Antibiotic use The number of antibiotic-free days was significantly less for all patients on the eight-day regime, but not for those patients with non-fermenting Gram-negative bacteria. No difference was found in the number of patients continuing to receive antibiotics after the end of the trial treatment regimen, or in the number of patients who received an additional course of antibiotics Antibiotic resistance For patients who developed recurrent pulmonary infections, those who had received the eight-day treatment of antibiotics had significantly less emergence of MDR pathogens compared with those who had received the 15-day treatment (42.1% vs 62.3% of recurrent 	

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
				infections, respectively; P=0.04)	
Cox 1987	To compare the efficacy of norfloxacin vs standard parenteral	Bacteria: <i>E. coli,</i> <i>Klebsiella</i> spp.,	Intervention Norfloxacin 400 mg x2/day,	Clinical success/improvement No significant differences were found	RCT Acceptable
RCT	treatment of non-bacteraemic, hospital-acquired UTI	Enterobacter spp., P. aeruginosa, Serratia	minimum treatment seven days. <i>N</i> =52 (46 evaluable patients)	between norfloxacin and standard parenteral antibiotic treatment in the	methodological quality (+)
Setting Secondary (two	Participants <i>N</i> =104	spp., C. freundii, M. morgagnii	Control group Aminoglycosides alone;	rate of participants that were clinically cured, showed clinical improvement or had treatment failure	
hospitals) USA	Age: not reported Male: not reported, female: not reported	Resistant to: not reported	aminoglycosides and meziocillin/ticarcillin; aminoglycosides and	Superinfection Rates of superinfection and early re-	
March 1985– December 1985	Inclusion criteria: 1. Hospitalized patients	Mechanism of resistance: not reported	cephalosporin; aminoglycosides and vancomycin, cephalosporin, cefotaxime alone, administered	infection also did not differ significantly between the norfloxacin and standard parenteral antibiotic	
1000	 2. >18 years of age 3. Documented UTI caused by an 		in accordance with the manufacturers' guidelines. <i>N</i> =52	treatment groups	
	organism known or presumed susceptible to norfloxacin		(48 evaluable patients) Length of follow-up: seven (SD	Antibiotic resistance No differences in the number of patients experiencing adverse	
	Exclusion criteria: 1. <18 years of age		two) days, optional four to six weeks	events were found between those receiving norfloxacin and those	
	2. Pregnant or not practising an effective means of birth control			receiving standard parenteral antibiotics	
	3. A history of allergic diathesis or an allergy to nalidixic acid, oxolinic acid or norfloxacin				
	4. Functional renal abnormalities or unstable deteriorating renal function				
	 Comatose or high probability of imminent death Serious concurrent infection 				
	 Serious concurrent infection Treated or recently completed treatment 				
	with antibiotics 8. History or visual disturbances, a				

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
	psychiatric disorder or central nervous system disease				
Giamarellou 1990 RCT Setting Tertiary (one ICU) Greece Dates not reported	To evaluate the efficacy of monotherapy with pefloxacin in secondary ICU pulmonary infections in comparison with imipenem Participants <i>N</i> =71 Adults 19–45 years, middle aged 46–64 years, aged 65–79 years, elderly 80+ years Male: 42, female: 29 Inclusion criteria: adult patients presenting serious bacterial infections of the respiratory tract Exclusion criteria: not reported	Bacteria: E. coli, K. pneumoniae, Enterobacter spp. (various Enterobacteriaceae), P. aeruginosa, A. anitratus, P. mira, S. marcescens Resistant to: aminoglycosides (gentamicine, tobramycin, netilmicin, amikacin), aztreonam, carbapenems (imipenem), cephalosporins (cefotaxime, ceftazidime, ceftriaxone), fluoroquinolones (ciprofloxacin) Mechanism of resistance: not reported	Intervention Pefloxacin intravenously 400 mg, every 8 h for 11.5 (SD 5.8) days. <i>N</i> =35 Control group Imipenem intravenously 1 g every 8 h for 12.9 (SD 6.2) days. <i>N</i> =36 Length of follow-up: duration of treatment	MortalityThere were three deaths related to sepsis in the imipenem group and one in the pefloxacin group (although the sepsis was not related to the bronchopneumonia, but to an underlying abdominal infection). All- cause mortality was not reportedClinical success/improvement No differences were found in the number of patients cured, the number with superinfection that was cured, the number showing improvement and the number experiencing treatment failure. Bacterial eradication rates were significantly lower in the imipemem group [55.3% vs 82.9%, respectively (P<0.001)]	RCT Acceptable methodological quality (+)
Huttner	To investigate if intestinal carriage of	Bacteria: Enterobacter	Intervention	Clinical success/improvement	RCT

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
2013	ESBL-E can be eradicated	spp. (ESBL-E)	Colistin sulfate 50 mg (equivalent to 42 mg colistin	The rate of eradication of ESBL-E was significantly different between	High methodological
RCT	Participants N=58	Resistant to: cefotaxime/	base or 1.26 million units 4x/day) and neomycin sulfate (250 mg	treatment regimes during treatment (day 6; RR 0.40; 95% CI 0.23–0.70)	quality (++)
Setting Secondary (all inpatient wards of a single	Adolescents 13–18 years, adults 19–45 years, middle aged 46–64 years, aged 65–79 years, elderly 80+ years Male: 34, female: 24	clavulanic acid, ceftazidime, ceftazidime/clavulanic acid, cefepime, cefepime/clavulanic	equivalent to 178 mg neomycin base 4xday) for 10 days. In the presence of ESBL-E bacteriuria, the patients were also treated with nitrofurantoin	or in the first day after treatment (RR 0.42; 95% CI 0.23–0.76), but did not differ in the end of follow-up Treatment adherence	
hospital) Switzerland	Inclusion criteria: aged ≥18 years; ESLB-E-positive rectal swab	acid Mechanism of	(100 mg 3x/day) for five days. <i>N</i> =27	There was no significant difference between groups in the number of patients that adhered to treatment,	
June 2009– June 2012	Exclusion criteria: patients with active ESLB infection, patients treated with antibiotics active against ESLB-E, pregnancy/breastfeeding, contraindication to the use of study drugs, previous study enrolment and resistance of the colonizing ESLB-E strain to colistin (defined as MIC >2 mg/L	resistance: ESBL	Control group Placebo. <i>N</i> =27 Length of follow-up: 28 (SD seven) days	measured by counting the number of pills on the boxes of study medication Adverse events No statistically significant difference was found between the treatment groups in the number of patients with at least one episode of liquid stool	
Moskowitz	To assess whether biofilm-growing	Bacteria: P. aeruginosa	Intervention	Antibiotic susceptibility	RCT
2011 RCT	bacteria susceptibility testing of <i>P.</i> <i>aeruginosa</i> correlates better with clinical outcomes in chronic cystic	Resistant to: aminoglycosides,	Biofilm testing: bioflim regimens of two antibiotics were selected centrally using a published	Participants were assigned to 12 different regimens. The most common regimens included	Acceptable methodological quality (+)
Setting	fibrosis airway infections, when compared with conventional	fluoroquinolones	algorithm, which calculated for each bacterial morphotype the	meropenem (52%) and ciprofloxacin (49%). Azithromycin-containing	Small sample
Secondary (seven cystic	antibiotic susceptibility testing	Mechanism of resistance: not	biofilm minimum inhibitory quotient of each drug, defined as	regimens were used for only two participants (5%), both in the biofilm	size
fibrosis centres) USA	Participants N=39 Adolescents 13–18 years, adults	reported	achievable serum concentration divided by biofilm MIC. <i>N</i> =20	group. No participant received ceftazidime and tobramycin, a combination commonly used in	
	19–45 years		Control group	cystic fibrosis clinical practice	

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
February 2007– October 2007	Male: 25, female: 14 Inclusion criteria: diagnosis of cystic fibrosis, history of persistent <i>P.</i> <i>aeruginosa</i> airway infection, clinical stability at the time of screening, ≥14 years with at least one prior course of intravenous antibiotics Exclusion criteria: sputum culture negative for <i>P. aeruginosa</i> , sputum culture positive for <i>B. cepacia</i> complex species, hospitalization or treatment for an acute pulmonary exacerbation, treatment with oral or inhaled antipseudomonal antibiotics, or azithromycin or other macrolides, within 14 days prior to screening		Conventional testing: conventional regimens of two antibiotics were selected centrally using a published algorithm, which calculated for each bacterial morphotype the conventional minimum inhibitory quotient of each drug defined as achievable serum concentration divided by conventional MIC. <i>N</i> =19 Length of follow-up: 14 days	Of the agents tested, meropenem was most active against biofilm- grown bacteria, but antibiotic regimens based on biofilm testing did not differ significantly from regimens based on conventional testing in terms of microbiological and clinical responses	
Rattanaump awan 2010	To determine whether nebulized	Bacteria: E. coli (ESBL	Intervention	Mortality	RCT
awan 2010	CMS as adjunctive therapy of Gram- negative VAP was safe and	+ve) and <i>E. coli</i> (ESBL - ve), <i>K. pneumoniae</i>	Systemic antibiotic and nebulized CMS (parenteral)	Rates of mortality due to VAP and all-cause mortality did not differ	Acceptable methodological
RCT	beneficial	(ESBL +ve) and K.	equivalent to 75 mg of colistin	between the groups receiving	quality (+)
•		pneumoniae (ESBL -	base reconstituted in 4 mL of	intervention or control	
Setting	Participants N=100	ve), E. cloacae, P.	NSS every 12 h via a nebulizer for 10 min. Continued until	Clinical augeneratimprovement	
Tertiary (one hospital)	Ne 100 Middle aged 46–64 years, aged 65–	aeruginosa, A. baumannii	systemic antibiotic therapy of	Clinical success/improvement Favourable microbiological outcome	
Thailand	79 years, elderly 80+ years	Saamamii	VAP was ended (decided by	was significantly higher in the	
	Male: 64, female: 36	Resistant to:	physician). <i>N</i> =51	intervention group compared with the	
July 2006–		aminoglycosides,		control group (RR 1.57, 95% CI	
September	Inclusion criteria: hospitalized	carbapenems,	Control group	1.03–2.37),but no significant	
2009	patients, ≥18 years of age, diagnosis	fluoroquinolones	Systemic antibiotic(s) plus NSS	difference was observed on clinical	
	of Gram-negative VAP	Maahaniam of	equivalent to 75 mg of colistin	outcomes	
	Exclusion criteria: not reported	Mechanism of resistance: ESBL	base reconstituted in 4 mL of NSS every 12 h via a nebulizer	The overall incidence of	
		TESISIANCE. EODL	for 10 min. Continued until	complications, bronchospasm and	

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
			systemic antibiotic therapy of VAP was ended. <i>N</i> =49	renal impairment did not differ between the two treatment groups	
			Length of follow-up: 28 days		
Stenderup 1983	To study the use of mecillinam as a prophylactic for travellers' diarrhoea	Bacteria: Enterotoxogeni <i>E. coli</i>	Intervention Mecillincam, 200 g, 1x per day for 25 days. <i>N</i> =38	Antibiotic resistance Only 8% of <i>E. coli</i> strains were resistant to three or more antibiotics	RCT Low methodological
RCT Setting Community Denmark Dates not reported	ParticipantsN=74 touristsAdults 19–45 years, middle aged46–64 years, aged 65–79 years,elderly 80+ yearsMale: not reported, female: notreportedInclusion criteria: Danish touriststravelling to Egypt and the Far EastExclusion criteria: not reported	Resistant to: mecillinam, tetracyline, sulfonamide, streptomycin, chloramphenicol, kanamycin, ampicillin, cephalosporin, carbenicillin Mechanism of resistance: not reported	Control group Placebo. <i>N</i> =36 Length of follow-up: duration of treatment	in the pre-travel samples. Post- travel, after participants had received either mecillinam or placebo, approximately 50% or more of the <i>E.</i> <i>coli</i> was resistant to more than three antibiotics	quality (0)
Tannock 2011	To test the efficacy of probiotic strain <i>E. coli</i> Nissle 1917 in reducing the carriage of MDR <i>E. coli</i>	Bacteria: <i>E. coli</i> Resistant to:	Intervention Probiotic: strain <i>E. coli</i> Nissle 1917, 5x10 ⁹ -5x10 ¹⁰ CFU one	Clinical success/improvement There was no significant difference between the probiotic and placebo	RCT Acceptable methodological
RCT Setting	Participants <i>N</i> =70	fluoroquinolones (norfloxacin)	capsule twice daily for five weeks. <i>N</i> =36	groups in the number of people with faecal and urine samples becoming negative or remaining positive.	quality (+)
Primary (14 long-term care facilities)	Age: not reported Male: not reported, female: not reported	Mechanism of resistance: ESBL	Control group Placebo starch powder capsule. <i>N</i> =33	Antibiotic resistance 103 norfloxacin-resistant <i>E. coli</i> isolates from 20 probiotic patients	
New Zealand	Inclusion criteria: not reported Exclusion criteria: not reported		Length of follow-up: five weeks	were tested for susceptibility. All isolates were resistant to norfloxacin (MIC >256 µg/mL) and ciprofloxacin.	
Dates not reported				The majority of norfloxacin-resistant <i>E. coli</i> isolates were MDR. The	
Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
--	--	--	--	---	--
				combination of MDRs differed among strains. None of the isolates were ESBL producers.	
Wang 2009 RCT Setting Tertiary (one ICU) China March 2006–July 2006	To report the effectiveness of extended-infusion meropenem compared with conventional bolus dosing in the management of HAP due to MDR <i>A. baumannii</i> Participants <i>N</i> =30 Adults 19–45 years, middle aged 46–64 years, aged 65–79 years Male: 19, female: 11 Inclusion criteria: HAP due to MDR <i>A. baumannii</i> Exclusion criteria: not reported	Bacteria: <i>A. baumanniii</i> Resistant to: carbapenems (meropenem) Mechanism of resistance: not reported	Intervention Extended intravenous meropenem infusion: 500 mg every 6 h over a 3-h infusion. <i>N</i> =15 Control group Conventional treatment: intravenous meropenem 1 g. every 8 h over a 1-h infusion. <i>N</i> =15 Length of follow-up: duration of treatment	Clinical success/improvement No significant differences were found between extended-infusion meropenem and conventional bolus dosing in the number of patients with treatment success at days 3, 5 and 7. The rates of relapse also did not significantly differ between the treatment groups Antibiotic resistance No patient developed a meropenem- resistant strain of <i>A. baumannii</i> , and the MIC ₉₀ for meropenem against <i>A. baumannii</i> remained at 2 µg/mL	RCT Acceptable methodological quality (+) Small sample size
Xue 2009 RCT Setting Tertiary (one ICU) China June 2007– December 2007	To determine the relation of carbapenem restriction with the incidence of MDR <i>A. baumannii</i> in VAP Participants <i>N</i> =26 Adults 19–45 years, middle aged 46–64 years, aged 65–79 years Male: 15, female: 11 Inclusion criteria: patients receiving mechanical ventilation for more than five days and diagnosed with VAP	Bacteria: <i>A. baumanniii</i> Resistant to: carbapenems Mechanism of resistance: ESBL	Intervention Carbapenem restriction policy limiting the use of third- generation carbapenems. Only used when severe sepsis and after consultation with a physician from the Department of Infectious Diseases. <i>N</i> =12 Control group Conventional treatment: no restrictions of carbapenem (doctors were able to prescribe if necessary). <i>N</i> =15	 Mortality The rates of mortality did not differ significantly between the treatment groups (RR 0.78; 95% CI 0.29–2.12). Antibiotic resistance More patients in the conventional group developed a carbapenem-resistant strain of <i>A. baumannii</i>, although the difference was not statistically significant (RR 0.63; 95% CI 0.38–1.04)	RCT Low methodological quality (0) Small sample size

Study details	Objective and participants	MDR Gram-negative bacteria	Intervention, control and follow-up	Results	Quality assessment
	Exclusion criteria: not reported		Length of follow-up: duration of treatment		

P. aeruginosa, Pseudomonas aeruginosa; E. coli, Escherichia coli; C. freundii, Citrobacter freundii; M. morgagnii, Morganella morgagnii; A. baumannii, Acinetobacter baumannii; A. anitratus, Acinetobacter anitratus; P. mira, Proteus mira; S.marcescens, Serratia marcescens; B. cepacia, Burkholderia cepacia; MDR, multi-drug resistant; VAP, ventilator-associated pneumonia; ESBL, extended-spectrum beta-lactamase; CMS, colistimethate sodium; RCT, randomized controlled trial; ICU, intensive care unit; UTI, urinary tract infection; HAP, hospital-acquired pneumonia; NSS, nebulized sterile normal saline; CFU, colonyforming unit; SD, standard deviation; RR, risk ratio; CI, confidence interval.

References

Antimicrobial Stewardship

Ben-David D, Maor Y, Keller N, *et al.* Potential role of active surveillance in the control of a hospital-wide outbreak of carbapenem-resistant *Klebsiella pneumoniae* infection. *Infect Control Hosp Epidemiol* 2010;**31**:620-626

Borer A, Eskira S, Nativ R, *et al.* A multifaceted intervention strategy for eradication of a hospital-wide outbreak caused by carbapenem-resistant *Klebsiella pneumoniae* in Southern Israel. *Infect Control Hosp Epidemiol* 2011;**32**:1158-1165.

Church EC, Mauldin PD, Bosso JA. Antibiotic resistance in *Pseudomonas aeruginosa* related to quinolone formulary changes: an interrupted time series analysis. *Infect Control Hosp Epidemiol* 2011;**32**:400-402.

Cohen MJ, Block C, Levin PD, *et al.* Institutional control measures to curtail the epidemic spread of carbapenem-resistant *Klebsiella pneumoniae*: A 4-year perspective. *Infect Control Hosp Epidemiol* 2011;**32**:673-678.

Dortch MJ, Fleming SB, Kauffmann RM, Dossett LA, Talbot TR, May AK. Infection reduction strategies including antibiotic stewardship protocols in surgical and trauma intensive care units are associated with reduced resistant Gram-negative healthcare-associated infections. *Surgical Infections* 2011;**12**:15-25.

Lewis GJ, Fang X, Gooch M, Cook PP. Decreased resistance of *Pseudomonas aeruginosa* with restriction of ciprofloxacin in a large teaching hospital's intensive care and intermediate care units. *Infect Control Hosp Epidemiol* 2012;**33**:368-373.

Meyer E, Lapatschek M, Bechtold A, Schwarzkopf G, Gastmeier P, Schwab F. Impact of restriction of third generation cephalosporins on the burden of third generation cephalosporin resistant *K. pneumoniae* and *E. coli* in an ICU. *Intensive Care Med* 2009;**35**:862-870.

Yong MK, Buising KL, Cheng AC, Thursky KA. Improved susceptibility of Gram-negative bacteria in an intensive care unit following implementation of a computerized antibiotic decision support system. *J Antimicrob Chemother* 2010;**65**:1062-1069.

Xue X-s, Wang B, Deng L-j, Kang Y. [Carbapenem restriction reduce the incidence of multidrugresistant *Acinetobacter baumannii* in ventilator associated pneumonia]. *Zhongguo Wei Zhong Bing Ji Jiu Yi Xue* 2009;**21**:234-236.

Other infection control measures

Levin PD, Golovanevski M, Moses AE, Sprung CL, Benenson S. Use of single patient rooms to decrease acquisition of antibiotic-resistant bacteria in the ICU. *Crit Care* 2010;**14**:S156-S157.

Selective decontamination

Agusti C, Pujol M, Argerich MJ, *et al.* Short-term effect of the application of selective decontamination of the digestive tract on different body site reservoir ICU patients colonized by multi-resistant *Acinetobacter baumannii. J Antimicrob Chemother* 2002;**49**:205-208.

Brun-Buisson C, Legrand P, Rauss A, *et al.* Intestinal decontamination for control of nosocomial multiresistant Gram-negative bacilli. Study of an outbreak in an intensive care unit. *Ann Intern Med* 1989;**110**:873-881.

Saidel-Odes L, Polachek H, Peled N, *et al.* A randomized, double-blind, placebo-controlled trial of selective digestive decontamination using oral gentamicin and oral polymyxin E for eradication of carbapenem-resistant *Klebsiella pneumoniae* carriage. *Infect Control Hosp Epidemiol* 2012;**33**:14-19.

Treatment

Betrosian AP, Frantzeskaki F, Xanthaki A, Georgiadis G. High-dose ampicillin-sulbactam as an alternative treatment of late-onset VAP from multidrug-resistant *Acinetobacter baumannii*. *Scand J Infect Dis* 2007;**39**:38-43.

Betrosian AP, Frantzeskaki F, Xanthaki A, Douzinas EE. Efficacy and safety of high-dose ampicillin/sulbactam vs. colistin as monotherapy for the treatment of multidrug resistant *Acinetobacter baumannii* ventilator-associated pneumonia. *J Infect* 2008;**56**:432-436.

Chastre J, Wolff M, Fagon JY, *et al.* Comparison of 8 vs 15 days of antibiotic therapy for ventilatorassociated pneumonia in adults: a randomized trial. *JAMA* 2003;**290**:2588-2598.

Cox CE, McCabe RE, Grad C. Oral norfloxacin versus parenteral treatment of nosocomial urinary tract infection. *Am J Med* 1987;**82(6B)**:59-64.

Giamarellou H, Mandragos K, Bechrakis P, Pigas K, Bilalis D, Sfikakis P. Pefloxacin versus imipenem in the therapy of nosocomial lung infections of intensive care unit patients. *J Antimicrob Chemother* 1990;**26 Suppl B**:117-127.

Huttner B, Haustein T, Uckay I, *et al.* Decolonization of intestinal carriage of extended spectrum betalactamase producing Enterobacteriaceae with oral colistin and neomycin: a randomized, double-blind, placebo-controlled trial. *J Antimicrob Chemother* 2013;**68**:2375-2382.

Moskowitz SM, Emerson JC, McNamara S, *et al.* Randomized trial of biofilm testing to select antibiotics for cystic fibrosis airway infection. *Pediatr Pulmonol* 2011;**46**:184-192.

Rattanaumpawan P, Lorsutthitham J, Ungprasert P, Angkasekwinai N, Thamlikitkul V. Randomized controlled trial of nebulized colistimethate sodium as adjunctive therapy of ventilator-associated pneumonia caused by Gram-negative bacteria. *J Antimicrob Chemother* 2010;**65**:2645-2649.

Stenderup J, Orskov I, Orskov F. Changes in serotype and resistance pattern of the intestinal *Escherichia coli* flora during travel. Results from a trial of mecillinam as a prophylactic against travellers' diarrhoea. *Scand J Infect Dis* 1983;**15**:367-373.

Tannock GW, Tiong IS, Priest P, *et al.* Testing probiotic strain *Escherichia coli* Nissle 1917 (Mutaflor) for its ability to reduce carriage of multidrug-resistant *E. coli* by elderly residents in long-term care facilities. *J Med Microbiol* 2011;**60**:366-370.

Wang D. Experience with extended-infusion meropenem in the management of ventilator-associated pneumonia due to multidrug-resistant *Acinetobacter baumannii*. *Int J Antimicrob Agents* 2009;**33**:290-291.

Xue X-s, Wang B, Deng L-j, Kang Y. [Carbapenem restriction reduce the incidence of multidrugresistant *Acinetobacter baumannii* in ventilator associated pneumonia]. *Zhongguo Wei Zhong Bing Ji Jiu Yi Xue* 2009;**21**:234-236.

Appendix D. Excluded clinical studies

Study design

Case-control study

Abbo A, Navon-Venezia S, Hammer-Muntz O, Krichali T, Siegman-Igra Y, Carmeli Y. Multidrugresistant *Acinetobacter baumannii. Emerg Infect Dis* 2005;**11**:22–29.

Al Jarousha AMK, El Jadba AHN, Al Afifi AS, El Qouqa IA. Nosocomial multidrug-resistant *Acinetobacter baumannii* in the neonatal intensive care unit in Gaza City, Palestine. *Int J Infect Dis* 2009;**13**:623–628.

Aloush V, Navon-Venezia S, Seigman-Igra Y, Cabili S, Carmeli Y. Multidrug-resistant *Pseudomonas aeruginosa*: risk factors and clinical impact. *Antimicrob Agents Chemother* 2006;**50**:43–48.

Anonymous. The cost of antibiotic resistance: effect of resistance among *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Acinetobacter baumannii*, and *Pseudomonas aeruginosa* on length of hospital stay. *Infect Control Hosp Epidemiol* 2002;**23**:106–108.

Apisarnthanarak A, Kiratisin P, Saifon P, Kitphati R, Dejsirilert S, Mundy LM. Clinical and molecular epidemiology of community-onset, extended-spectrum beta-lactamase-producing *Escherichia coli* infections in Thailand: a case–case–control study. *Am J Infect Control* 2007;**35**:606–612.

Arnoni MV, Berezin EN, Martino MDV. Risk factors for nosocomial bloodstream infection caused by multidrug resistant Gram-negative bacilli in pediatrics. *Braz J Infect Dis* 2007;**11**:267–271.

Arruda EA, Marinho IS, Boulos M, *et al.* Nosocomial infections caused by multiresistant *Pseudomonas aeruginosa. Infect Control Hosp Epidemiol* 1999;**20**:620–623.

Asensio A, Oliver A, Gonzalez-Diego P, *et al.* Outbreak of a multiresistant *Klebsiella pneumoniae* strain in an intensive care unit: antibiotic use as risk factor for colonization and infection. *Clin Infect Dis* 2000;**30**:55–60.

Aslan Gulen T, Guner R, Yilmaz GR, Keske S, Tasyaran MA. Clinical impact and cost analysis of multidrug-resistant nosocomial *Acinetobacter baumannii* bacteraemia: a case–control study. *Clin Microbiol Infect* 2012;**18**:765.

Aydemir H, Celebi G, Piskin N, *et al.* Mortality attributable to carbapenem-resistant nosocomial *Acinetobacter baumannii* infections in a Turkish university hospital. *Jpn J Infect Dis* 2012;**65**:66–71.

Banu A, Sathyanarayana BC, Chattannavar G. Efficacy of fresh Aloe vera gel against multi-drug resistant bacteria in infected leg ulcers. *Australas Med J* 2012;**5**:305–309.

Baran G, Erbay A, Bodur H, *et al.* Risk factors for nosocomial imipenem-resistant *Acinetobacter baumannii* infections. *Int J Infect Dis* 2008;**12**:16–21.

Bermejo J, Lesnaberes P, Arnesi N, *et al.* Risk factors associated with ceftazidime-resistant *Klebsiella pneumoniae* infection. *Enferm Infec Microbiol Clin* 2003;**21**:72–76.

Bisson G, Fishman NO, Patel JB, Edelstein PH, Lautenbach E. Extended-spectrum beta-lactamaseproducing *Escherichia coli* and *Klebsiella* species: risk factors for colonization and impact of antimicrobial formulary interventions on colonization prevalence. *Infect Control Hosp Epidemiol* 2002;**23**:254–260.

Borer A, Saidel-Odes L, Riesenberg K, *et al.* Attributable mortality rate for carbapenem-resistant *Klebsiella pneumoniae* bacteremia. *Infect Control Hosp Epidemiol* 2009;**30**:972–976.

Cao B, Wang H, Sun H, Zhu Y, Chen M. Risk factors and clinical outcomes of nosocomial multi-drug resistant *Pseudomonas aeruginosa* infections. *J Hosp Infect* 2004;**57**:112–118.

Cao B, Wang H, Zhu Y-J, Chen MJ. Risk factors and clinical outcomes of nosocomial infections caused by multidrug resistant *Pseudomonas aeruginosa*. *Chin J Tubercul Respir Dis* 2004;**27**:31–35.

Carmeli Y, Eliopoulos G, Mozaffari E, Samore M. Health and economic outcomes of vancomycinresistant enterococci. *Arch Intern Med* 2002;**162**:2223–2228.

Çelebi S, Hacimustafaoglu M, Yüce N, *et al.* Risk factors and clinical outcomes of infections caused by *Acinetobacter* spp. in children: results of a 5 year study. *J Pediatr Infect* 2010;**4**:15–20.

Centers for Disease Control and Prevention. Carbapenem-resistant *Klebsiella pneumoniae* associated with a long-term-care facility – West Virginia, 2009–2011. *MMWR Morb Mortal Wkly Rep* 2011;**60**:1418–1420.

Chen H, Li H, He L, *et al.* Analysis of hospital-acquired pneumonia caused by carbapenem-resistant *Acinetobacter baumannii. Chin J Infect Chemother* 2010;**10**:94–99.

Cipriano Souza R, Vicente AC, Vieira VV, *et al.* Clindamycin and metronidazole as independent risk factors for nosocomial acquisition of multidrug-resistant *Pseudomonas aeruginosa. J Hosp Infect* 2008;**69**:402–403.

Cohen MJ, Anshelevich O, Raveh D, Broide E, Rudensky B, Yinnon AM. Acquisition of multidrugresistant organisms among hospital patients hospitalized in beds adjacent to critically ill patients. *Infect Control Hosp Epidemiol* 2006;**27**:675–681.

Cohen-Nahum K, Saidel-Odes L, Riesenberg K, Schlaeffer F, Borer A. Urinary tract infections caused by multi-drug resistant *Proteus mirabilis*: risk factors and clinical outcomes. *Infection* 2010;**38**:41–46.

Cornejo-Juarez P, Perez-Jimenez C, Silva-Sanchez J, *et al.* Molecular analysis and risk factors for *Escherichia coli* producing extended-spectrum beta-lactamase bloodstream infection in hematological malignancies. *PLoS One* 2012;**7**:e35780.

Cortes JA, Cuervo SI, Urdaneta AM, *et al.* Identifying and controlling a multiresistant *Pseudomonas aeruginosa* outbreak in a Latin-American cancer centre and its associated risk factors. *Braz J Infect Dis* 2009;**13**:99–103.

Cranendonk DR, van der Valk M, Langenberg ML, van der Meer JT. Clinical consequences of increased ciprofloxacin and gentamicin resistance in patients with *Escherichia coli* bacteraemia in the Netherlands. *Scand J Infect Dis* 2012;**44**:363–368.

Dantas SRPE, Moretti-Branchini ML. Impact of antibiotic-resistant pathogens colonizing the respiratory secretions of patients in an extended-care area of the emergency department. *Infect Control Hosp Epidemiol* 2003;**24**:351–355.

Defez C, Fabbro-Peray P, Bouziges N, *et al.* Risk factors for multidrug-resistant *Pseudomonas aeruginosa* nosocomial infection. *J Hosp Infect* 2004;**57**:209–216.

del Mar Tomas M, Cartelle M, Pertega S, *et al.* Hospital outbreak caused by a carbapenem-resistant strain of *Acinetobacter baumannii*: patient prognosis and risk-factors for colonisation and infection. *Clin Microbiol Infect* 2005;**11**:540–546.

Deris ZZ, Harun A, Shafei MN, Rahman RA, Johari MR. Outcomes and appropriateness of management of nosocomial Acinetobacter bloodstream infections at a teaching hospital in northeastern Malaysia. *Southeast Asian J Trop Med Public Health* 2009;**40**:140–147.

Di Martino P, Gagniere H, Berry H, Bret L. Antibiotic resistance and virulence properties of *Pseudomonas aeruginosa* strains from mechanically ventilated patients with pneumonia in intensive care units: comparison with imipenem-resistant extra-respiratory tract isolates from uninfected patients. *Microbes Infect* 2002;**4**:613–620.

Durakovic N, Radojcic V, Boban A, *et al.* Efficacy and safety of colistin in the treatment of infections caused by multidrug-resistant *Pseudomonas aeruginosa* in patients with hematologic malignancy: a matched pair analysis. *Intern Med* 2011;**50**:1009–1013.

Eagye KJ, Kuti JL, Nicolau DP. Risk factors and outcomes associated with isolation of meropenem high-level-resistant *Pseudomonas aeruginosa*. *Infect Control Hosp Epidemiol* 2009;**30**:746–752.

Falagas ME, Rafailidis PI, Kofteridis D, *et al.* Risk factors of carbapenem-resistant *Klebsiella pneumoniae* infections: a matched case–control study. *J Antimicrob Chemother* 2007;**60**:1124–1130.

Fernandez A, Pereira MJ, Suarez JM, *et al.* Emergence in Spain of a multidrug-resistant *Enterobacter cloacae* clinical isolate producing SFO-1 extended-spectrum beta-lactamase. *J Clin Microbiol* 2011;**49**:822–828.

Fierobe L, Lucet J, Decre D, *et al.* An outbreak of imipenem-resistant *Acinetobacter baumannii* in critically ill surgical patients. *Infect Control Hosp Epidemiol* 2001;**22**:35–40.

Fortaleza CMCB, Freire MP, Filho Dde C, de Carvalho Ramos M. Risk factors for recovery of imipenem- or ceftazidime-resistant *Pseudomonas aeruginosa* among patients admitted to a teaching hospital in Brazil. *Infect Control Hosp Epidemiol* 2006;**27**:901–906.

Furtado GHC, Bergamasco MD, Menezes FG, et al. Imipenem-resistant Pseudomonas aeruginosa

infection at a medical-surgical intensive care unit: risk factors and mortality. *J Crit Care* 2009;**24**:625.e9–e14.

Garnacho J, Sole-Violan J, Sa-Borges M, Diaz E, Rello J. Clinical impact of pneumonia caused by *Acinetobacter baumannii* in intubated patients: a matched cohort study. *Crit Care Med* 2003;**31**:2478–2482.

Gasink LB, Fishman NO, Nachamkin I, Bilker WB, Lautenbach E. Risk factors for and impact of infection or colonization with aztreonam-resistant *Pseudomonas aeruginosa*. *Infect Control Hosp Epidemiol* 2007;**28**:1175–1180.

Gregory CJ, Llata E, Stine N, *et al.* Outbreak of carbapenem-resistant *Klebsiella pneumoniae* in Puerto Rico associated with a novel carbapenemase variant. *Infect Control Hosp Epidemiol* 2010;**31**:476–484.

Gulay Z, Atay T, Amyes SG. Clonal spread of imipenem-resistant *Pseudomonas aeruginosa* in the intensive care unit of a Turkish hospital. *J Chemother* 2001;**13**:546–554.

Hussein K, Sprecher H, Mashiach T, Oren I, Kassis I, Finkelstein R. Carbapenem resistance among *Klebsiella pneumoniae* isolates: risk factors, molecular characteristics, and susceptibility patterns. *Infect Control Hosp Epidemiol* 2009;**30**:666–671.

Hyle EP, Ferraro MJ, Silver M, Lee H, Hooper DC. Ertapenem-resistant Enterobacteriaceae: risk factors for acquisition and outcomes. *Infect Control Hosp Epidemiol* 2010;**31**:1242–1249.

Hyle EP, Lipworth AD, ZaoutisTE, *et al.* Risk factors for increasing multidrug resistance among extended-spectrum beta-lactamase-producing *Escherichia coli* and *Klebsiella* species. *Clin Infect Dis* 2005;**40**:1317–1324.

Jeon M-H, Choi S-H, Kwak YG, *et al.* Risk factors for the acquisition of carbapenem-resistant *Escherichia coli* among hospitalized patients. *Diagn Microbiol Infect Dis* 2008;**62**:402–406.

Johnson JR, Kuskowski MA, Gajewski A, Sahm DF, Karlowsky JA. Virulence characteristics and phylogenetic background of multidrug-resistant and antimicrobial-susceptible clinical isolates of *Escherichia coli* from across the United States, 2000–2001. *J Infect Dis* 2004;**190**:1739–1744.

Kallel H, Hergafi L, Bahloul M, *et al.* Safety and efficacy of colistin compared with imipenem in the treatment of ventilator-associated pneumonia: a matched case–control study. *Intensive Care Med* 2007;**33**:1162–1167.

Kanafani ZA, Mehio-Sibai A, Araj GF, Kanaan M, Kanj SS. Epidemiology and risk factors for extended-spectrum beta-lactamase-producing organisms: a case–control study at a tertiary care center in Lebanon. *Am J Infect Control* 2005;**33**:326–332.

Kang C, Kim S, Park WB, *et al.* Clinical epidemiology of ciprofloxacin resistance and its relationship to broad-spectrum cephalosporin resistance in bloodstream infections caused by *Enterobacter* species. *Infect Control Hosp Epidemiol* 2005;**26**:88–92.

Kim YA, Choi JY, Kim CK, *et al.* Risk factors and outcomes of bloodstream infections with metallobeta-lactamase-producing *Acinetobacter*. *Scand J Infect Dis* 2008;**40**:234–240.

Kim JY, Lautenbach E, Chu J, *et al.* Fluoroquinolone resistance in pediatric bloodstream infections because of *Escherichia coli* and *Klebsiella* species. *Am J Infect Control* 2008;**36**:70–73.

Kohlenberg A, Weitzel-Kage D, Sohr D, *et al.* Outbreak of carbapenem-resistant *Pseudomonas aeruginosa* infection in a surgical intensive care unit. *J Hosp Infect* 2010;**74**:350–357.

Krcmery, Jr, V, Spanik S, Krupova I, *et al.* Bacteremia due to multiresistant Gram-negative bacilli in neutropenic cancer patients: a case controlled study. *J Chemother* 1998;**10**:320–325.

Kritsotakis EI, Tsioutis C, Roumbelaki M, Christidou A, Gikas A. Antibiotic use and the risk of carbapenem-resistant extended-spectrum-{beta}-lactamase-producing *Klebsiella pneumoniae* infection in hospitalized patients: results of a double case–control study. *J Antimicrob Chemother* 2011;**66**:1383–1391.

Kuo KC, Shen YH, Hwang KP. Clinical implications and risk factors of extended-spectrum betalactamase-producing *Klebsiella pneumoniae* infection in children: a case–control retrospective study in a medical center in southern Taiwan. *J Microbiol Immunol Infect* 2007;**40**:248–254.

Kuo SC, Lee YT, Yang SP, *et al.* Eradication of multidrug-resistant *Acinetobacter baumannii* from the respiratory tract with inhaled colistin methanesulfonate: a matched case–control study. *Clin Microbiol Infect* 2012;**18**:870–876.

Lautenbach E, Strom BL, Bilker WB, *et al.* Epidemiological investigation of fluoroquinolone resistance in infections due to extended-spectrum beta-lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae*. *Clin Infect Dis* 2001;**33**:1288–1294.

Lautenbach E, Metlay JP, Weiner MG, *et al.* Gastrointestinal tract colonization with fluoroquinoloneresistant *Escherichia coli* in hospitalized patients: changes over time in risk factors for resistance. *Infect Control Hosp Epidemiol* 2009;**30**:18–24.

Lautenbach E, Synnestvedt M, Weiner MG, *et al.* Epidemiology and impact of imipenem resistance in *Acinetobacter baumannii. Infect Control Hosp Epidemiol* 2009;**30**:1186–1192.

Lautenbach E, Synnestvedt M, Weiner MG, *et al.* Imipenem resistance in *Pseudomonas aeruginosa*: emergence, epidemiology, and impact on clinical and economic outcomes. *Infect Control Hosp Epidemiol* 2010;**31**:47–53.

Lee NY, Lee HC, Ko NY, *et al.* Clinical and economic impact of multidrug resistance in nosocomial *Acinetobacter baumannii* bacteremia. *Infect Control Hosp Epidemiol* 2007;**28**:713–719.

Lin MF, Yang CM, Lin CH, Huang ML, Tu CC, Liou ML. Clinical features and molecular epidemiology of multidrug-resistant *Acinetobacter calcoaceticus–A baumannii* complex in a regional teaching hospital in Taiwan. *Am J Infect Control* 2009;**37**:e1–e3.

Lodise TP, Miller CD, Graves J, *et al.* Clinical prediction tool to identify patients with *Pseudomonas aeruginosa* respiratory tract infections at greatest risk for multidrug resistance. *Antimicrob Agents Chemother* 2007;**51**:417–422.

Lopez-Dupla M, Martinez JA, Vidal F, *et al.* Previous ciprofloxacin exposure is associated with resistance to beta-lactam antibiotics in subsequent *Pseudomonas aeruginosa* bacteremic isolates. *Am J Infect Control* 2009;**37**:753–758.

Lytsy B, Lindback J, Torell E, Sylvan S, Velicko I, Melhus A. A case–control study of risk factors for urinary acquisition of *Klebsiella pneumoniae* producing CTX-M-15 in an outbreak situation in Sweden. *Scand J Infect Dis* 2010;**42**:439–444.

Maragakis LL, Winkler A, Tucker MG, *et al.* Outbreak of multidrug-resistant *Serratia marcescens* infection in a neonatal intensive care unit. *Infect Control Hosp Epidemiol* 2008;**29**:418–423.

Martinez-Aguilar G, Alpuche-Aranda CM, Anaya C, et al. Outbreak of nosocomial sepsis and pneumonia in a newborn intensive care unit by multiresistant extended-spectrum beta-lactamase-producing *Klebsiella pneumoniae*: high impact on mortality. *Infect Control Hosp Epidemiol* 2001;**22**:725–728.

Matthaiou DK, Michalopoulos A, Rafailidis PI, *et al.* Risk factors associated with the isolation of colistin-resistant Gram-negative bacteria: a matched case–control study. *Crit Care Med* 2008;**36**:807–811.

Mentzelopoulos SD, Pratikaki M, Platsouka E, *et al.* Prolonged use of carbapenems and colistin predisposes to ventilator-associated pneumonia by pandrug-resistant *Pseudomonas aeruginosa. Intensive Care Med* 2007;**33**:1524–1532.

Montero M, Dominguez M, Orozco-Levi M, Salvado M, Knobel H. Mortality of COPD patients infected with multi-resistant *Pseudomonas aeruginosa*: a case and control study. *Infection* 2009;**37**:16–19.

Montero M, Sala M, Riu M, et al. Risk factors for multidrug-resistant *Pseudomonas aeruginosa* acquisition. Impact of antibiotic use in a double case–control study. *Eur J Clin Microbiol Infect Dis* 2010;**29**:335–339.

Mosqueda-Gomez JL, Montano-Loza A, Rolon AL, *et al.* Molecular epidemiology and risk factors of bloodstream infections caused by extended-spectrum beta-lactamase-producing *Klebsiella pneumoniae*. A case–control study. *Int J Infect Dis* 2008;**12**:653–659.

Mouloudi E, Protonotariou E, Zagorianou A, *et al.* Bloodstream infections caused by metallo-betalactamase/*Klebsiella pneumoniae* carbapenemase-producing K. pneumoniae among intensive care unit patients in Greece: risk factors for infection and impact of type of resistance on outcomes. *Infect Control Hosp Epidemiol* 2010;**31**:1250–1256.

Muder RR, Brennen C, Drenning SD, Stout JE, Wagener MM. Multiply antibiotic-resistant Gramnegative bacilli in a long-term-care facility: a case–control study of patient risk factors and prior antibiotic use. *Infect Control Hosp Epidemiol* 1997;**18**:809–813. Nateghian AR, Parvin M, Rohani P, Tabrizi M. Incidence and risk factors for gentamicin and ceftriaxone resistant E.coli causing urinary tract infection in children admitted in Hazrat-E-Ali Asghar hospital. *J Iran Univ Med Sci* 2009;**16**:56.

Nouer SA, Nucci M, de-Oliveira MP, Pellegrino FLPC, Moreira BM. Risk factors for acquisition of multidrug-resistant *Pseudomonas aeruginosa* producing SPM metallo-beta-lactamase. *Antimicrob Agents Chemother* 2005;**49**:3663–3667.

Nseir S, Di Pompeo C, Diarra M, *et al.* Relationship between immunosuppression and intensive care unit-acquired multidrug-resistant bacteria: a case–control study. *Crit Care Med* 2007;**35**:1318–1323.

Ohmagari N, Hanna H, Graviss L, *et al.* Risk factors for infections with multidrug-resistant *Pseudomonas aeruginosa* in patients with cancer. *Cancer* 2005;**104**:205–212.

Onguru P, Erbay A, Bodur H, *et al.* Imipenem-resistant *Pseudomonas aeruginosa*: risk factors for nosocomial infections. *J Korean Med Sci* 2008;**23**:982–987.

Palmore TN, Michelin AV, Bordner M, *et al.* Use of adherence monitors as part of a team approach to control clonal spread of multidrug-resistant *Acinetobacter baumannii* in a research hospital. *Infect Control Hosp Epidemiol* 2011;**32**:1166–1172.

Paramythiotou E, Lucet JC, Timsit JM, *et al.* Acquisition of multidrug-resistant *Pseudomonas aeruginosa* in patients in intensive care units: role of antibiotics with antipseudomonal activity. *Clin Infect Dis* 2004;**38**:670–677.

Park YS, Lee H, Chin BS, *et al.* Acquisition of extensive drug-resistant *Pseudomonas aeruginosa* among hospitalized patients: risk factors and resistance mechanisms to carbapenems. *J Hosp Infect* 2011;**79**:54–58.

Park SY, Kang CI, Joo EJ, *et al.* Risk factors for multidrug resistance in nosocomial bacteremia caused by extended-spectrum beta-lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae*. *Microb Drug Resist* 2012;**18**:518–524.

Patel G, Huprikar S, Factor SH, Jenkins SG, Calfee DP. Outcomes of carbapenem-resistant Klebsiella pneumoniae infection and the impact of antimicrobial and adjunctive therapies. *Infect Control Hosp Epidemiol* 2008;**29**:1099–1106

Pereira GH, Levin AS, Oliveira HB, Moretti ML. Controlling the clonal spread of *Pseudomonas* aeruginosa infection. *Infect Control Hosp Epidemiol* 2008;**29**:549–552.

Pinheiro MRS, Lacerda HR, Melo RGL, Maciel MA. *Pseudomonas aeruginosa* infections: factors relating to mortality with emphasis on resistance pattern and antimicrobial treatment. *Braz J Infect Dis* 2008;**12**:509–515.

Playford EG, Craig JC, Iredell JR. Carbapenem-resistant *Acinetobacter baumannii* in intensive care unit patients: risk factors for acquisition, infection and their consequences. *J Hosp Infect* 2007;**65**:204–211

Pop-Vicas A, Tacconelli E, Gravenstein S, Bing L, D'Agata EMC. Influx of multidrug-resistant, Gramnegative bacteria in the hospital setting and the role of elderly patients with bacterial bloodstream infection. *Infect Control Hosp Epidemiol* 2009;**30**:325–331.

Qavi A, Segal-Maurer S, Mariano N, *et al.* Increased mortality associated with a clonal outbreak of ceftazidime-resistant *Klebsiella pneumoniae*: a case–control study. *Infect Control Hosp Epidemiol* 2005;**26**:63–68.

Rattanaumpawan P, Tolomeo P, Bilker WB, Fishman NO, Lautenbach E. Risk factors for fluoroquinolone resistance in Gram-negative bacilli causing healthcare-acquired urinary tract infections. *J Hosp Infect* 2010;**76**:324–327.

Ray A, Perez F, Beltramini AM, *et al.* Use of vaporized hydrogen peroxide decontamination during an outbreak of multidrug-resistant *Acinetobacter baumannii* infection at a long-term acute care hospital. *Infect Control Hosp Epidemiol* 2010;**31**:1236–1241.

Ribeiro Gomes MZ, de Oliveira RVC, Machado CR, *et al.* Factors associated with epidemic multiresistant *Pseudomonas aeruginosa* infections in a hospital with AIDS-predominant admissions. *Braz J Infect Dis* 2012;**16**:219–225.

Rocha Lde A, Vilela CAP, Cezario RC, Almeida AB, Gontijo Filho P. Ventilator-associated pneumonia in an adult clinical-surgical intensive care unit of a Brazilian university hospital: incidence, risk factors, etiology, and antibiotic resistance. *Braz J Infect Dis* 2008;**12**:80–85.

Rodriguez-Bano J, Navarro MD, Romero L, *et al.* Clinical and molecular epidemiology of extendedspectrum beta-lactamase-producing *Escherichia coli* as a cause of nosocomial infection or colonization: implications for control. *Clin Infect Dis* 2006;**42**:37–45.

Romanelli RM, Jesus LA, Clemente WT, *et al.* Outbreak of resistant *Acinetobacter baumannii*measures and proposal for prevention and control. *Braz J Infect Dis* 2009;**13**:341–347.

Saito R, Okugawa S, Kumita W, *et al.* Clinical epidemiology of ciprofloxacin-resistant *Proteus mirabilis* isolated from urine samples of hospitalised patients. *Clin Microbiol Infect* 2007;**13**:1204–1206.

Salgado FXC, Goncalves JC, de Souza CM, da Silva NB, Sanchez TEG, de Oliveira Karnikowski MG. Cost of antimicrobial treatment in patients infected with multidrug-resistant organisms in the intensive care unit. *Medicina* 2011;**71**:531–535.

Sanchez M, Herruzo R, Marban A, *et al.* Risk factors for outbreaks of multidrug-resistant *Klebsiella pneumoniae* in critical burn patients. *J Burn Care Res* 2012;**33**:386–392.

Scerpella EG, Wanger AR, Armitige L, Anderlini P, Ericsson CD. Nosocomial outbreak caused by a multiresistant clone of *Acinetobacter baumannii*: results of the case–control and molecular epidemiologic investigations. *Infect Control Hosp Epidemiol* 1995;**16**:92–97.

Schechner V, Kotlovsky T, Tarabeia J, *et al.* Predictors of rectal carriage of carbapenem-resistant Enterobacteriaceae (CRE) among patients with known CRE carriage at their next hospital encounter. *Infect Control Hosp Epidemiol* 2011;**32**:497–503.

Serefhanoglu K, Turan H, Timurkaynak FE, Arslan H. Bloodstream infections caused by ESBLproducing *E. coli* and *K. pneumoniae*: risk factors for multidrug-resistance. *Braz J Infect Dis* 2009;**13**:403–407.

Soderstrom M, Vikatmaa P, Lepantalo M, Aho PS, Kolho E, Ikonen T. The consequences of an outbreak of multidrug-resistant Pseudomonas aeruginosa among patients treated for critical leg ischemia. *J Vasc Surg* 2009;**50**:806–812.

Song KH, Jeon JH, Park WB, *et al.* Clinical outcomes of spontaneous bacterial peritonitis due to extended-spectrum beta-lactamase-producing *Escherichia coli* and *Klebsiella* species: a retrospective matched case–control study. *BMC Infect Dis* 2009;**9**:41.

Spanik S, Krupova I, Trupl J, *et al.* Bacteremia due to multiresistant Gram-negative bacilli in neutropenic cancer patients: a case–controlled study. *J Infect Chemother* 1999;**5**:180–184.

Sunenshine RH, Wright MO, Maragakis LL, *et al.* Multidrug-resistant *Acinetobacter* infection mortality rate and length of hospitalization. *Emerg Infect Dis* 2007;**13**:97–103.

Superti SV, Augusti G, Zavascki AP. Risk factors for and mortality of extended-spectrum-betalactamase-producing *Klebsiella pneumoniae* and *Escherichia coli* nosocomial bloodstream infections. *Rev Inst Med Trop Sao Paulo* 2009;**51**:211–216.

Surasarang K, Narksawat K, Danchaivijitr S, *et al.* Risk factors for multi-drug resistant *Acinetobacter baumannii* nosocomial infection. *J Med Assoc Thailand* 2007;**90**:1633–1699.

Tacconelli E, Cataldo MA, De Pascale G, *et al.* Prediction models to identify hospitalized patients at risk of being colonized or infected with multidrug-resistant *Acinetobacter baumannii* calcoaceticus complex. *J Antimicrob Chemother* 2008;**62**:1130–1137.

Tsai H-T, Wang J-T, Chen C-J, Chang S-C. Association between antibiotic usage and subsequent colonization or infection of extensive drug-resistant *Acinetobacter baumannii*: a matched case–control study in intensive care units. *Diagn Microbiol Infect Dis* 2008;**62**:298–305.

Tumbarello M, Repetto E, Trecarichi EM, *et al.* Multidrug-resistant *Pseudomonas aeruginosa* bloodstream infections: risk factors and mortality. *Epidemiol Infect* 2011;**139**:1740–1749.

Tumbarello M, Trecarichi EM, Fiori B, *et al.* Multidrug-resistant *Proteus mirabilis* bloodstream infections: risk factors and outcomes. *Antimicrob Agents Chemother* 2012;**56**:3224–3231.

Tuncer Ertem G, Sonmezer MC, Tulek N, *et al.* Evaluation of risk factors for nosocomial multidrugresistant *Pseudomonas aeruginosa* infections. *Clin Microbiol Infect* 2012;**18**:517.

Turkoglu M, Dizbay M, Ciftci A, Aksakal FN, Aygencel G. Colistin therapy in critically ill patients with chronic renal failure and its effect on development of renal dysfunction. *Int J Antimicrob Agents* 2012;**39**:142–145.

Valencia R, Arroyo LA, Conde M, *et al.* Nosocomial outbreak of infection with pan-drug-resistant *Acinetobacter baumannii* in a tertiary care university hospital. *Infect Control Hosp Epidemiol* 2009;**30**:257–263.

Vigil KJ, Adachi JA, Aboufaycal H, *et al.* Multidrug-resistant *Escherichia coli* bacteremia in cancer patients. *Am J Infect Control* 2009;**37**:741–745.

Villers D, Espaze E, Coste-Burel M, *et al.* Nosocomial *Acinetobacter baumannii* infections: microbiological and clinical epidemiology. *Ann Intern Med* 1998;**129**:182–189.

von Dolinger de Brito D, Oliveira EJ, Abdallah VOS, da Costa Darini AL, Filho PPG. An outbreak of *Acinetobacter baumannii* septicemia in a neonatal intensive care unit of a university hospital in Brazil. *Braz J Infect Dis* 2005;**9**:301–309.

Weingarten CM, Rybak MJ, Jahns BE, Stevenson JG, Brown WJ, Levine DP. Evaluation of *Acinetobacter baumannii* infection and colonization, and antimicrobial treatment patterns in an urban teaching hospital. *Pharmacotherapy* 1999;**19**:1080–1085.

Wendt C, Lin D, von Baum H. Risk factors for colonization with third-generation cephalosporinresistant enterobacteriaceae. *Infection* 2005;**33**:327–332.

Yakupogullari Y, Otlu B, Dogukan M, *et al.* Investigation of a nosocomial outbreak by alginateproducing pan-antibiotic-resistant *Pseudomonas aeruginosa. Am J Infect Control* 2008;**36**:e13–e18.

Ye, Jr J, Huang C-T, Shie S-S, *et al.* Multidrug resistant *Acinetobacter baumannii*: risk factors for appearance of imipenem resistant strains on patients formerly with susceptible strains. *PLoS One* 2010;**5**:e9947.

Young LS, Sabel AL, Price CS. Epidemiologic, clinical, and economic evaluation of an outbreak of clonal multidrug-resistant *Acinetobacter baumannii* infection in a surgical intensive care unit. *Infect Control Hosp Epidemiol* 2007;**28**:1247–1254.

Case series/report

Allou N, Kermarrec N, Muller C, *et al.* Risk factors and prognosis of post-operative pneumonia due to *Pseudomonas aeruginosa* following cardiac surgery. *J Antimicrob Chemother* 2010;**65**:806–807.

Bercault N, Linassier P. Interest of septic isolation to decrease the acquisition of multiply antibioticresistant bacteria in intensive care unit. Effect on nosocomial infections. *Rev Med Intern* 1999;**20**:86– 87.

Bint AJ, Bullock DW, Speller DC, Stern SR, Turner A. Cefuroxime therapy for urinary tract infections caused by a multi-resistant, epidemic *Klebsiella aerogenes*. *J Antimicrob Chemother* 1979;**5**:189–193.

Conejo MC, Dominguez MC, Lopez-Cerero L, Serrano L, Rodriguez-Bano J, Pascual A. Isolation of multidrug-resistant *Klebsiella oxytoca* carrying blaIMP-8, associated with OXY hyperproduction, in the intensive care unit of a community hospital in Spain. *J Antimicrob Chemother* 2010;**65**:1071–1073.

Cree M, Stacey S, Graham N, Wainwright C. Fosfomycin – investigation of a possible new route of administration of an old drug. A case study. *J Cystic Fibrosis* 2007;**6**:244–246.

Evagelopoulou P, Myrianthefs P, Markogiannakis A, Baltopoulos G, Tsakris A. Multidrug-resistant *Klebsiella pneumoniae* mediastinitis safely and effectively treated with prolonged administration of tigecycline. *Clin Infect Dis* 2008;**46**:1932–1933.

Fraser TG, Reiner S, Malczynski M, Yarnold PR, Warren J, Noskin GA. Multidrug-resistant *Pseudomonas aeruginosa* cholangitis after endoscopic retrograde cholangiopancreatography: failure of routine endoscope cultures to prevent an outbreak. *Infect Control Hosp Epidemiol* 2004;**25**:856–859.

Furtado GHC, d'Azevedo PA, Santos AF, Gales AC, Pignatari ACC, Medeiros EAS. Intravenous polymyxin B for the treatment of nosocomial pneumonia caused by multidrug-resistant *Pseudomonas aeruginosa*. *Int J Antimicrob Agents* 2007;**30**:315–319.

Goldoni S, Galassi P, Gandolfi P, *et al.* Evaluation of the efficacy and safety of aztreonam in the treatment of urinary infections due to multiresistant Gram-negative bacteria. *Curr Ther Res Clin Exp* 1987;**42**:880–888.

Helm EB, Munk I, Shah PM, Stille W. Elimination of bacteria during antibacterial chemotherapy – a neglected parameter of chemotherapy. *Infection* 1979;**7**(Suppl. 5):S492–S494.

Khawcharoenporn T, Apisarnthanarak A, Mundy LM. Intrathecal colistin for drug-resistant *Acinetobacter baumannii* central nervous system infection: a case series and systematic review. *Clin Microbiol Infect* 2010;**16**:888–894.

Kumarasamy K, Kalyanasundaram A. Emergence of *Klebsiella pneumoniae* isolate co-producing NDM-1 with KPC-2 from India. *J Antimicrob Chemother* 2012;**67**:243–244.

Manzar S. Outbreak of multidrug resistant *Acinetobacter* in the neonatal intensive care unit. *Saudi Med J* 2004;**25**:961–963.

Min SS, Weber DJ, Donovan BJ, *et al.* Multidrug-resistant *Enterococcus faecium* in a patient with burns. *Clin Infect Dis* 2003;**36**:1210–1211.

Palasubramaniam S, Subramaniam G, Muniandy S, Parasakthi N. SHV-5 extended-spectrum betalactamase from *Klebsiella pneumoniae* associated with a nosocomial outbreak in a paediatric oncology unit in Malaysia. *Int J Infect Dis* 2005;**9**:170–172.

Paz A, Bauer H, Potasman I. Multiresistant *Pseudomonas aeruginosa* outbreak associated with contaminated transrectal ultrasound. *J Hosp Infect* 2001;**49**:148–149.

Reish O, Ashkenazi S, Naor N, Samra Z, Merlob P. An outbreak of multiresistant *Klebsiella* in a neonatal intensive care unit. *J Hosp Infect* 1993;**25**:287–291.

Rodriguez CH, Barberis C, Nastro M, *et al.* Impact of heteroresistance to colistin in meningitis caused by *Acinetobacter baumannii. J Infect* 2012;**64**:119–121.

Rosanova M, Epelbaum C, Noman A, *et al.* Use of colistin in a pediatric burn unit in Argentina. *J Burn Care Res* 2009;**30**:612–615.

Spanik S, Lacka J, Koren P, *et al.* Increasing incidence of carbapenem-resistant *Pseudomonas aeruginosa* bacteraemia in a cancer centre over a seven-year period. *J Hosp Infect* 1997;**35**:250–251.

Cross-sectional

Augustin A, Shahum A, Kalavsky E, Liskova A, Kisac P, Krcmery V. Colonization of the respiratory tract by drug-resistant bacteria in HIV-infected children and prior exposure to antimicrobials. *Med Sci Monitor* 2008;**14**:SC19–SC22.

Beerepoot MAJ, den Heijer CDJ, Penders J, Prins JM, Stobberingh EE, Geerlings SE. Predictive value of *Escherichia coli* susceptibility in strains causing asymptomatic bacteriuria for women with recurrent symptomatic urinary tract infections receiving prophylaxis. *Clin Microbiol Infect* 2012;**18**:E84–E90.

Behiry IK, Abada EA, Ahmed EA, Labeeb RS. Enteropathogenic *Escherichia coli* associated with diarrhea in children in Cairo, Egypt. *Sci World* 2011;**11**:2613–2619.

Cordero L, Rau R, Taylor D, Ayers LW. Enteric Gram-negative bacilli bloodstream infections: 17 years' experience in a neonatal intensive care unit. *Am J Infect Control* 2004;**32**:189–195.

Crandon JL, Kuti JL, Jones RN, Nicolau DP. Comparison of 2002–2006 OPTAMA programs for US hospitals: focus on Gram-negative resistance. *Ann Pharmacother* 2009;**43**:220–227.

Ejrnaes K. Bacterial characteristics of importance for recurrent urinary tract infections caused by *Escherichia coli*. *Danish Med Bull* 2011;**58**:B4187.

Fadel R, Dakdouki GK, Kanafani ZA, Araj GF, Kanj SS. Clinical and microbiological profile of urinary tract infection at a tertiary-care center in Lebanon. *Infect Control Hosp Epidemiol* 2004;**25**:82–85.

Giamarellou H, Galanakis N. Use of intravenous ciprofloxacin in difficult-to-treat infections. *Am J Med* 1987;**82**:346–351.

losifidis E, Antachopoulos C, Tsivitanidou M, *et al.* Differential correlation between rates of antimicrobial drug consumption and prevalence of antimicrobial resistance in a tertiary care hospital in Greece. *Infect Control Hosp Epidemiol* 2008;**29**:615–622.

Jeddi R, Ghedira H, Ben Amor R, *et al.* Risk factors of septic shock in patients with hematologic malignancies and *Pseudomonas* infections. *Hematology* 2011;**16**:160–165.

Jenkinson L, Smullen J, Weightman NC, Kerr KG. Changes in gastrointestinal carriage of multiresistant Gram-negative bacilli in a predominantly rural population served by a district general hospital. *J Clin Pathol* 2012;**65**:376–377.

Jukemura EM, Burattini MN, Pereira CAP, Braga ALF, Medeiros EAS. Control of multi-resistant bacteria and ventilator-associated pneumonia: Is it possible with changes in antibiotics? *Braz J Infect Dis* 2007;**11**:418–422.Livesey JE, Chiew Y-F. Antimicrobial drug utilisation in Dunedin Hospital, New Zealand, and its association with antimicrobial resistance. *Pathology* 2006;**38**:245–248.

Khosravi Y, Tay ST, Jamuna V. First characterization of blaIMP and blaVIM cassette containing novel integron in metallo-beta-lactamase producing *Pseudomonas aeruginosa* in Malaysia. *Int J Infect Dis* 2010;**14**:e37.

Kiani QH, Amir M, Ghazanfar MA, Iqbal M. Microbiology of wound infections among hospitalised patients following the 2005 Pakistan earthquake. *J Hosp Infect* 2009;**73**:71–78.

Martins-Loureiro M, de Moraes BA, de Mendonca VL, Rocha-Quadra MR, dos Santos-Pinheiro G, Dutra-Asensi M. Molecular epidemiology of extended-spectrum beta-lactamase-producing *Klebsiella pneumoniae* isolated from neonatal intensive care unit patients involved in hospital infection cases in Rio de Janeiro, Brazil. *Rev Latinoam Microbiol* 2001;**43**:88–95.

Paladino JA, Sunderlin JL, Singer ME, Adelman MH, Schentag JJ. Influence of extended-spectrum beta-lactams on Gram-negative bacterial resistance. *Am J Health Syst Pharm* 2008;**65**:1154–1159.

Ramakant P, Verma AK, Misra R, *et al.* Changing microbiological profile of pathogenic bacteria in diabetic foot infections: time for a rethink on which empirical therapy to choose? *Diabetologia* 2011;**54**:58–64.

Rosenthal EJK. Epidemiology of septicaemia pathogens. *Deutsche Med Wochenschr* 2002;**127**:2435–2440.

Savas L, Duran N, Savas N, Onlen Y, Ocak S. The prevalence and resistance patterns of *Pseudomonas aeruginosa* in intensive care units in a university hospital. *Turk J Med Sci* 2005;**35**:317–322.

Sepp E, Stsepetova J, Loivukene K, *et al.* The occurrence of antimicrobial resistance and class 1 integrons among commensal *Escherichia coli* isolates from infants and elderly persons. *Ann Clin Microbiol Antimicrob* 2009;**8**:34.

Urbánek K, Kolář M, Lovečková Y, Strojil J, Šantavá L. Influence of third-generation cephalosporin utilization on the occurrence of ESBL-positive *Klebsiella pneumoniae* strains. *J Clin Pharm Ther* 2007;**32**:403–408.

Vavilov VN, Ponomarenko OB, Popova MO, *et al.* Epidemiology of bacterial infections and antibiotic resistance in BMT clinic: a single center experience. *Cell Ther Transplant* 2009;**2**:123.

Veldman K, Cavaco LM, Mevius D, *et al.* International collaborative study on the occurrence of plasmid-mediated quinolone resistance in *Salmonella enterica* and *Escherichia coli* isolated from animals, humans, food and the environment in 13 European countries. *J Antimicrob Chemother* 2011;**66**:1278–1286.

Viray M, Linkin D, Maslow JN, *et al.* Longitudinal trends in antimicrobial susceptibilities across long-term-care facilities: emergence of fluoroquinolone resistance. *Infect Control Hosp Epidemiol* 2005;**26**:56–62.

Wiener J, Quinn JP, Bradford PA, et al. Multiple antibiotic-resistant *Klebsiella* and *Escherichia coli* in nursing homes. *JAMA* 1999;**281**:517–523.

In-vitro studies

Al-Kaabi MR, Tariq WUZ, Hassanein A. Rising bacterial resistance to common antibiotics in Al Ain, United Arab Emirates. *East Med Health J* 2011;**17**:479–484.

Blaettler L, Mertz D, Frei R, *et al.* Secular trend and risk factors for antimicrobial resistance in *Escherichia coli* isolates in Switzerland 1997–2007. *Infection* 2009;**37**:534–539.

Campana S, Taccetti G, Farina S, Ravenni N, De Martino M. Antimicrobial susceptibility and synergistic activity of meropenem against Gram-negative non-fermentative bacteria isolated from cystic fibrosis patients. *J Chemother* 2003;**15**:551–554.

Chromy BA, Elsheikh M, Christensen TL, *et al.* Repurposing screens identify rifamycins as potential broad-spectrum therapy for multidrug-resistant *Acinetobacter baumannii* and select agent microorganisms. *Future Microbiol* 2012;**7**:1011–1020.

Erlandsson M, Gill H, Nordlinder D, *et al.* Antibiotic susceptibility patterns and clones of *Pseudomonas aeruginosa* in Swedish ICUs. *Scand J Infect Dis* 2008;**40**:487–494.

Falagas ME, Kanellopoulou MD, Karageorgopoulos DE, *et al.* Antimicrobial susceptibility of multidrugresistant Gram negative bacteria to fosfomycin. *Eur J Clin Microbiol Infect Dis* 2008;**27**:439–443.

Fooladi AAI, Sattari M, Pourbabaei AA, Gholami M. Relation between quinolones and beta-lactams resistance with feature of producing capsules in *Pseudomonas aeruginosa* isolated from urine. *Med Sci J Islamic Azad Univ Tehran Med Branch* 2009;**19**:7.

Gustafsson I, Sjolund M, Torell E, *et al.* Bacteria with increased mutation frequency and antibiotic resistance are enriched in the commensal flora of patients with high antibiotic usage. *J Antimicrob Chemother* 2003;**52**:645–650.

Guyot A, Barrett SP, Threlfall EJ, Hampton MD, Cheasty T. Molecular epidemiology of multi-resistant *Escherichia coli. J Hosp Infect* 1999;**43**:39–48.

Kansal R, Pandey A, Asthana AK. beta-lactamase producing *Acinetobacter* species in hospitalized patients. *Ind J Pathol Microbiol* 2009;**52**:456–457

Luo Y, Cui S, Li J, *et al.* Characterization of *Escherichia coli* isolates from healthy food handlers in hospital. *Microb Drug Resist* 2011;**17**:443–448.

Messai L, Achour W, Ben Hassen A. Epidemiological profile of enterobacteria isolated from neutropenic patients. *Pathol Biol* 2007;**55**:230–234.

Moskowitz SM, Garber E, Chen Y, *et al.* Colistin susceptibility testing: evaluation of reliability for cystic fibrosis isolates of *Pseudomonas aeruginosa* and *Stenotrophomonas maltophilia*. *J Antimicrob Chemother* 2010;**65**:1416–1423.

Oguttu JW, Veary CM, Picard JA. Antimicrobial drug resistance of *Escherichia coli* isolated from poultry abattoir workers at risk and broilers on antimicrobials. *J S Afr Vet Assoc* 2008;**79**:161–166.

Shigemura K, Tanaka K, Okada H, *et al.* Pathogen occurrence and antimicrobial susceptibility of urinary tract infection cases during a 20-year period (1983–2002) at a single institution in Japan. *Jpn J Infect Dis* 2005;**58**:303–308.

Shin J, Kim DH, Ko KS. Comparison of CTX-M-14- and CTX-M-15-producing *Escherichia coli* and *Klebsiella pneumoniae* isolates from patients with bacteremia. *J Infect* 2011;**63**:39–47.

Sinirtas M, Akalin H, Gedikoglu S. Investigation of colistin sensitivity via three different methods in *Acinetobacter baumannii* isolates with multiple antibiotic resistance. *IntJ Infect Dis* 2009;**13**:e217–e220.

Spence RP, Towner KJ, Henwood CJ, James D, Woodford N, Livermore DM. Population structure and antibiotic resistance of *Acinetobacter* DNA group 2 and 13TU isolates from hospitals in the UK. *J Med Microbiol* 2002;**51**:1107–1112.

Stephens C, Francis SJ, Abell V, DiPersio JR, Wells P. Emergence of resistant *Acinetobacter baumannii* in critically ill patients within an acute care teaching hospital and a long-term acute care hospital. *Am J Infect Control* 2007;**35**:212–215.

Szucs O, Ozse M, Kristof K, Darvas K, Csomos A. Pattern of pathogens in hospital-acquired bloodstream infections: 2 years comparison. *Intensive Care Med* 2011;**37**:S19.

Taherikalani M, Etemadi G, Geliani KN, Fatollahzadeh B, Soroush S, Feizabadi MM. Emergence of multi and pan-drug resistance *Acinetobacter baumannii* carrying blaOXA-type-carbapenemase genes among burn patients in Tehran, Iran. *Saudi Med J* 2008;**29**:623–624.

Tato M, Valverde A, Coque TM, Canton R. PER-1 multiresistant *Pseudomonas aeruginosa* strain in Spain. *Enferm Infec Microbiol Clin* 2006;**24**:472–473.

Vettoretti L, Floret N, Hocquet D, *et al.* Emergence of extensive-drug-resistant *Pseudomonas aeruginosa* in a French university hospital. *Eur J Clin Microbiol Infect Dis* 2009;**28**:1217–1222.

Wada K, Kariyama R, Mitsuhata R, *et al.* Experimental and clinical studies on fluoroquinoloneinsusceptible *Escherichia coli* isolated from patients with urinary tract infections from 1994 to 2007. *Acta Med Okayama* 2009;**63**:263–272.

Zavascki AP, Cruz RP, Goldani LZ. High rate of antimicrobial resistance in *Pseudomonas aeruginosa* at a tertiary-care teaching hospital in southern Brazil. *Infect Control Hosp Epidemiol* 2004;**25**:805–807.

Prospective cohort

Acar A, Oncul O, Ozyurt M, Budak S, Gorenek L, Haznedaroglu T. *Acinetobacter baumannii* infections in intensive care units patients: predictors of risk factors of multidrug resistance. *Clin Microbiol Infect* 2010;**16**:S368.

Acolet D, Ahmet Z, Houang E, Hurley R, Kaufmann ME. *Enterobacter cloacae* in a neonatal intensive care unit: account of an outbreak and its relationship to use of third generation cephalosporins. *J Hosp Infect* 1994;**28**:273–286.

Alvarez-Lerma F, Palomar M, Olaechea P, *et al.* Changes of multiresistant markers in the ICU. 2005-2008 data. *Intensive Care Med* 2009;**35**:S268.

Brown BJ, Asinobi AO, Fatunde OJ, Osinusi K, Fasina NA. Antimicrobial sensitivity pattern of organisms causing urinary tract infection in children with sickle cell anaemia in Ibadan, Nigeria. *W Afr J Med* 2003;**22**:110–113.

Calil R, Marba STM, Von Nowakonski A, Tresoldi AT. Reduction in colonization and nosocomial infection by multiresistant bacteria in a neonatal unit after institution of educational measures and restriction in the use of cephalosporins. *Am J Infect Control* 2001;**29**:133–138.

Ciobotaro P, Oved M, Nadir E, Bardenstein R, Zimhony O. An effective intervention to limit the spread of an epidemic carbapenem-resistant *Klebsiella pneumoniae* strain in an acute care setting: from theory to practice. *Am J Infect Control* 2011;**39**:671–677.

Elnasser ZA, Al Aseel SM. Antibiotic resistance of *Pseudomonas aeruginosa* isolates from patients in King Abdullah University Hospital in Jordan. *J Chemother* 2009;**21**:356–359.

Fernandez J, Acevedo J, Castro M, *et al.* Prevalence and risk factors of infections by multiresistant bacteria in cirrhosis: a prospective study. *Hepatology* 2012;**55**:1551–1561.

Gomes CC, Vormittag E, Santos CR, Levin AS. Nosocomial infection with cephalosporin-resistant *Klebsiella pneumoniae* is not associated with increased mortality. *Infect Control Hosp Epidemiol* 2006;**27**:907–912.

Gouin F, Papazian L, Martin C, *et al.* A non-comparative study of the efficacy and tolerance of cefepime in combination with amikacin in the treatment of severe infections in patients in intensive care. *J Antimicrob Chemother* 1993;**32**(Suppl. B):205–214.

Gudiol C, Tubau F, Calatayud L, *et al.* Bacteraemia due to multidrug-resistant Gram-negative bacilli in cancer patients: risk factors, antibiotic therapy and outcomes. *J Antimicrob Chemother* 2011;**66**:657–663.

Heppt W, Lutz H. Clinical experiences with ofloxacin sequential therapy in chronic ear infections. *Eur Arch Otorhinolaryngol* 1993;**250**(Suppl. 1):S19–S21.

Limat S, Cornette C, Deconinck E, Woronoff-Lemsi MC, Cahn JY. Antibiotic therapy of febrile neutropenic patients: prospective study at a hematologic department. *Presse Med* 1999;**28**:729–733.

Lu Q, Luo R, Bodin L, *et al.* Efficacy of high-dose nebulized colistin in ventilator-associated pneumonia caused by multidrug-resistant *Pseudomonas aeruginosa* and *Acinetobacter baumannii*. *Anesthesiology* 2012;**117**:1335–1347.

Markogiannakis A, Fildisis G, Tsiplakou S, *et al.* Cross-transmission of multidrug-resistant *Acinetobacter baumannii* clonal strains causing episodes of sepsis in a trauma intensive care unit. *Infect Control Hosp Epidemiol* 2008;**29**:410–417.

Martin C, Ofotokun I, Rapp R, *et al.* Results of an antimicrobial control program at a university hospital. *Am J Health Syst Pharm* 2005;**62**:732–738.

Mastoraki A, Douka E, Kriaras I, Stravopodis G, Manoli H, Geroulanos S. *Pseudomonas aeruginosa* susceptible only to colistin in intensive care unit patients. *Surg Infect* 2008;**9**:153–160.

Metan G, Sariguzel F, Sumerkan B. Factors influencing survival in patients with multi-drug-resistant *Acinetobacter* bacteraemia. *Eur J Intern Med* 2009;**20**:540–544.

Meyer KS, Urban C, Eagan JA, Berger BJ, Rahal JJ. Nosocomial outbreak of *Klebsiella* infection resistant to late-generation cephalosporins. *Ann Intern Med* 1993;**119**:353–358.

Millar M, Philpott A, Wilks M, *et al.* Colonization and persistence of antibiotic-resistant Enterobacteriaceae strains in infants nursed in two neonatal intensive care units in East London, United Kingdom. *J Clin Microbiol* 2008;**46**:560–567.

Mukhopadhyay C, Chawla K, Krishna S, Nagalakshmi N, Rao SP, Bairy I. Emergence of *Burkholderia pseudomallei* and pandrug-resistant non-fermenters from southern Karnataka, India. *Trans R Soc Trop Med Hyg* 2008;**102**(Suppl. 1):S12–S17.

Narchi H, Al-Hamdan MA. Antibiotic resistance trends in paediatric community-acquired first urinary tract infections in the United Arab Emirates. *East Med Health J* 2010;**16**:45–50.

Nseir S, Di Pompeo C, Soubrier S, *et al.* First-generation fluoroquinolone use and subsequent emergence of multiple drug-resistant bacteria in the intensive care unit. *Crit Care Med* 2005;**33**:283–289.

Ortega B, Groeneveld ABJ, Schultsz C. Endemic multidrug-resistant *Pseudomonas aeruginosa* in critically ill patients. *Infect Control Hosp Epidemiol* 2004;**25**:825–831.

Paauw A, Verhoef J, Fluit AC, *et al.* Failure to control an outbreak of qnrA1-positive multidrugresistant *Enterobacter cloacae* infection despite adequate implementation of recommended infection control measures. *J Clin Microbiol* 2007;**45**:1420–1425.

Parra Moreno ML, Arias Rivera S, de La Cal Lopez MA, *et al.* Effect of selective digestive decontamination on the nosocomial infection and multiresistant microorganisms incidence in critically ill patients. *Med Clin* 2002;**118**:361–364.

Presentado JCM, Lopez DP, Castiglioni MB, Gerez J. Ceftriaxone and ciprofloxacin restriction in an intensive care unit: less incidence of *Acinetobacter* spp. and improved susceptibility of *Pseudomonas aeruginosa. Rev Panam Salud Publ* 2011;**30**:603–609.

Rafat C, Vimont S, Ancel PY, *et al.* Ofloxacin: new applications for the prevention of urinary tract infections in renal graft recipients. *Transplant Infect Dis* 2011;**13**:344–352.

Rogers BA, Sidjabat HE, Paterson DL, Kennedy K, Collignon P, Jones M. Prolonged carriage of resistant *E. coli* by returned travellers: clonality, risk factors and bacterial characteristics. *Eur J Clin Microbiol Infect Dis* 2012;**31**:2413–2420.

Saavedra S, Ramirez-Ronda CH, Nevarez M. Ciprofloxacin in the treatment of urinary tract infections caused by *Pseudomonas aeruginosa* and multiresistant bacteria. *Eur J Clin Microbiol* 1986;**5**:255–

257.

Sandiumenge A, Lisboa T, Gomez F, Hernandez P, Canadell L, Rello J. Effect of antibiotic diversity on ventilator-associated pneumonia caused by ESKAPE organisms. *Chest* 2011;**140**:643–651.

Slim E, Smit CA, Bos AJ, Peerbooms PG. Nosocomial transmission of highly resistant microorganisms on a spinal cord rehabilitation ward. *J Spinal Cord Med* 2009;**32**:422–427.

Stein GE. Serum bactericidal activity of trovafloxacin against drug-resistant respiratory pathogens. *Drugs* 1999;**58**:356–357.

Suankratay C, Jutivorakool K, Jirajariyavej S. A prospective study of ceftriaxone treatment in acute pyelonephritis caused by extended-spectrum beta-lactamase-producing bacteria. *J Med Assoc Thailand* 2008;**91**:1172–1181.

Tacconelli E, De Angelis G, Cataldo MA, *et al.* Antibiotic usage and risk of colonization and infection with antibiotic-resistant bacteria: a hospital population-based study. *Antimicrob Agents Chemother.* 2009;**53**:4264–4269.

Thabet L, Memmi M, Turki A, Messadi AA. The impact of fluoroquinolones use on antibiotic resistance in an intensive care burn department. *Tunisie Med* 2010;**88**:696–699.

Van Der Voort PHJ, Van Roon EN, *et al.* A before–after study of multi-resistance and cost of selective decontamination of the digestive tract. *Infection* 2004;**32**:271–277.

Van Ruler O, Kiewiet JJS, Van Ketel RJ, Boermeester MA. Initial microbial spectrum in severe secondary peritonitis and relevance for treatment. *Eur J Clin Microbiol Infect Dis* 2012;**31**:671–682.

Wybo I, Blommaert L, De Beer T, *et al.* Outbreak of multidrug-resistant *Acinetobacter baumannii* in a Belgian university hospital after transfer of patients from Greece. *J Hosp Infect* 2007;**67**:374–380.

Yang K, Guglielmo BJ. Diagnosis and treatment of extended-spectrum and AmpC beta-lactamaseproducing organisms. *Ann Pharmacother* 2007;**41**:1427–1435.

Zhang J-P, Yang X-S, Chen J, Peng Y-Z, Huang Y-S. Clinical assessment of colistin in treating infections caused by multidrug-resistant Gram-negative bacillus in patients with severe burn. *Zhonghua Shao Shang Za Zhi* 2009;**25**:372–376.

Surveillance

Andrade SS, Jones RN, Gales AC, Sader HS. Increasing prevalence of antimicrobial resistance among *Pseudomonas aeruginosa* isolates in Latin American medical centres: 5 year report of the SENTRY Antimicrobial Surveillance Program (1997–2001). *J Antimicrob Chemother* 2003;**52**:140–141.

Baang JH, Axelrod P, Decker BK, *et al.* Longitudinal epidemiology of multidrug-resistant (MDR) *Acinetobacter* species in a tertiary care hospital. *Am J Infect Control* 2012;**40**:134–137.

Behera B, Mathur P. High levels of antimicrobial resistance at a tertiary trauma care centre of India. *Ind J Med Res* 2011;**133**:343–345.

Bou G, Cervero G, Dominguez MA, Quereda C, Martinez-Beltran J. Characterization of a nosocomial outbreak caused by a multiresistant *Acinetobacter baumannii* strain with a carbapenem-hydrolyzing enzyme: high-level carbapenem resistance in *A. baumannii* is not due solely to the presence of beta-lactamases. *J Clin Microbiol* 2000;**38**:3299–3305.

Brink A, Feldman C, Richards G, Moolman J, Senekal M. Emergence of extensive drug resistance (XDR) among Gram-negative bacilli in South Africa looms nearer. *S Afr Med J* 2008;**98**:586–592.

Cabrita J, Oleastro M, Lopes MM, Barros R, Peres I, Pires I. Molecular epidemiologic analysis of multiresistant *Klebsiella pneumoniae* isolates from a pediatric hospital. *Clin Microbiol Infect* 1999;**5**:109–112.

Cardoso O, Alves AF, Leitao R. Surveillance of antimicrobial susceptibility of *Pseudomonas aeruginosa* clinical isolates from a central hospital in Portugal. *J Antimicrob Chemother* 2007;**60**:452–454.

Daoud Z, Moubareck C, Hakime N, Doucet-Populaire F. Extended spectrum beta-lactamase producing enterobacteriaceae in Lebanese ICU patients: epidemiology and patterns of resistance. *J Gen Appl Microbiol* 2006;**52**:169–178.

De Gheldre Y, Maes N, Rost F, *et al.* Molecular epidemiology of an outbreak of multidrug-resistant *Enterobacter aerogenes* infections and in vivo emergence of imipenem resistance. *J Clin Microbiol* 1997;**35**:152–160.

Filius PMG, Gyssens IC, Kershof IM, *et al.* Colonization and resistance dynamics of Gram-negative bacteria in patients during and after hospitalization. *Antimicrob Agents Chemother* 2005;**49**:2879–2886.

Furtado GHC, Perdiz LB, Medeiros EAS. The effect of a 4th generation-cephalosporin introduction upon the incidence of multidrug-resistant Gram-negative bacteria in a non-teaching hospital. *Am J Infect Dis* 2008;**4**:267–271.

Gomes MZR, Machado CR, De Souza da Conceicao M, *et al.* Outbreaks, persistence, and high mortality rates of multiresistant *Pseudomonas aeruginosa* infections in a hospital with AIDS-predominant admissions. *Braz J Infect Dis* 2011;**15**:312–322.

Gottig S, Pfeifer Y, Wichelhaus TA, *et al.* Global spread of New Delhi metallo-beta-lactamase 1. *Lancet Infect Dis* 2010;**10**:828–829.

Gottlieb T, Bradbury R, Cheong E. The resistible rise and fall of a burns and ICU-related hospital outbreak of multi-resistant *Acinetobacter baumannii*. *Clin Microbiol Infect* 2010;**16**:S411.

Kim JY, Sohn JW, Park DW, Yoon YK, Kim YM, Kim MJ. Control of extended-spectrum betalactamase-producing *Klebsiella pneumoniae* using a computer-assisted management program to restrict third-generation cephalosporin use. *J Antimicrob Chemother* 2008;**62**:416–421.

Kochar S, Sheard T, Sharma R, *et al.* Success of an infection control program to reduce the spread of carbapenem-resistant *Klebsiella pneumoniae. Infect Control Hosp Epidemiol* 2009;**30**:447–452.

Lemmen SW, Hafner H, Kotterik S, Lutticken R, Topper R. Influence of an infectious disease service on antibiotic prescription behavior and selection of multiresistant pathogens. *Infection* 2000;**28**:384–387.

Liu WL, Chang PC, Chen YY, Lai CC. Impact of fluoroquinolone consumption on resistance of healthcare-associated *Pseudomonas aeruginosa*. *J Infect* 2012;**64**:335–337.

Livermore DM, Hill RLR, Thomson H, *et al.* Antimicrobial treatment and clinical outcome for infections with carbapenem- and multiply-resistant *Acinetobacter baumannii* around London. *Int J Antimicrob Agents* 2010;**35**:19–24.

Low DE, Markovic MJ, Dowzicky MJ. Antimicrobial susceptibility among bacterial isolates from ICU and non-ICU setting and different age groups: results from the tigecycline evaluation and surveillance trial in North America. *J Chemother* 2009;**21**:16–23.

Luzzaro F, Vigano EF, Fossati D, *et al.* Prevalence and drug susceptibility of pathogens causing bloodstream infections in northern Italy: a two-year study in 16 hospitals. *Eur J Clin Microbiol Infect Dis* 2002;**21**:849–855.

MacDougall C, Harpe SE, Powell JP, Johnson CK, Edmond MB, Polk RE. *Pseudomonas aeruginosa*, *Staphylococcus aureus*, and fluoroquinolone use. *Emerg Infect Dis* 2005;**11**:1197–1204.

Marra AR, de Almeida SM, Correa L, *et al.* The effect of limiting antimicrobial therapy duration on antimicrobial resistance in the critical care setting. *Am J Infect Control* 2009;**37**:204–209.

Mazzarello MG. Action of several new antibiotics on multi-resistant bacterial strains isolated in the urine of hospitalised patients. *Gazz Med Ital Arch Sci Med* 1988;**147**:155–158.

Miano TA, Powell E, Schweickert WD, Morgan S, Binkley S, Sarani B. Effect of an antibiotic algorithm on the adequacy of empiric antibiotic therapy given by a medical emergency team. *J Crit Care* 2012;**27**:45–50.

Monzon H, Salvado M, Estelrich M, Gasos A, Serrano C, Montaner F. Impact of an antimicrobial stewardship program in a second level hospital. *Clin Microbiol Infect* 2011;**17**:S725.

Munoz-Price LS, Hayden MK, Lolans K, *et al.* Successful control of an outbreak of *Klebsiella pneumoniae* carbapenemase-producing *K. pneumoniae* at a long-term acute care hospital. *Infect Control Hosp Epidemiol* 2010;**31**:341–347.

Muzaheed,Doi Y, Adams-Haduch JM, *et al.* High prevalence of CTX-M-15-producing *Klebsiella pneumoniae* among inpatients and outpatients with urinary tract infection in Southern India. J Antimicrob Chemother 2008;**61**:1393–1394.

Pardo Serrano FJ, Tirado Balaguer MD, Garcia Zuniga ED, Granados Ortega J, Campos Aznar A, Moreno Munoz R. *Pseudomonas aeruginosa*: antimicrobial resistance in clinical isolates. Castellon 2004–2008. *Rev Espanol Quimioter* 2010;**23**:20–26.

Pires dos Santos R, Jacoby T, Pires Machado D, *et al.* Hand hygiene, and not ertapenem use, contributed to reduction of carbapenem-resistant pseudomonas aeruginosa rates. *Infect Control Hosp Epidemiol* 2011;**32**:584–590.

Polemis M, Trifinopoulou K, Chrisochoidou S, *et al.* Bacteraemia and antibiotic resistance of its pathogens reported in Greece between 2000 and 2009: trend analysis. *Clin Microbiol Infect* 2011;**17**:S423.

Pop-Vicas AE, D'Agata EMC. The rising influx of multidrug-resistant Gram-negative bacilli into a tertiary care hospital. *Clin Infect Dis* 2005;**40**:1792–1798.

Suman E, Gopalkrishna Bhat K. Urinary tract infection in children due to drug-resistant bacteria - a study from South India. *J Trop Pediatr* 2001;**47**:374.

Szucs O, Kristof K, Darvas K, Csomos A. Changes in the incidence of multiresistant pathogens and its consequences in the intensive care unit. *Orvosi Hetilap* 2011;**152**:1486–1491.

Takesue Y, Nakajima K, Ichiki K, *et al.* Impact of a hospital-wide programme of heterogeneous antibiotic use on the development of antibiotic-resistant Gram-negative bacteria. *J Hosp Infect* 2010;**75**:28–32.

Tamayo J, Orden B, Cacho J, Cuadros J, Gomez-Garces JL, Alos JI. Activity of ertapenem and other antimicrobials against ESBL-producing enterobacteria isolated from urine in patients from Madrid. *Rev Espanol Quimioter* 2007;**20**:334–338.

Tomasoni D, Gattuso G, Scalzini A, *et al. Enterobacter cloacae* in an Italian Neonatal Intensive Care Unit: pattern of drug resistance compared with an international database (SENTRY Antimicrobial Surveillance Program). *J Chemother* 2006;**18**:110–111.

Turner PJ. Meropenem activity against European isolates: report on the MYSTIC (Meropenem Yearly Susceptibility Test Information Collection) 2006 results. *Diagn Microbiol Infect Dis* 2008;**60**:185–192.

Van Der Mee-Marquet N, Valentin-Domelier AS, Charbonnier T, *et al.* Characteristics of the multiresistant bacteria that actually diffuse in the Centre region, France, in and out of healthcare institutions. *Clin Microbiol Infect* 2010;**16**:S29.

Wroblewska MM, Rudnicka J, Marchel H, Luczak M. Multidrug-resistant bacteria isolated from patients hospitalised in intensive care units. *Int J Antimicrob Agents* 2006;**27**:285–289.

Narrative reviews, commentaries or editorials

Agarwal R. Do fluoroquinolones actually increase mortality in community-acquired pneumonia? *Crit Care* 2006;**10**:403.

Aguado JM. Role of new carbapenems in nosocomial intraabdominal infection. *Enferm Infec Microbiol Clin* 2010;**28**(Suppl. 2):65–68

Arya SC, Agarwal N, Solanki BS, Agarwal S. Use of cefepime for the treatment of infections caused by extended spectrum: beta-lactamase-producing *Klebsiella pneumoniae* and *Escherichia coli*. *Singapore Med J* 2007;**48**:600–601.

Barie PS. Clinical issues in the management of surgical infections, with an emphasis on antibiotic management of infections caused by multi-drug-resistant pathogens. *Surg Infect* 2005;**6**:S1–S3.

Barraclough KA, Hawley CM, Playford EG, Johnson DW. Prevention of access-related infection in dialysis. *Exp Rev Anti-infect Ther* 2009;**7**:1185–1200.

Baughman RP, Glauser MP. Managing serious infections in the hospital: a new model. *Clin Microbiol Infect* 2005;**11**:1–3.

Bhavnani SM. Antimicrobial usage and resistance problems: surveillance issues and a strategy for the future. *Antimicrob Infect Dis Newslett* 1998;**17**:41–47.

Bishop M. New strains of E. coli cause concern. Pharmaceut J 2005;275:770.

Bragesjo F, Hallberg M. Back to basics: Governing antibacterial resistance by means of mundane technoscience and accountability relations in a context of risk. *Health Risk Soc* 2011;**13**:691–709.

Cooper TW, Pass SE, Brouse SD, Hall IRG. Can pharmacokinetic and pharmacodynamic principles be applied to the treatment of multidrug-resistant *Acinetobacter? Ann Pharmacother* 2011;**45**:229–240.

Cunha BA. Pharmacokinetic considerations regarding tigecycline for multidrug-resistant (MDR) *Klebsiella pneumoniae* or MDR *Acinetobacter baumannii* urosepsis. *J Clin Microbiol* 2009;**47**:1613.

Cunha BA. Oral doxycycline for non-systemic urinary tract infections (UTIs) due to *P. aeruginosa* and other Gram negative uropathogens. *Eur J Clin Microbiol Infect Dis* 2012;**31**:2865–2868.

Curcio D. Tigecycline for treating ventilator-associated pneumonia: a practical perspective. *Diagn Microbiol Infect Dis* 2011;**69**:466–467.

Curcio D. Resistant pathogen-associated skin and skin-structure infections: antibiotic options. *Exp Rev Anti-infectTher* 2010;**8**:1019–1036.

Diomedi A. *Acinetobacter baumannii* pandrug-resistant: update in epidemiological and antimicrobial managing issues. *Rev Chil Infectol* 2005;**22**:298–320.

Drinka P, Niederman MS, El-Solh AA, Crnich CJ. Assessment of risk factors for multi-drug resistant organisms to guide empiric antibiotic selection in long term care: a dilemma. *J Am Med Direct Assoc* 2011;**12**:321–325.

Drissi M, Poirel L, Mugnier PD, Baba Ahmed Z, Nordmann P. Carbapenemase-producing *Acinetobacter baumannii*, Algeria. *Eur J Clin Microbiol Infect Dis* 2010;**29**:1457–1458.

Emonet S, Schrenzel J. How could rapid bacterial identification improve the management of septic patients? *Exp Rev Anti-infect Ther* 2011;**9**:707–709.

Evans ME, Feola DJ, Rapp RP. Polymyxin B sulfate and colistin: old antibiotics for emerging multiresistant Gram-negative bacteria. *Ann Pharmacother* 1999;**33**:960–967.

Falagas ME, Kasiakou SK, Michalopoulos A. Treatment of multidrug-resistant *Pseudomonas aeruginosa* and *Acinetobacter baumannii* pneumonia. *J Cystic Fibrosis* 2005;**4**:149–150.

Falagas ME, Kasiakou SK, Michalopoulos A. Polymyxins: a word of caution for prudent use of valuable 'old antibiotics' . *Infect Control Hosp Epidemiol* 2006;**27**:995.

Falagas ME, Koletsi PK, Bliziotis IA. The diversity of definitions of multidrug-resistant (MDR) and pandrug-resistant (PDR) *Acinetobacter baumannii* and *Pseudomonas aeruginosa*. *J Med Microbiol* 2006;**55**:1619–1629.

Filius PMG, Verbrugh HA. Epidemiology of antibiotic resistant bacteria: what is the course of resistance development. *Pharmaceut Weekblad* 2001;**136**:958–963.

Fish DN. Meropenem in the treatment of complicated skin and soft tissue infections. *Ther Clin Risk Manag* 2006;**2**:401–415.

Furukawa K. Importance of appropriate carbapenem use to reduce carbapenem-resistant *Pseudomonas aeruginosa. Nippon Rinsho* 2007;**65**(Suppl. 2):258–264.

Giamarellou H. Prescribing guidelines for severe *Pseudomonas* infections. *J Antimicrob Chemother* 2002;**49**:229–233.

Giske CG, Monnet DL, Cars O, Carmeli Y. Clinical and economic impact of common multidrugresistant Gram-negative bacilli. *Antimicrob Agents Chemother* 2008;**52**:813–821.

Gotoh N. Antibiotic resistant Pseudomonas aeruginosa. Nippon Rinsho 2003;61(Suppl. 3):196–201.

Heininger A, Unertl K. Antibiotic therapy: ventilator-associated pneumonia and multiresistant bacteria. *Anasthesiol Intensivmed Notfallmed Schmerzther* 2007;**42**:122–129.

Heyn D, Kober P. Results of a review of disinfectant use in public health institutions. *Hyg Med* 1992;**17**:145–148.

Hoban DJ, Zhanel GG. Introduction to the CANWARD Study (2007–2009). *Diagn Microbiol Infect Dis* 2011;**69**:289–290.

Horcajada JP, Farinas MC. Involvement of bacterial resistances in community-acquired urinary infections. *Enferm Infec Microbiol Clin* 2005;**23**:1–3.

Inglis TJJ, Beer CD. Multiresistant *Escherichia coli* in aged care: the gathering storm. The growing infection control challenges facing an ageing population. *Med J Aust* 2011;**195**:489–490.

Jain R, Danziger LH. Multidrug-resistant *Acinetobacter* infections: an emerging challenge to clinicians. *Ann Pharmacother* 2004;**38**:1449–1459.

Kaier K, Frank U, Meyer E. Economic incentives for the (over-)prescription of broad-spectrum antimicrobials in German ambulatory care. *J Antimicrob Chemother* 2011;**66**:1656–1658.

Kelesidis T, Karageorgopoulos DE, Kelesidis I, Falagas ME. Tigecycline for the treatment of multidrug-resistant Enterobacteriaceae: a systematic review of the evidence from microbiological and clinical studies. *J Antimicrob Chemother* 2008;**62**:895–904.

Kim C, Kim DG, Kang HR, *et al.* A trial of aerosolized colistin for the treatment of nosocomial pneumonia due to multidrug-resistant *Acinetobacter baumannii. Tuberc Respir Dis* 2008;**64**:102–108.

Kollef MH. An empirical approach to the treatment of multidrug-resistant ventilator-associated pneumonia. *Clin Infect Dis* 2003;**36**:1119–1121.

Kretzschmar A. Antibiotic therapy in intensive care medicine: new options for risk-adapted strategies. *Krankenhauspharmazie* 2009;**30**:560.

Kuper KM, Hirsch EB, Tam VH. Significant publications on infectious diseases pharmacotherapy in 2008. *Am J Health Syst Pharm* 2009;**66**:1726–1734.

Kwa AL, Tam VH, Falagas ME. Polymyxins: a review of the current status including recent developments. *Ann Acad Med Singapore* 2008;**37**:870–883.

Leggiadro RJ. Pediatric antimicrobial therapy. Curr Prob Pediatr 1993;23:315-321.

Levy SB, Zimmermann O, De Ciman R, *et al.* Bacteremia among Kenyan children (multiple letters). *N Engl J Med* 2005;**352**:1379–1381.

Lindemann H. Recent developments in prevention and treatment of *Pseudomonas* infection in CF patients. *Monatsschrift Kinderheilkunde* 2002;**150**:1224–1232.

Lopardo HA. Acinetobacter spp. and time-kill studies. J Antimicrob Chemother 2008;61:464.

Lowy FD. Antimicrobial resistance: the example of *Staphylococcus aureus*. *J Clin Investig* 2003;**111**:1265–1273.

Lubelski J, Konings WN, Driessen AJM. Distribution and physiology of ABC-type transporters contributing to multidrug resistance in bacteria. *Microbiol Molec Biol Rev* 2007;**71**:463–476.

Lynch AS. Antimicrobial resistance - 2005 Annual Conference. Science-prevention-control. 27–29 June 2005, Bethesda, MD, USA. *IDrugs* 2005;**8**:697–700.

Marcos RJ, Torres Marti A, Ariza Cardenal FJ, Alvarez Lerma F, Barcenilla Gaite F. Recommendations for the treatment of severe nosocomial pneumonia. *Med Intens* 2004;**28**:262–278.

Martinez-Martinez L. Bacterial death and heteroresistance to antimicrobial agents. *Enferm Infec Microbiol Clin* 2008;**26**:481–484.

Mathai E. Nosocomial bacteraemia & antimicrobial resistance in intensive care units. *Ind J Med Res* 2005;**122**:285–287.

Maviglia R, Nestorini R, Pennisi M. Role of old antibiotics in multidrug resistant bacterial infections. *Curr Drug Targets* 2009;**10**:895–905.

Mendelson M, Whitelaw A, Nicol M, Brink A. Wake up, South Africa! The antibiotic 'horse' has bolted. *S Afr Med J* 2012;**102**:607–608.

Michalopoulus A, Kasiakou SK, Falagas ME, Mubareka S, Rubinstein E. The significance of different formulations of aerosolized colistin (multiple letters). *Crit Care* 2005;**9**:417–418.

Mojtahedzadeh M, Mahmoudi L. Aminoglycoside resistance in ICUs: are we running out of drugs, for bad bugs. *Iran J Pharm Res* 2011;**10**:391–392.

Montefour K, Frieden J, Hurst S, *et al. Acinetobacter baumannii*: an emerging multidrug-resistant pathogen in critical care. *Crit Care Nurs* 2008;**28**:15–25; quiz 26.

Montero A, Corbella X, Ariza J. Clinical relevance of *Acinetobacter baumannii* ventilator-associated pneumonia. *Crit Care Med* 2003;**31**:2557–2559.

Moriyama B, Henning SA, Neuhauser MM, Danner RL, Walsh TJ. Continuous-infusion beta-lactam antibiotics during continuous venovenous hemofiltration for the treatment of resistant Gram-negative bacteria. *Ann Pharmacother* 2009;**43**:1324–1337.

Motaouakkil S, Charra B, Hachimi A, Benslama A. Nosocomial pneumonia caused by multiresistant *Acinetobacter baumanii* treated by colistin and rifampicin. *Ann Francais Anesthes Reanim* 2006;**25**:543–544.

Mulvey MR, Simor AE. Antimicrobial resistance in hospitals: how concerned should we be? *CMAJ* 2009;**180**:408–415.

Munoz Bellido JL. Problematic bacteria. Rev Espanol Quimioter 2008;21:2-6.

Nguyen S, Whitehill J. Treatment of urinary tract infections in children. US Pharmacist 2011;**36**:HS2–HS7.

Nicolau DP. Carbapenems: a potent class of antibiotics. Exp Opin Pharmacother 2008;9:23-37.

Nicolau DP. Management of complicated infections in the era of antimicrobial resistance: the role of tigecycline. *Exp Opin Pharmacother* 2009;**10**:1213–1222.

Niederman MS. Reexamining quinolone use in the intensive care unit: use them right or lose the fight against resistant bacteria. *Crit Care Med* 2005;**33**:443–444.

Nseir S. Aerosolized antibiotics are not a good idea – don't go with the flow: Premum Non Nocere! *Crit Care Med* 2009;**37**:800–801.

Obritsch MD, Fish DN, MacLaren R, Jung R. Nosocomial infections due to multidrug-resistant *Pseudomonas aeruginosa*: epidemiology and treatment options. *Pharmacotherapy* 2005;**25**:1353–1364.

Oliver A. Carbapenem resistance and *Acinetobacter baumannii*. Enferm Infec Microbiol Clin 2004;**22**:259–261.

Oncul O. Tertiary trauma care centre & antimicrobial resistance. Ind J Med Res 2011;134:238.

Ong CT, Kuti JL, Nightingale CH, Nicolau DP. Emerging *Pseudomonas aeruginosa* resistance: implications in clinical practice. *Connecticut Med* 2004;**68**:11–15.

Orhan-Sungur M, Akca O. Ventilator-associated pneumonia by multidrug-resistant bacteria: pathogen-specific risks versus care-related risks. *J Crit Care* 2007;**22**:26–27.

Oteo J, Alos JI. What's new in bacterial resistance to antimicrobials? *Enferm Infec Microbiol Clin* 2002;**20**:28–33.

Parry CM, Hien TT, Dougan G, White NJ, Farrar JJ. Typhoid fever. *N Engl J Med* 2002;**347**:1770–1782.

Perez F, Endimiani A, Bonomo RA. Why are we afraid of *Acinetobacter baumannii*? *Exp Rev Anti-Infect Ther* 2008;**6**:269–271.

Philippon A, Arlet G, Lagrange P. Extended spectrum beta-lactamases. *Rev Pratic* 1993;**43**:2387–2395.

Pickering LK. Emerging antibiotic resistance in enteric bacterial pathogens. *Semin Pediatr Infect Dis* 1996;**7**:272–280.

Poulakou G, Giamarellou H. Doripenem: an expected arrival in the treatment of infections caused by multidrug-resistant Gram-negative pathogens. *Exp Opin Investig Drugs* 2008;**17**:749–771.

Rapp RP. Antimicrobial resistance: insights into control and treatment of complicated infections – introduction. *Pharmacotherapy* 2005;**25**:41S–43S.

Rapp RP, Empey KM. Antimicrobial cycling to control bacterial resistance. *Ann Pharmacother* 2001;**35**:1289–1290.

Rice LB. Emerging issues in the management of infections caused by multi-drug-resistant, Gramnegative bacilli. *Surg Infect* 2005;**6**:S37–S47.

Rodriguez-Bano J, Bonomo RA. Multidrug-resistant *Acinetobacter baumannii*: 'eyes wide shut'? *Enferm Infec Microbiol Clin* 2008;**26**:185–186.

Rodriguez-Bano J, Pascual A. Hospital infection control in Spain. J Hosp Infect 2001;48:258–260.

Rotimi VO, Jamal W, Salama M. Control of acinetobacter outbreaks in the intensive care unit. *J Hosp Infect* 2009;**73**:286–287.

Ruf BR, Kern WV. Infectiology. Internist 1999;40:369-380.

Sabella C, Goldfarb J. Fluoroquinolone therapy in pediatrics: where we stand. *Clin Pediatr* 1997;**36**:445–448.

Sandel DC, Wang C-T, Kessler S. Urinary tract infections and a multidrug-resistant *Escherichia coli* clonal group. *N Engl J Med* 2002;**346**:535–536.

Sarma S, Nair D, Rawat D, *et al.* Burn wound septicemia – a pilot study from a tertiary care hospital. *Ann Trop Med Public Health* 2011;**4**:146–148.

Schwaber MJ, Carmeli Y. Carbapenem-resistant enterobacteriaceae: a potential threat. *JAMA* 2008;**300**:2911–2913.

Shears P. A review of bacterial resistance to antimicrobial agents in tropical countries. *Ann Trop Paediatr* 1993;**13**:219–226.

Silver LL. Antibacterial drug discovery & development - SRI's 11th Annual Summit. Antibacterial trends and current research, 10–11 April 2006, Princeton, NJ, USA. *IDrugs* 2006;**9**:394–397.

Simon A, Krawtschenko O, Reiffert SM, Exner M, Trautmann M, Engelhart S. Outbreaks of *Pseudomonas aeruginosa* in pediatric patients – clinical aspects, risk factors and management. *J Pediatr Infect Dis* 2008;**3**:249–269.

Slover CM, Rodvold KA, Danziger LH. Tigecycline: a novel broad-spectrum antimicrobial. *Ann Pharmacother* 2007;**41**:965–972.

Strateva T, Markova B, Marteva-Proevska Y, Ivanova D, Mitov I. Widespread dissemination of multidrug-resistant *Acinetobacter baumannii* producing OXA-23 carbapenemase and ArmA 16S ribosomal RNA methylase in a Bulgarian university hospital. *Brazi J Infect Dis* 2012;**16**:307–310.

Sun HY, Fujitani S, Quintiliani R, Yu VL. Pneumonia due to *Pseudomonas aeruginosa*. Part II: Antimicrobial resistance, pharmacodynamic concepts, and antibiotic therapy. *Chest* 2011;**139**:1172–1185.

Sydnor ERM, Perl TM. Hospital epidemiology and infection control in acute-care settings. *Clin Microbiol Rev* 2011;**24**:141–173.

Taubes G. The bacteria fight back. Science 2008;321:356-360, 361.

Tellado JM. The need for new antimicrobials for intra-abdominal infections (IAI): defining the forthcoming scenario. *Surg Infect* 2006;**7**:1–4.

Totsuka K, Kaku M, Kuwabara M, Rikitomi N, Fuchigami T. Antibiotic therapy in routine medical care: progress in diagnosis of and therapy for bacterial infections (discussion). *Nihon Naika Gakkai Zasshi* 2006;**95**:2256–2276.

Turkoglu M, Dizbay M. Multidrug-resistant *Acinetobacter baumannii* infection is not an independent risk factor for mortality in critically ill patients with hematologic malignancy. *J Crit Care* 2011;**26**:526–527.

Urban C, Segal-Maurer S, Rahal JJ. Considerations in control and treatment of nosocomial infections due to multidrug-resistant *Acinetobacter baumannii*. *Clin Infect Dis* 2003;**36**:1268–1274.

Van Delden C, Blumberg EA. Multidrug resistant Gram-negative bacteria in solid organ transplant recipients. *Am J Transplant* 2009;**9**:S27–S34.

Van Saene HKF, Stoutenbeek CP, Gilbertson AA. Review of available trials of selective decontamination of the digestive tract (SDD). *Infection* 1990;**18**:S5–S9.

Wagenlehner FME, Schmiemann G, Hoyme U, et al. National S3 guideline on uncomplicated urinary tract infection: recommendations for treatment and management of uncomplicated community-acquired bacterial urinary tract infections in adult patients. *Urologe* 2011;**50**:153–169.

Wagner BA, Dargatz DA, Morley PS, Keefe TJ, Salman MD. Analysis methods for evaluating bacterial antimicrobial resistance outcomes. *Am J Vet Res* 2003;**64**:1570–1579.

Wang F. Strategies for control of serious infections caused by multi-drug resistant *Pseudomonas* aeruginosa and Acinetobacter baumannii. Chin J Infect Chemother 2007;**7**:230–232.

Waterman P, Kwak Y, Clifford R, et al. A multidrug-resistance surveillance network: 1 year on. *Lancet Infect Dis* 2012;**12**:587–588.

Wunderink RG, Niederman MS. Update in respiratory infections 2011. *Am J Respir Crit Care Med* 2012;**185**:1261–1265.

Xu JF, Wu JF. Infection caused by multidrug resistant *Pseudomonas aeruginosa*. *Chin J Infect Chemother* 2007;**7**:141–144.

Yankaskas JR, Marshall BC, Sufian B, Simon RH, Rodman D. Cystic fibrosis adult care: consensus conference report. *Chest* 2004;**125**:1S–39S.

Yuan JY, Yang F. Latest advances on the diagnosis and management of ventilator-associated pneumonia. *Chin J Infect Chemother* 2007;**7**:382–385.

Zavascki AP. Treatment of multidrug-resistant *Pseudomonas aeruginosa* infections: more attention required to in-vitro studies. *Clin Microbiol Infect* 2005;**11**:856–857.

Zavascki AP, Li J. Intravenous colistimethate for multidrug-resistant Gram-negative bacteria. *Lancet Infect Dis* 2008;**8**:403–405.

Zavascki AP, Machado ABMP, de Oliveira KRP, *et al.* KPC-2-producing *Enterobacter cloacae* in two cities from Southern Brazil. *Int J Antimicrob Agents* 2009;**34**:286–288.

Retrospective cohort

Alexiou VG, Michalopoulos A, Makris GC, Peppas G, Samonis G, Falagas ME. Multi-drug-resistant Gram-negative bacterial infection in surgical patients hospitalized in the ICU: a cohort study. *Eur J Clin Microbiol Infect Dis* 2012;**31**:557–566.

Branski LK, Al-Mousawi A, Rivero H, Jeschke MG, Sanford AP, Herndon DN. Emerging infections in burns. *Surg Infect* 2009;**10**:389–397.

Cheng C, Tsai M, Huang Y, *et al.* Antibiotic resistance patterns of community-acquired urinary tract infections in children with vesicoureteral reflux receiving prophylactic antibiotic therapy. *Pediatrics* 2008;**122**:1212–1217.

Daniels TL, Deppen S, Arbogast PG, Griffin MR, Schaffner W, Talbot TR. Mortality rates associated with multidrug-resistant *Acinetobacter baumannii* infection in surgical intensive care units. *Infect Control Hosp Epidemiol* 2008;**29**:1080–1083.

das Neves MT, de Lorenzo MEP, Almeida RAMB, Fortaleza CMCB. Antimicrobial use and incidence of multidrug-resistant *Pseudomonas aeruginosa* in a teaching hospital: an ecological approach. *Rev Soc Brasil Med Trop* 2010;**43**:629–632.

Florent A, Chichmanian RM, Cua E, Pulcini C. Adverse events associated with intravenous fosfomycin. *Int J Antimicrob Agents* 2011;**37**:82–83.

Fortaleza CMCB, Figueiredo LC, Beraldo CC, De Melo EC, Pola PMS, Aragao VDN. Risk factors of oropharyngeal carriage of *Pseudomonas aeruginosa* among patients from a medical-surgical intensive care unit. *Braz J Infect Dis* 2009;**13**:173–176.

Geyik MF, Aldemir M, Hosoglu S, Tacyildiz HI. Epidemiology of burn unit infections in children. *Am J Infect Control* 2003;**31**:342–346.

Hachem RY, Chemaly RF, Ahmar CA, *et al.* Colistin is effective in treatment of infections caused by multidrug-resistant *Pseudomonas aeruginosa* in cancer patients. *Antimicrob Agents Chemother* 2007;**51**:1905–1911.

Hyle EP, Lipworth AD, Zaoutis TE, Nachamkin I, Bilker WB, Lautenbach E. Impact of inadequate initial antimicrobial therapy on mortality in infections due to extended-spectrum beta-lactamase-producing enterobacteriaceae: variability by site of infection. *Arch Intern Med* 2005:**165**:1375–1380.

Johansen HK, Moskowitz SM, Ciofu O, Pressler T, Hoiby N. Spread of colistin resistant non-mucoid *Pseudomonas aeruginosa* among chronically infected Danish cystic fibrosis patients. *J Cystic Fibrosis* 2008;**7**:391–397.

Johnson LE, D'Agata EMC, Paterson DL, *et al. Pseudomonas aeruginosa* bacteremia over a 10-year period: multidrug resistance and outcomes in transplant recipients. *Transplant Infect Dis* 2009;**11**:227–234.

Joung MK, Kwon KT, Kang CI, *et al.* Impact of inappropriate antimicrobial therapy on outcome in patients with hospital-acquired pneumonia caused by *Acinetobacter baumannii. J Infect* 2010;**61**:212–218.

Kasiakou SK, Michalopoulos A, Soteriades ES, Samonis G, Sermaides GJ, Falagas ME. Combination therapy with intravenous colistin for management of infections due to multidrug-resistant Gramnegative bacteria in patients without cystic fibrosis. *Antimicrob Agents Chemother* 2005;**49**:3136–3146.

Kent L, Bradley JM, France M, *et al.* Temocillin in cystic fibrosis: a retrospective pilot study. *J Cystic Fibrosis* 2008;**7**:551–554.

Leflon-Guibout V, Ternat G, Heym B, Nicolas-Chanoine M-H. Exposure to co-amoxiclav as a risk factor for co-amoxiclav-resistant *Escherichia coli* urinary tract infection. *J Antimicrob Chemother* 2002;**49**:367–371.

Le Hello S, Falcot V, Lacassin F, Mikulski M, Baumann F. Risk factors for carbapenem-resistant *Acinetobacter baumannii* infections at a tertiary care hospital in New Caledonia, South Pacific. *Scand J Infect Dis* 2010;**42**:821–826.

Lipovy B, Rihova H, Hanslianova M, Gregorova N, Suchanek I, Brychta P. Prevalence and resistance of *Pseudomonas aeruginosa* in severely burned patients: a 10-year retrospective study. *Acta Chirurg Plastic* 2010;**52**:39–43.

Medvedev DS, Zakharova NV. De-escalation strategy reduces Gram-negative pathogen resistance in infectious complications of thermal burns. *Clin Microbiol Infect* 2011;**17**:S774.

Navarro JL, Somodevilla A, Martinez MC, *et al.* Nosocomial outbreak of *Pseudomonas aeruginosa* in adult inpatients: multidrug- vs. non-multidrug-resistant strains. *Clin Microbiol Infect* 2011;**17**:S717.

Poulakou G, Kontopidou FV, Paramythiotou E, *et al.* Tigecycline in the treatment of infections from multi-drug resistant Gram-negative pathogens. *J Infect* 2009;**58**:273–284.

Prevotat A, Leroy S, Perez T, Wallet F, Wallaert B. Tolerance and efficacy of ceftazidime in combination with aztreonam for exacerbations of cystic fibrosis. *Rev Malad Respir* 2010;**27**:449–456.

Schwaber MJ, Navon-Venezia S, Kaye KS, Ben-Ami R, Schwartz D, Carmeli Y. Clinical and economic impact of bacteremia with extended-spectrum-beta- lactamase-producing Enterobacteriaceae. *Antimicrob Agents Chemother* 2006;**50**:1257–1262.

Sousa M, Trindade I, Cortesao N, Alves V, Granja C. Successful change in epidemiology of ICU multiresistant bacteria with implementation of evidence-based interventions. *Crit Care Med* 2010;**38**:A84.

Strenger V, Gschliesser T, Grisold A, *et al.* Orally administered colistin leads to colistin-resistant intestinal flora and fails to prevent faecal colonisation with extended-spectrum beta-lactamase-producing enterobacteria in hospitalised newborns. *Int J Antimicrob Agents* 2011;**37**:67–69.

Tasbakan MS, Pullukcu H, Sipahi OR, Tasbakan MI, Aydemir S, Bacakoglu F. Is tigecyclin a good choice in the treatment of multidrug-resistant *Acinetobacter baumannii* pneumonia? *J Chemother* 2011;**23**:345–349.

Trottier V, Namias N, Pust DG, *et al.* Outcomes of *Acinetobacter baumannii* infection in critically ill surgical patients. *Surg Infect* 2007;**8**:437–443.

Study design not relevant

Bercault N, Linassier P. The value of sepsis isolation to diminish the spread of multidrug-resistant bacteria in intensive care. Consequences on the incidence of nosocomial infections. *Rev Med Intern/ Fondee* 1999;**20**:86–87.

Drapeau CMJ, Grilli E, Petrosillo N. Rifampicin combined regimens for Gram-negative infections: data from the literature. *Int J Antimicrob Agents* 2010;**35**:39–44.

Eveillard M, Eb F, Tramie B, *et al.* Evaluation of the contribution of isolation precautions in prevention and control of multi-resistant bacteria in a teaching hospital. *J Hosp Infect* 2001;**47**:116–124.

Falagas ME, Kasiakou SK. Toxicity of polymyxins: a systematic review of the evidence from old and recent studies. *Crit Care* 2006;**10**:R27.

Furtado GHC, Martins ST, Machado AMO, Wey SB, Medeiros EAS. Clinical culture surveillance of carbapenem-resistant *Pseudomonas aeruginosa* and *Acinetobacter* species in a teaching hospital in São Paulo, Brazil: a 7-year study. *Infect Control Hosp Epidemiol* 2006;**27**:1270–1273.

Hashino S, Morita L, Kanamori H, *et al.* Clinical impact of cycling the administration of antibiotics for febrile neutropenia in Japanese patients with hematological malignancy. *Eur J Clin Microbiol Infect Dis* 2012;**31**:173–178.

Noy A, Orni-Wasserlauf R, Sorkine P, Siegman-Igra Y. Epidemiology of ceftazidime-resistant *Klebsiella pneumoniae* in a large university hospital in Tel Aviv. *Israel Med Assoc J* 2000;**2**:908–911.

Oostdijk EA, Leverstein-van Hall M, Muilwijk J, Kesecioglu J, Bonten MJ. Colistin resistance in Gramnegative bacteria during prophylactic colistin use in intensive care units. *Clin Microbiol Infect* 2011;**17**:S294.
Patterson JE, Hardin TC, Kelly CA, Garcia RC, Jorgensen JH. Association of antibiotic utilization measures and control of multiple-drug resistance in *Klebsiella pneumoniae*. *Infect Control Hosp Epidemiol* 2000;**21**:455–458.

Prescott WA Jr, Gentile AE, Nagel JL, Pettit RS. Continuous-infusion antipseudomonal beta-lactam therapy in patients with cystic fibrosis. *P* T 2011;**36**:723–763.

Schmitt F, Clermidi P, Dorsi M, Cocquerelle V, Gomes CF, Becmeur F. Bacterial studies of complicated appendicitis over a 20-year period and their impact on empirical antibiotic treatment. *J Pediatr Surg* 2012;**47**:2055–2062.

Controlled before-after studies without a minimum of two intervention and control sites

Bennett KM, Scarborough JE, Sharpe M, *et al.* Implementation of antibiotic rotation protocol improves antibiotic susceptibility profile in a surgical intensive care unit. *J Trauma* 2007;**63**:307–311.

De Champs CL, Guelon DP, Garnier RM, *et al.* Selective digestive decontamination by erythromycinbase in a polyvalent intensive care unit. *Intensive Care Med* 1993;**19**:191–196.

Martínez J, Nicolás J, Marco F, *et al.* Comparison of antimicrobial cycling and mixing strategies in two medical intensive care units. *Crit Care Med* 2006;**34**:329–336.

Interrupted time series studies without at least three data points before and after the intervention

Geissler A, Gerbeaux P, Granier I, Blanc P, Facon K, Durand-Gasselin J. Rational use of antibiotics in the intensive care unit: impact on microbial resistance and costs. *Intensive Care Med* 2003;**29**:49–54.

Participants not relevant

Giamarellou H, Efstratiou A, Tsagarakis J, Petrikkos G, Daikos GK. Experience with ciprofloxacin in vitro and in vivo. *Arzneimittel Forschung* 1984;**34**:1775–1778.

Karageorgopoulos DE, Kelesidis T, Kelesidis I, Falagas ME. Tigecycline for the treatment of multidrug-resistant (including carbapenem-resistant) *Acinetobacter* infections: a review of the scientific evidence. *J Antimicrob Chemother* 2008;**62**:45–55.

Kastoris A, Rafailidis P, Vouloumanou E, Gkegkes I, Falagas M. Synergy of fosfomycin with other antibiotics for Gram-positive and Gram-negative bacteria. *Eur J Clin Pharmacol* 2010;**66**:359–368.

Wistrom J, Gentry LO, Palmgren AC, *et al.* Ecological effects of short-term ciprofloxacin treatment of travellers' diarrhoea. *J Antimicrob Chemother* 1992;**30**:693–706.

Zervos M, Mandell LA, Vrooman PS, *et al.* Comparative efficacies and tolerabilities of intravenous azithromycin plus ceftriaxone and intravenous levofloxacin with step-down oral therapy for hospitalized patients with moderate to severe community-acquired pneumonia. *Treat Respir Med* 2004;**3**:329–336.

Antibiotics used not relevant for the review

Galanakis N, Giamarellou H, Moussas T, Dounis E. Chronic osteomyelitis caused by multi-resistant Gram-negative bacteria: evaluation of treatment with newer quinolones after prolonged follow-up. *J Antimicrob Chemother* 1997;**39**:241–246.

Giamarellou H, Tsagarakis J, Petrikkos G, Daikos GK. Norfloxacin versus cotrimoxazole in the treatment of lower urinary tract infections. *Eur J Clin Microbiol* 1983;**2**:266–269.

Goldstein EJ, Alpert ML, Najem A, *et al.* Norfloxacin in the treatment of complicated and uncomplicated urinary tract infections. A comparative multicenter trial. *Am J Med* 1987;**82**:65–69.

Henry Jr DC, Bettis RB, Riffer E, *et al.* Comparison of once-daily extended-release ciprofloxacin and conventional twice-daily ciprofloxacin for the treatment of uncomplicated urinary tract infection in women. *Clin Ther* 2002;**24**:2088–2104.

Holloway WJ, Palmer D. Clinical applications of a new parenteral antibiotic in the treatment of severe bacterial infections. *Am J Med* 1996;**100**:52S–59S.

Koomanachai P, Tiengrim S, Kiratisin P, Thamlikitkul V. Efficacy and safety of colistin (colistimethate sodium) for therapy of infections caused by multidrug-resistant *Pseudomonas aeruginosa* and *Acinetobacter baumannii* in Siriraj Hospital, Bangkok, Thailand. *Int J Infect Dis* 2007;**11**:402–406.

Vlieghe E, Phoba MF, Tamfun JJM, Jacobs J. Antibiotic resistance among bacterial pathogens in Central Africa: a review of the published literature between 1955 and 2008. *Int J Antimicrob Agents* 2009;**34**:295–303.

Not multi-drug-resistant infections

Hooton TM, Roberts PL, Stapleton AE. Cefpodoxime vs ciprofloxacin for short-course treatment of acute uncomplicated cystitis: a randomized trial. *JAMA* 2012;**307**:583–589.

Sole Violan J, Fernandez JA, Benitez AB, Cendrero JAC, De Castro FR. Impact of quantitative invasive diagnostic techniques in the management and outcome of mechanically ventilated patients with suspected pneumonia. *Crit Care Med* 2000;**28**:2737–2741.

Appendix E. Peer review

Healthcare Infection Society

Consultation – Joint Working Party on multi-drug resistant Gram-negative infections: prevention and control

Closing date: 5pm 31st October 2014

Organization	Ayrshire and Arran	
Title (e.g. Dr, Mr, Ms, Prof)	Dr	
Name	Alan McDonald	
Job title or role	Consultant Microbiologist	
Address and post code	Crosshouse Hospital	
	Kilmarnock	
Telephone number	01563 825285	
Email address	Alan.MacDonald@aaaht.scot.nhs.uk	
Please note: comments will only be accepted electronically on this proforma.		

Section		
	Comments	
Line 256	Delete word 'in'	Amended
Line 315	Amended	Amended
Line 318	I don't think this is necessary and could give false assurance.	Sentence
	See Scottish Guidance Version 2.0 July 2014. Guidance for neonatal units, adult and paediatric intensive care units in Scotland to minimise the risk of <i>Pseudomonas aeruginosa</i> infection from water.	
	Page 10 'Routine sampling of water to detect Pseudomonas aeruginosa should not be carried out.'	
Line 324	Do you mean 'selective decontamination of the digestive tract'?	Amended
Line 327	What is this?	Amended
Line 330	and nursing staff	Amended
Line 334	Why is this percentage sign at the end of the lines?	Amended

Consultation – Joint Working Party on multi-drug-resistant Gram-negative infections: prevention and control

Closing date: 5pm 31st October 2014

Organization	NHS Lanarkshire	
Title (e.g. Dr, Mr, Ms, Prof)	Mrs	
Name	Carol Whitefield	
Job title or role	Senior Infection Control Nurse	
Address and post code	Infection Control Department	
	Wishaw General Hospital	
Telephone number	01698 366453	
Email address	Carol.whitefield@lanarkshire.scot.nhs.uk	
Please note: comments will only be accepted electronically on this proforma.		

Section		
	Comments	
Line 256		Amended
	Remove word 'in'	
Line 292 and line 1501	Why gowns instead of aprons?	Amended
Line 297	Isolate those colonized. This can prove extremely difficult due to pressures for single rooms.	Amended
Line 310	ATP testing for monitoring the environment. Not widely used at present.	Amended
Line 320	Hydrogen peroxide vaporizers. Not many hospitals own these.	Not changed as hospitals can hire them
Line 688	Enterobacteriaceae. This word is repeated.	Amended

Consultation – Joint Working Party on multi-drug-resistant Gram-negative infections: prevention and control

Closing date: 5pm 31st October 2014

Organization	Health Protection Scotland	
Title (e.g. Dr, Mr, Ms, Prof)	Dr	
Name	Camilla Wiuff	
Job title or role	Strategic Lead Microbiology	
Address and post code	5 Cadogan Street	
	Glasgow G2 6QE	
Telephone number	0141 282 2927	
Email address	camilla.wiuff@nhs.net	
Please note: comments will only be accepted electronically on this proforma.		

Section		
	Comments	
General	The document presents a very thorough and systematic review of current evidence in the area of prevention and control of MDR Gram-negatives and provides the strength of the evidence and associated recommendations. It provides a solid basis for the development of guidance in this area. The initial review of the current epidemiology of a range of MDR Gram-negative organisms is also very helpful.	
Page 25	In the Recommendation: 'Screening for carbapenem-resistant organisms should be prioritized to patients admitted from ICU and from post-acute facilities' – please clarify if post-acute facilities includes long-term care facilities, and if this recommendation refers to all admissions to any ward from post-acute facilities.	Amended
Page 50	A Vietnamese study on patient isolation is mentioned (Schultsz, 2013) – but in the second sentence, it is stated that the study did not include patient isolation. Please clarify.	Amended

Consultation – Joint Working Party on multi-drug-resistant Gram-negative infections: prevention and control

Closing date: 5pm 31st October 2014

Organization	NHS Dumfries & Galloway	
Title (e.g. Dr, Mr, Ms, Prof)	Mrs	
Name	Elaine Ross	
Job title or role	Infection Control manager	
Address and post code	Infection Control Department	
	Residences	
	Dumfries & Galloway Royal Infirmary	
	Bankend Road	
	Dumfries DG1 4TG	
Telephone number	01387 241426	
Email address	Elaine.ross2@nhs.net	
Please note: comments will only be accepted electronically on this proforma.		

Section	
	Comments
General	Well-designed and presented review of the evidence.
	Will have significant implications for practice in terms of screening and isolation as expected.

Consultation – Joint Working Party on multi-drug-resistant Gram-negative infections: prevention and control

Organization	Tayside	
Title (e.g. Dr, Mr, Ms, Prof)	Dr	
Name	Gabby Phillips	
Job title or role	Consultant Microbiologist	
Address and post code		
Telephone number		
Email address	gabby.phillips@nhs.net	
Please note: comments will only be accepted electronically on this proforma.		

Closing date: 5pm 31st October 2014

Section	
	Comments
	Very comprehensive document

Consultation – Joint Working Party on multi-drug-resistant Gram-negative infections: prevention and control

Organization	HIS	
Title (e.g. Dr, Mr, Ms, Prof)	Dr	
Name	Alaric Colville	
Job title or role	Coordinator of responses to consultations for Healthcare infection Society	
Address and post code	Department of Microbiology	
	Royal Devon and Exeter NHS Foundation Trust	
	Church Lane	
	Exeter EX2 5AD	
Telephone number	01392 402961	
Email address	alaric.colville@nhs.net	
Please note: comments will only be accepted electronically on this proforma.		

Closing date: 5pm 31st October 2014

Section		
Controll	Comments	
HIS comments	The following are a summary of comments received from those who have responded through HIS.	
	All those who submitted comments were highly appreciative of the guidelines in general and welcomed their imminent appearance. Also recognized the huge amount of work undertaken.	
	The guidance is welcomed as a substantial improvement on the PHE guidance 'CPE Toolkit.	
General comments	 As someone trained on a diet of concern about gentamicin resistance, it concerns me greatly that there is no mention of aminoglycoside resistance in this document. Even quinolone resistance gets a brief mention but other than that, it is all carbapenemase, ESBLs, AmpC and Acinetobacters. If aminoglycoside resistance is no longer a concern, you should at least state why. Meanwhile, here on the ground, we will just monitor its inexorable rise in these days of avoiding broader spectrum agents! 	
	 Standard infection control precautions, measures, standard precautions – too many terms. Use one and define please. 	Amended
	3. The term 'high-risk patients' is used often in the document. Sometimes loosely. Is it high risk of having or acquiring CPE?	Defined in glossary
Section 4 Summary of Guidelines	The term 'post-acute care' is not a commonly used term and is ambiguous. When I read the recommendation in the summary at the beginning, I took it to mean 'admission from acute care elsewhere'. It was only on reading the body of the text that I see that you mean admission from rehab, nursing and	Amended

Section		
Section	Comments	
	residential facilities. Please clarify.	
	Line 343 Not just decontaminating respiratory equipment in handwash stations (not wash basin), it is not tipping respiratory secretions or ventilator exudates into handwash stations – these are to be used for handwashing only. See relevant Chief Executive Letters, Health Protection Scotland.	Amended
	Line 492 The terms 'plasmid outbreaks' and 'plasmid-related outbreaks' are both used – please use one.	Amended
	Line 647 There is a notable seasonality to all Gram-negative bacteraemias – which season?	Amended
	Line 704 For ESBL carriers is a risk factor for what – infection or colonization or both?	Amended
	Line 785 Needs a line after surveillance 'of what to detect what'.	Heading amended
Section 7.3.1 Testing of diagnostic samples	Line 840 Whilst recognizing the lack of specificity, is ertapenem a more convenient screening test for widespread screening for urine Enterobacteriaceae isolates than testing meropenem and cefpodoxime?	Not changed as counter to
	Line 870–871 Which means what for the UK?	Amended
	Line 891 Note no limit of time here, no risk assessment of degree of exposure or amount of organism on the patient. Are we sure?	Amended
Section 7.3.3 How should we undertake local surveillance, why is it important	Line 940 'Passive surveillance is not recommended when outbreaks are anticipated and clinical risk is high.'	Amended
and how should it be interpreted?	This recommendation doesn't make sense. Even after reading the section, I'm unclear as to what is being recommended — except when outbreaks are anticipated, or not at all?	

Section		
	Comments	
Section 7.4.2.2 What organisms should screening include?	Line 1130 This is too non-specific re: <i>Pseudomonas</i> are often resistant to carbapenems and I would not want to necessarily be chasing these – perhaps specify VIM/IMP-resistant strains.	
Section 7.4.2.3.1 Whom to screen	Line 1134 Until such times as PHE are prepared to allow us mere mortals on the front line access to up-to- date UK epidemiology, there is no point in recommending screening from UK institutions with a high prevalence (line 1163). How about an additional recommendation to make UK epidemiology available to us all?	Probably outside remit
Section 7.4.3.2 Disposable aprons and gloves	Line 1501 Comment: The evidence supplied does not suggest gowns over aprons, although from a practical perspective, gowns may be preferable on occasions (close body contact with patient), but are we suggesting entering the room wearing gowns?	Changed all to say apron
Section 7.4.2.1 What is the role of screening in patients and staff?	Line 1084–1091 You provide no evidence to back up the statement that cross-transmission is by members of staff via hands with the statements below. In fact, the evidence presented in this paragraph does not support the assertion.	Sentence deleted
	Line 1093 Does not make sense This has to be combined with the full implementation of standard infection control precautions throughout the care area.	Amended
Section 7.4.2.3.2 How to screen	Line 1179 In practice, staff don't like doing rectal swabs and patients don't like having them done. We find that if it is possible to visualize faeces on the swab, then you don't have long to wait for a stool sample so why not just request a swab from a stool sample unless in an outbreak situation. Don't request a stool sample or it will inevitable get diverted to the stool bench.	Amended
Section 7.4.5.1 When should the environment be	Line 1742 Comment: There is not enough guidance about how to screen and the sensitivity of screening. I would say 'consider screening'.	Amended recommendatio

Section	Comments	
sampled?		
	Line 1742 Comment: It doesn't seem to me that the evidence supplied suggests this is 'strong' evidence outside of <i>Acinetobacter</i> spp. The major problem at present is carbapenemase-producing Enterobacteriaceae rather than <i>Acinetobacter</i> spp., and to place great relevance on environmental screening is not correct or helpful practically.	As above
	Line 1287 and further lines elsewhere. If they are in isolation, transmission-based precautions are being used. Everyone is applying SICPs for all patients all of the time.	Amended
Section 7.4.2.6 Is there evidence for effective interventions on positive patients i.e. can they be cleared?	Line 1345 YOU NEVER DISCONTINUE SICP! This paragraph needs rewriting.	Contact precautions introc

Consultation – Joint Working Party on multi-drug resistant Gram-negative infections: prevention and control

Closing date: 5pm 31st October 2014

Organization	MRSA Action UK	
Title (e.g. Dr, Mr, Ms, Prof)	Mr	
Name	Derek Butler	
Job title or role	Chair	
Address and post code	6 Lunesdale Road	
	Kirkham PR4 2HS	
Telephone number	07762 741114	
Email address	derek.butler6@btinternet.com	
Please note: comments will only be accepted electronically on this proforma.		

Section		
Occion	Comments	
General	MRSA Action UK welcomes this guidance. We would like to see a plain English guide for patients with leaflets and publicity to raise awareness of this important and significant issue. There is patient acceptability with regard to MRSA screening in most instances, and this is largely attributable to the well-publicised information on MRSA and the interventions that have been put in place to prevent it.	Leaflets are planned
	Information about the need to diagnose MDR Gram-negative bacteria in high risk patients and the measures needed to deal with it should, we believe, be well publicised.	
Summary of guidelines	Establish a robust flagging system for patient notes. Change 'weak' to 'strong'.	Weak relates to the strer
Line 301	Rationale:	
	Information on carriage or infection is important, particularly when transferring patients between facilities. This information is essential in identifying high-risk patients for screening.	
Summary of guidelines	In areas where numbers justify, consider a separate dedicated nursing unit and monitoring hand hygiene of shared medical staff.	
Line 328–329	Change 'weak' to 'strong'.	
	Rationale:	
	Monitoring of hand hygiene of medical staff at the point of care is important, particularly in an outbreak with	

Section	Comments	
	these significant bacteria. This matches the 'strong' recommendation 'To prevent any hospital-acquired infection, hand hygiene is required before and after direct patient contact, after contact with body fluids, mucous membranes and non-intact skin, after contact with the immediate patient environment and immediately after the removal of gloves.' in line 302–305.	

Healthcare Infection Society

Consultation – Joint Working Party on multi-drug-resistant Gram-negative infections: prevention and control

Closing date: 5pm 31st October 2014

Organization	PHE & Associated Working Groups
Title (e.g. Dr, Mr, Ms, Prof)	Mrs
Name	Liz Stokle
Job title or role	AMRS & HCAI Programme Lead
Address and post code	Public Health England
	Wellington House

	133–155 Waterloo Road	
	London, SE1 8UG	
Telephone number	020 78117240 Blackberry: 07733306918	
Email address	Liz.stokle@phe.gov.uk	
Please note: comments will only be accepted electronically on this proforma.		

Section	Comments	
General comments	With the current focus on carbapenemase-producing Enterobacteriaceae and, given this guidance includes other MDR organisms (e.g. ESBL, AmpC producers), it will be important to emphasize that prevention of transmission for any of these organisms should be paramount.	
	In relation to lay/patient representation, it may be that the lay summary would sit better below the executive summary. An introductory statement would help to set the context, describing what the report sets out to do and group representatives, including patient representatives. Sentence two of the summary could be clearer.	Amended
	The words bacteraemia/bacteremia and colonised/colonized have been used	

Section		
	Comments	
	interchangeably throughout. Generally, recommendations need to be more prescriptive and consistent with that highlighted in the narrative.	Corrected
Specific line comments		
Section 4: Line 326: Code of Practice HPA 2013	Should this be the Code of Practice which is part of the Health and Social Care Act or the HPA/DH 'Prevention and control of infection in care homes – an information resource'?	HPA 2013 but as va
Section 5: Line 335: Audit measure 'All patients infected with meropenem-resistant Gram-negative bacteria to be reported to Public Health England or equivalent bodies'	It is unclear whether this should include colonization; reporting to PHE centres or equivalent.	Amended
Section 7: Line 423: 'healthcare-acquired infections often become apparent after hospital discharge'	It should be noted that healthcare -acquired infections could denote health care in a hospital or the community – it is assumed hospital-acquired infection is what is implied.	Amended
	In assessing community vs hospital-acquired infection, it is not clear, by considering hospitalization in the last year, whether this will elucidate place of acquisition (considering the difficulty determining chronology) in revolving door patients; it would be useful to reference this.	Referenced

Section	Comments	
Section 7.1.4.2 Line 495: 'This is found, for example, in the current spread of pKpQIL plasmids encoding KPC carbapenemases in and around Manchester.'	This is a working assumption but is not proven.	Amended
7.1.4.4 Line 531 'Long-term care facilities range from establishments offering assisted-living to largely independent residents through to those providing complex medical support (CDC 2014, Lievesley 2011)'	Do you mean the Centre for Policy on Ageing rather than CDC?	CDC is correct
Section 7.1.5 Line 604 ' numbers'	Typographical error.	Corrected
Section 7.2 Line 682 Recommendation; 'Screening for carbapenem-resistant organisms should be prioritized to patients admitted to ICU and from post- acute care facilities.'	The evidence does not directly support this recommendation. The key issue is whether we are trying to prevent infection (and thus focus on critical care may be appropriate) or transmission (and thus we need to target all at-risk patients). This is inconsistent with the Acute Toolkit. In Greater Manchester, risk is more associated with previous hospital admissions than post-acute care facilities.	Amended
Section 7.2.1 Line 688 Enterobacteriaceae repeated	Typographical error.	Corrected
Line 694 'common environmental sources have occasionally been described and should be sought where no other plausible vectors can be found'	It is unclear what this means without identifying common sources/supporting with evidence.	Amended
Section 7.2.1 Line 711–716 'Screening for carriers with subsequent isolation of those identified is effective in preventing transmission and is important for early recognition of individuals at high risk of carriage of carbapenem-resistant Enterobacteriaceae	Should there be recommendations associated with this statement?	Background informa

Section		
	Comments	
Awareness of carriage is important and, therefore,		
communications regarding patients who are known to		
be infected or colonized with MDR strains is essential		
when transferring patients within and between		
institutions.'		
Section 7.3.2 Line 876–877 Recommendation	It would be useful to have a recommendation about outputs of recommended	Amended
Antimicrobial susceptibility data on all routine isolates	surveillance in addition to inputs. The guidance makes recommendations for	
should be reported electronically to a central national	reporting of sensitivities on all significant isolates to PHE, but makes no	
database, preferably from all body sites.	comments on what output is required. Analysis of data by acute trust is required.	
	Mandatory reporting is an approach supported by some as a way of ensuring that	
	all trusts report.	
Section 7.3.3 Line 880–881 How should we	It would be helpful to clarify what is meant by local – is this trust/laboratory level,	Amended
undertake local surveillance, why is it important and	or local authority level?	
how should it be interpreted?		
Line 882: Where warranted to track resistance	Is this the only reason for screening? It will also inform and evaluate infection	Amended
	prevention and control practices.	Amenueu
types (e.g. carbapenemase producers), local		
screening should be performed'		
Line 886: 'it is critical to ensure the compliance of	It is not clear whether this means compliance with guidance indicating when to	Amended
staff taking the samples by means of audit and	take samples or how to take samples or other.	
feedback'		
Line 898–899 Local surveillance provides more rapid	The reference laboratory does not actually undertake surveillance. A definition of	Amended
notification of an emergent problem than the	local surveillance is required. The presumption is that this is trust level	

Section		
	Comments	
reference laboratory surveillance, particularly if a single clone and species is responsible	surveillance.	
Line 910–913: Screening on admission and weekly until discharge should be performed on patients at risk, known to be colonized or their nearby contacts as part of a package of measures to control an outbreak	If a patient is known to be colonized on admission, it is unclear why they should be screened on admission. The meaning of the wording 'known to be colonized or their nearby contacts' is unclear.	Amended
Section 7.3.4 Line 926	Appears to have lost text as next sentence appears to be a 'follow-on' rather than the start of a topic.	Amended
Line 939 Recommendation: 'Passive surveillance is not recommended when outbreaks are anticipated and clinical risk is high'	It is unclear what is meant by 'when outbreaks are anticipated'; by their very nature, outbreaks are not expected.	Amended
Section 7.4.1: Line 966–967: 'of hospital patients without the clear identification of such movements to the laboratory'	This line appears muddled/misplaced.	Amended
Line 986: 'Public Health England, Centers for Disease Control, ESCMID all recommend contact precautions'	PHE toolkit recommends standard precautions not contact precautions.	Amended
Line 993: 'emphases'	Typographical error.	Amended
Section 7.4.1 Line 1030–1031 'Assess all patients for infection risk on arrival at the care area (if possible, prior to accepting a patient from another healthcare area) and continuously review patient infection status throughout their stay.'	Infection risk or colonization risk? Or should it be transmission risk?	Amended

Section		
	Comments	
Line 1039–1040 Do not discard body fluids, secretions or exudates into handwash basins	Should this also include water used to wash patients or the environment?	Amended
Section 7.4.2 Line 1103: 'Screening of potential carriage sites in patients should be undertaken as part of a package of infection control measures for carbapenemase-producing Enterobacteriaceae to prevent the spread of outbreak strains'	This should clarify which patients.	At risk, addressed la
Line 1169 'Given the likelihood of prolonged gastrointestinal carriage of MDR Gram-negative organisms, clearance samples are not recommended'	As this is termed as 'likelihood' rather than clearly evidenced, would clearance samples provide valuable information about the patient's status at the same time as increasing our knowledge about carriage duration?	Amended. The table
Section 7.4.2.3.3 Line 1266 'Effective communications between healthcare settings will help facilitate efficient patient transfers'	As this is a crucial measure in reducing spread, could a recommendation be associated with this?	Amended
Section 7.4.2.4 Line 1272 'In situations where a patient is incapacitated and cannot sign, it may be considered permissible for those giving care to proceed with any interventions'	Suggest strengthen from 'may be permissible' to 'those giving care may proceed'.	Amended
Line 1281: 'Patients should be informed, whenever possible, of the need and reason for screening (i.e. that it is for their benefit)'	To some extent, this is contradictory to line 882 re tracking resistance types.	No change

Section	Comments	
Line 1283: 'They should be given the option of who carries it out, including self-screening if the patient is able and prefers it'	Depending on the patient, self-screening with a rectal swab may not be safe or realistic.	Amended
Section 7.4.2.6 Line 1345: 'SICP'	This needs to be spelled out.	Amended
Line 1348: 'organizations should be cautious in discontinuing contact precautions'	Advice appears to switch between standard and contact precautions.	Amended
Line 1377–1379 'Local screening policies should be developed to define those patients at high risk of carriage of, for example, carbapenemase producers. All patients transferred from healthcare facilities with endemic carbapenemase-producing Enterobacteriaceae at home or abroad should be screened'	Consider adding 'or with a history of admission to'.	Amended
Line 1386–1387 Patients colonized with carbapenem- resistant organisms should be isolated for the duration of their stay where possible	It needs to be clear what the implications are for future stays.	Amended
Section 7.4.5.3 Line 1835 'Water sources should be sampled at least twice a year for <i>P. aeruginosa</i> in augmented care units and point-of-use filters installed or taps changed when levels of patient colonization or infections rise.'	The group recommend twice-yearly testing of water in augmented care units for <i>Pseudomonas</i> spp. This is in line with national guidelines, but it is even more important to stress the requirement for a full risk assessment in relation to these guidelines. For example, removal of automatic taps, removal of thermostatic mixing valves, removal of flow straighteners, design of sinks etc. The Health Technical Memorandum does not specifically advise frequency of testing. There is no evidence to support twice-yearly testing.	Amended

Section	Comments	
	The group recommend the use of filters. These may be of short-term use whilst engineering solutions are implemented, but the <i>Pseudomonas</i> advisory group also found evidence that the filters themselves can become a source of <i>Pseudomonas</i> spp. because of where they are fitted. There is also some evidence that these filters actually deflect the <i>Pseudomonas</i> spp. (and <i>Legionella</i> spp.) to other parts of the pipework. Further data can be obtained from the PHE team at Porton Down.	
Table 4: 'Carbapenem-resistant Enterobacteriaceae – screen all patient contacts in ward of case who has not been identified and isolated'	Advice in the table in areas such as this are confusing/do not make sense.	Amended

Consultation – Joint Working Party on multi-drug-resistant Gram-negative infections: prevention and control

Closing date: 5pm 31st October 2014

Organization	Public Health Wales
Title (e.g. Dr, Mr, Ms, Prof)	Dr
Name	Neil Wigglesworth
Job title or role	Nurse Consultant
Address and post code	Temple of Peace and Health
	Cathays Park
	Cardiff CF10 3 NW
Telephone number	029 2040 2473
Email address	neil.wigglesworth@wales.nhs.uk
Please note: comments will only be accepted electronically on this proforma.	

Section		
	Comments	
Numerous (1)	There are a number of confusing references to terms to describe different levels of IPC precautions; when discussing managing cases, the terms 'isolation', 'contact precautions' and on one occasion at least 'standard infection control precautions' are used seemingly interchangeably. The guidance also refers to long-sleeved gowns without a discussion (as these do not routinely form part of contact precautions, a discussion is merited). Examples given below (some not all, there are many).	Amended
Numerous (2)	There are a number of England only references to regulation and regulatory structures – is this an England only document? Examples given below.	Amended
Section 7.3.4	When are the situations when outbreaks can be 'anticipated'?	Amended
Section 7.4.2.4	Example of IPC precautions terminology: ' isolation with standard infection control precautions'.	Amended
Section 7.4.2.6	As above: 'criteria for discontinuing SICP' (SICP should never be discontinued).	Amended
" "	As above: it then goes on to refer to contact precautions.	Amended
((As above: term used is isolated (nothing wrong with that but no consistency of language).	Amended
Section 7.4.5.4	Example of England only: the NHS constitution is England only I believe.	Amended
Section 7.5	As above: the Code of Practice and Care Quality Commission are England only.	Amended
ί (As above: the recommendation is England only.	Amended

Section	Comments	

Healthcare Infection Society

Consultation – Joint Working Party on multi-drug-resistant Gram-negative infections: prevention and control

Closing date: 5pm 31st October 2014

Organization	Southern Health and Social Care Trust
Title (e.g. Dr, Mr, Ms, Prof)	Dr
Name	Martin Brown
Job title or role	Consultant Microbiologist
Address and post code	Microbiology Department
	Craigavon Area Hospital
	68 Lurgan Road
	Portadown BT63 5QQ
Telephone number	02838612654
Email address	martin.brown@southerntrust.hscni.net

Please note: comments will only be accepted electronically on this proforma.

Section	Comments	
Section 4 Line 253– 255	 'Screening on admission and weekly until discharge should be performed on patients at risk, known to be colonized or their nearby contacts as part of a package of measures to control an outbreak' Please could it be clarified with the working group as to what the goal of screening patients known to be colonized with MDR Gram negatives is. The use of screening with regards to contacts and those at risk is clear; however, we would view patients known to be colonized as colonized and isolate them and take infection control precautions for them as a matter of course. As we have no method of decolonizing them, and the sensitivity of the screening test is not absolute, we would not view a negative result as one that would allow us to relax precautions. If the guidance pertains to trying to get specimens to link them to a potential outbreak, I can see the use in this, but otherwise screening known carriers will likely utilize resources to generate results that do not alter management. 	Amended
Section 4 Line 256– 257 and Section 7.3 Line 940–941	'Passive surveillance is not recommended in when outbreaks are anticipated and clinical risk is high' 'in when' is likely a typo	Amended
Section 7.18 Line 682–683	 'Screening for carbapenem-resistant organisms should be prioritized to patients admitted to ICU and from post-acute care facilities' Please could it be clarified if it is recommended that all patients admitted to ICU and from post-acute care facilities should be screened, and if so, whether they should all be screened weekly until discharge. I found this 	Amended

Section		
	Comments	
	a bit unclear.	
Section 7.3 Line 874– 875	'Laboratories should test meropenem susceptibility in all clinically significant Gram-negative isolates if possible and blood isolates as a minimum'	Amended – currently
	It lies beyond the capacity of many laboratories to test meropenem in all Gram-negative isolates, especially urine cultures and sputum cultures which are high volume and where the significance is not always known. It might be better to focus this on resistant organisms or potentially on sterile site cultures. I do appreciate though that it is aspirational and only says blood cultures as a minimum.	

Healthcare Infection Society

Consultation – Joint Working Party on multi-drug-resistant Gram-negative infections: prevention and control

Closing date: 5pm 31st October 2014

Organization	European Society of Clinical Microbiology and Infectious Diseases (ESCMID)	
Title (e.g. Dr, Mr, Ms, Prof)	Prof Doz. Dr	
Name	Jesus Rodriguez-Bano	Markus Hell
Job title or role	ESCMID Education Officer	ESCMID Study Group for Nosocomial Infections (ESGNI) Secretary
For further questions, please contact: Dr Henri Saenz, ESCMID Science Manager		

Address and post code	ESCMID Executive Office
	P.O. Box 214
	4010 Basel
	Switzerland
Telephone number	+41 61 508 0156
Email address	henri.saenz@escmid.org
Please note: comments will only be accepted electronically on this proforma.	

Section				
	Comments			
Page 15 Line 397	ESCMID, not ECCMID	Amended		
Page 17 Line 448	Diarrhoea and incontinence maybe	Amended		
Page 29 Line 813	Please allow us to ask why not use ertapenem as a first-step screening to detect carbapenemases, as it is the most sensitive carbapenem	Amended		
Page 51 Line 1501	We would like to point out that glove use is not advised systematically by some national infection control societies (e.g. France), as it is associated with decreased hand hygiene	Noted		

Section		
	Comments	
Page 54, Line 1616	In Section 7.4.4, we would suggest to reconsider 'equality of liquid soap and water WITH alcoholic hand rub', we think alcoholic hand disinfection should rather be favoured and handwashing be restricted to visibly soiled hands only (otherwise it may be confusing/unclear and could be inefficient in breaking the chain of transmission of all Gram-negative bacteria).	Amended
Section 7.4.5	Environmental cleaning could address surface disinfection techniques of the surfaces adjacent to the patient and sanitary cell.	Discussed later
General	May we suggest to add a section/subsection about reprocessing bed pans (automated bedpan reprocessing is a crucial issue for at least all MDR Enterobacteriaceae and <i>Pseudomonas</i> spp.) and maybe also about reprocessing flexible colonoscopies.	Query added to text
General	Clarification of the specific organisms: the ESCMID guidelines and the point about ESBL <i>E. coli</i> not to target anymore is mentioned, but we do not see this mirrored in the recommendation; we would suggest that for ESBL <i>E. coli</i> standard precautions are sufficient except there is evidence for a so-called high-risk clone/superspreader of ESBL <i>E. coli</i> in an institution or within a region; in addition, the high potential of transmission of <i>Klebsiella</i> spp. (ESBL + KPC) may be stressed more strongly.	Sentence added
General	We would like to suggest to harmonize the definition of 'multi-drug-resistant' with the one recommended internationally by ECDC and CDC, amongst others (Magiorakos, Clinical Microbiology & Infection).	Definition kept cons
General	Can we suggest that detection of ESBL producers focuses on <i>K. pneumoniae</i> , <i>Enterobacter</i> spp., <i>Serratia</i> spp. (and not <i>E. coli</i>). Tacconelli E, Cataldo MA, Dancer SJ, <i>et al.</i> ; European Society of Clinical Microbiology. ESCMID guidelines for the management of the infection control measures to reduce transmission of multidrug-resistant Gram-negative bacteria in hospitalized patients. <i>Clin Microbiol Infect</i> 2014; 20 (Suppl. 1):1–55.	Amended
General	ESGARS: The use of rapid diagnostic tests for ESBL and carbapenemases (Carba NP, ESBL NDP, MALDI-TOF application) may be considered.	Added for considera

Section	Comments	
	Nordmann P, Poirel L. The difficult-to-control spread of carbapenemase producers among Enterobacteriaceae worldwide. Clin Microbiol Infect 2014;20:821–830.	
	Burckhardt I, Zimmermann S. Using matrix-assisted laser desorption ionization-time of flight mass spectrometry to detect carbapenem resistance within 1 to 2.5 hours. <i>J Clin Microbiol</i> 2011; 49 :3321–3324.	
	Nordmann P, Poirel L, Dortet L. Rapid detection of carbapenemase-producing Enterobacteriaceae. <i>Emerg Infect Dis</i> 2012; 18 :1503–1507.	
	Nordmann P, Dortet L, Poirel L. Rapid detection of extended-spectrum-β-lactamase-producing Enterobacteriaceae. <i>J Clin Microbiol</i> 2012; 50 :3016–3022.	
Healthcare Infection Society

Consultation – Joint Working Party on multi-drug-resistant Gram-negative infections: prevention and control

Closing date: 5pm 31st October 2014

Organization	Infection Prevention Society	
Title (e.g. Dr, Mr, Ms, Prof)	Mrs	
Name	Debbie Wright	
Job title or role	IPS Honorary Secretary	
Address and post code	Blackburn House	
	Redhouse Road	
	Seafield	
	Bathgate EH47 &AQ	
Telephone number	01506 292 023 (Lynne Duncan, PA to the IPS Board)	
Email address	pa@ips.uk.net	
Please note: comments will	Please note: comments will only be accepted electronically on this proforma.	

Please provide comments on the draft guideline on the form below, putting each new comment in a new row. When feeding back, please note the section you are commenting on (for example, Section 1 Introduction and line number). If your comment relates to the guideline as a whole, please put 'general'. Add extra rows if required.

Section	Comments	
General	The recommendations for different MDR Gram-negative bacteria need to be separated because the infection control precautions for ESBLs, <i>Pseudomonas</i> spp. and carbapenamase-resistant organisms are very different. Combining them make the document confusing and it is difficult to easily see what precautions are relevant for different MDR Gram-negative bacteria.	Amended
General	Not all the recommendations are clearly mapped to the evidence presented, and it is therefore difficult to see how or why they are justified. Table IV would be improved by separating out the different organisms (see Point 1) and indicting the grade of evidence – not clear why this is there for the pseudomonas column but not the others.	Amended
General	The critical appraisal of evidence derived from outbreak studies or other poor-quality studies is minimal and there is a danger that what is presented reflects 'opinion' rather than robust evidence. Whilst the former may be reasonable, it is important to distinguish where recommendations are based on high-quality evidence and give some indication of the, often strong, biases evident in outbreak studies. Where the recommendation is based on expert opinion, the measure of certainty that they are effective and balance between desirable/undesirable effects should be clear.	Amended
General	Maybe it is intended to include as an appendix, but it would be helpful to see the evidence tables to understand how the recommendations have been supported by evidence. As many of the studies included are outbreak, it is difficult to see how these could be described as 2+. Many other recommendations including strong recommendations are not linked to a level of evidence.	Amended. The systematic review found only
General	GRADE recommends that recommendations are 'specific' and 'actionable'; not all of those included meet these criteria. GRADE is not advised for use with 'good practice recommendations' may be the problem with the approach taken here. It is also not advised to attempt to grade obvious procedures or standard practice. The list of	Amended

Section		
	Comments	
	recommendations may be easier to follow if they are divided by subheadings into areas of practice, e.g. screening, infection control procedures, management of outbreaks etc., and main groups of MDR Gram-negative bacteria.	
General	Previous high-quality evidence reviews should have been used to either support recommendations or to refer to, rather than attempt to undertake a superficial review that comes up with incomplete recommendations. In particular, this relates to aspects of practice covered by EPIC 3 (Loveday <i>et al.</i> , 2014) and by the systematic review of <i>Pseudomonas</i> spp. in healthcare water systems (Loveday <i>et al.</i> , 2014).	Amended
General	The terminology of 'screening', active and passive surveillance is confusing and the terms seem to be used interchangeably.	Amended
General	Target audience should be clarified. One mention of care homes at end of document – rest not relevant to this setting and care home guidance is not detailed.	Disagree care homes mentioned several tin
General	 There are a number of confusing references to different levels of IPC precautions; when discussing managing cases, the terms 'isolation', 'contact precautions' and on one occasion at least 'standard infection control precautions' are used seemingly interchangeably. The guidance also refers to long-sleeved gowns without a discussion (as these do not routinely form part of contact precautions, a discussion is merited). Examples given below (some not all, there are many). NOTE: I have not checked the tables for this issue. 	Removed long sleeve reference, rechecked
General	There are a number of England only references to regulation and regulatory structures – is this an England only document? Examples given below.	Amended

Section		
	Comments	
Line 227–232	There are costs associated with extended screening/surveillance, isolation and cohorting	Amended
	patients. This includes adverse effects to the patient of being put in prolonged isolation.	
	The efficacy of some of the proposed measures is, at best, uncertain and therefore it	'''''''''''''''''''''''''''''''''''''
	cannot be assumed that the costs will be offset by reduced transmission.	
Line 241	Does this mean you should or shouldn't screen these patients? It seems to conflict with	Amended
	Page 11 Line 253.	
Line 248	This is not a statement based on evidence; more a desired practice.	Removed
Line 251	What travel information and why/what should be done with it?	Amended
Line 256	This doesn't make sense and anyway is a double negative which in combination with a	Amended
	'weak' recommendation makes it difficult to understand what the requirement is.	
Line 258	This is not specific. Assess infection risk against what standards? What does 'infection	Amended
	status' mean and what action would be taken if it changed? Does it mean any infection or	
	an MDR Gram-negative bacteria infection?	
Line 261	Not specific.	Amended
Line 263, 265	These would be better cited as good practice recommendations and referenced to EPIC 3	Changed
	which fully reviewed the evidence underpinning them.	
Line 267	Whilst this may be 'guidance', it is based on extremely skimpy evidence. Again, suggest it	Disagree this is evidence based but moved
	should be listed as a good practice recommendation.	
Line 269	This is not specific or actionable. What all patients? Which body sites? What package of	Amended
	measures? How is an outbreak strain defined?	

Section		
	Comments	
Line 272	Not specific or actionable. Either screening is recommended or it is not. If it is going to be a recommendation, it should be 'preferred', preferred to what?	Amended
Line 282	This is two separate recommendations, although actually the first one is more of a 'good practice'.	Split
Line 286	Not specific or actionable. What does vigorously reinforced mean?	Amended
Line 289	Says the same thing twice.	Amended
Line 291	Why gowns? This is a very US approach. In the UK, we use aprons. There may be evidence that gowns might be necessary for abacters, but is there any evidence to suggest that they have a significant effect on transmission of other Gram-negative bacteria?	Gowns or aprons
Line 295	See earlier reference to EPIC 3.	Amended
Line 299	The priority is not just about MDR Gram-negative bacteria, the decision will depend on other infections too (<i>Clostridium difficile</i> infection, meticillin-resistant <i>Staphylococcus aureus</i> , tuberculosis etc.). Other recommendations are conflicting (i.e must isolate carbapenem-resistant Enterobacteriaceae but for ESBL isolate if possible). Cohort isolation should be a separate recommendation.	Amended. Not conflicting in that CRO highe
Line 303	See earlier reference to EPIC 3.	Amended
Line 307	Not specific.	Disagree
Line 310	There is no good evidence that monitoring based on ATP is effective in improving cleaning, and no evidence that it prevents transmission (which is presumably the outcome of interest for these recommendations).	Changed to 'weak'

Section		
	Comments	
Line 313	This is a good practice recommendation but actually should apply to any equipment, not just respiratory.	Amended
Line 315	What is the reason/evidence for this?	Removed as a recommendation
Line 317	This reflects (some of) the guidance on pseudomonas control – better to refer to other sources of guidance rather than partially repeat here. Evidence base is anyway minimal.	Amended
Line 320	What about cleaning with other disinfectants? There should be a more general recommendation – terminal cleaning after a case? Routinely for cleaning of area with infected patients? Or just for outbreaks? Is there evidence? Please note – there are costs associated with environmental damage of routine use of chlorine. It is frequently not feasible/practical to use H_2O_2 so important to recommend alternatives.	Added
Line 323	Think this means gut decontamination.	Amended
Line 325	Not specific – it is recommended for ICU patients to prevent VAP (and also for other patients at risk of HAP).	Removed
Line 326	Not specific or actionable.	Amended
Line 327	Is this for all patients or just those with MDR Gram-negative bacteria?	Amended
Line 329	Not specific or actionable. Monitoring hand hygiene a separate recommendation – why just medical staff?	Amended
Line 333	Isolates from blood cultures.	Amended
Line 334	What does significant Gram-negative isolates mean?	Amended

Section		
	Comments	
Line 335	This is not really an audit measure	Removed
Line 337	This is not really an audit measure	Removed
Line 343	This would be very difficult to audit	Removed
Line 441	References for this statement?	Referenced
Line 442	Vague and not referenced	Removed
Line 451	Are generally more likely to be associated with	Amended
Line 505	References?	Not required
Line 511	Reference?	Added Villegas
Line 512–521	There is a comprehensive systematic review of evidence for risks and control of <i>P. aeruginosa</i> related to water systems (Loveday <i>et al.</i> , 2014). Breathnach does not provide high-quality evidence (outbreak report), but there are others studies that provide better evidence to support control measure and risk factors.	Amended
Line 682	Does this mean you should or should not do it? Or should other patients be prioritized?	Amended
Line 700	This is fine but is not clearly reflected in the recommendations. Statement is not linked to evidence.	Deleted
Line 714	Reference?	Added
Line 748	What is the evidence for risk of resistance to chlorhex?	Amended
Line 759	There is no robust evidence to support this statement.	Amended

Section		
	Comments	
Line 760–767	This paragraph has not included critical appraisal of the evidence cited. See Loveday <i>et al.</i> for robust assessment of the quality of evidence of routes of transmission.	Added
Line 783	These are very old references.	Nil new
Line 871	Reference for this?	In section
Line 880	See previous comment about terms 'screening/surveillance'.	Amended
Line 883	No such section.	Amended
Line 912	At risk of what?	Amended
Line 925	References?	Added
Line 940	When are the situations when outbreaks can be 'anticipated'?	Removed
Line 987	Suggest define what is meant by contact precautions to avoid confusion and reference it (e.g. Seigel, 2007).	Amended
Line 1003	These refer to a combination of Seigel 2007 and EPIC (which does not include all these elements). Patient placement should be explained.	Referred to Siegel 2007
Line 1019–1021	Is this level of protection relevant to Gram-negative bacteria? If so, in which circumstances? Evidence/references?	Deleted
Line 1027	Discussion of the evidence suggests that contact precautions are required. These are different to the standard precautions recommended here. It may be necessary to recommend different things for different MDR Gram-negative bacteria.	Amended

Section		
	Comments	
Line 1030	These recommendations are not specific, especially in the absence of recommendations on how to decide whether a patient is an 'infection risk'.	Amended
Line 1066	This refers to transmission-based precautions not contact precautions – which and when?	Amended
Line 1093	Not sure what this means. Low-grade evidence.	Amended
Line 1135	This implies screening of the majority of patients in hospital.	Amended
Line 1287	Example of IPC precautions terminology: ' isolation with standard infection control precautions'.	Removed
Line 1391–1400	 There are no references or critical appraisal of evidence to support these statements or indication of situations where cohorting may be indicated. Conflicts with other statement about isolation not being practical for ESBLs. Example of IPC precautions terminology 'criteria for discontinuing SICP' (SICP should never be discontinued). 	Amended
Line 1348	Example of IPC precautions terminology, it then goes on to refer to contact precautions.	Amended
Line 1386	Example of IPC precautions terminology, term used is isolated (nothing wrong with that but no consistency of language).	Amended
Line 1501	Why gowns? So is the advice to use contact precautions or standard precautions? It is not clear.	Amended
Line 1598	Conflicts with other evidence presented/recommendations that suggests isolation or ESBLs not practical. Evidence cited seems to be largely based on multiple interventions in outbreaks, therefore may not be very robust to support strong statement.	Amended

Section		
	Comments	
Line 1611	There are other much better source references of this evidence that can be cited. The general statement is not necessary for Gram-negative bacteria guidance.	Amended
Line 1726	Evidence for efficacy of ATP in preventing transmission is not sufficiently robust to support recommendations.	Changed to weak
Line 1737	The evidence for the efficacy of cleaning is systematically reviewed in EPIC 3. There is limited robust evidence. This section would be better focusing on strategies to ensure decontamination of the environment relevant to Gram-negative bacteria rather than sampling it.	There is already a decontamination section
Line 1752	Very-poor-quality evidence.	Amended
Line 1767–1776	See previous comments about review of evidence re <i>P. aeruginosa</i> .	Recommendation amended to include other
Line 1845	Example of England only: the NHS constitution is England only.	Added 'In England'
Line 1916	Studies on H ₂ O ₂ are not associated with low risk of bias (see EPIC 3). H ₂ O ₂ may be a useful strategy for eliminating environmental reservoirs of MDR Gram-negative bacteria in some circumstances (e.g outbreaks of <i>Acinetobacter</i> spp.). Not sure there is sufficient evidence to say definitively it is effective in reducing environmental reservoirs.	Amended
Line 2002	The Code of Practice and Care Quality Commission are England only.	Amended
Line 2106	Source of this grading system not clear (previously referred to SIGN). Not clear how recommendations are linked to quality of evidence/balance between desirable/undesirable effects/values and preferences/costs as per GRADE.	Changed to updated 2014 SIGN system – s
Line 2109	Table IV good but needs heading on each page and best to have separate column for	Not changed

Section		
	Comments	
	ESBL/carbapenem-resistant Enterobacteriaceae throughout.	
Line 2024	The recommendation is England only.	Amended
Table	Not clear what 'other room or cohort' refers to. What are strict contact precautions?	Amended
Table	Why contact precautions when previously said standard precautions OK for ESBLs? Why soap and water? Why not alcohol hand gel?	Amended
Table	But what about respiratory equipment?	Amended
Table	Definition/evidence for increased cleaning frequency? What about other forms of environmental decontamination (e.g hypochlorite)?	New recommendation introduced
Table	Staff cohorting – extremely difficult, very costly and not relevant for ESBLs.	Agreed

Closing date: Please forward this electronically by 5pm on 31st October 2014 at the very latest to consultations@his.org.uk

Healthcare Infection Society

Consultation – Joint Working Party on multi-drug-resistant Gram-negative infections: prevention and control

Closing date: 5pm 31st October 2014

Organization	NHS Western Isles
Title (e.g. Dr, Mr, Ms, Prof)	Ms
Name	Jennifer Macdonald
Job title or role	ICM
Address and post code	NHS Western Isles
	Western Isles Hospital
	Macaulay Road
	Stornoway
Telephone number	01851 708 399
Email address	Jennifer.macdonald1@nhs.net
Please note: comments will only be accepted electronically on this proforma.	

Please provide comments on the draft guideline on the form below, putting each new comment in a new row. When feeding back, please note the section you are commenting on (for example, Section 1 Introduction and line number). If your comment relates to the guideline as a whole, please put 'general'. Add extra rows if required.

Section		
	Comments	
Section 7.3.4	Page 44 Line 1287 – describes SICPs – should this be 'transmission-based precautions (TBPs)' and again in line 1300. We should be using the same language as in the National Infection Prevention Control Manual.	Amended to single room, contact precautions, avoiding TBP as include droplet and airborne as well as contact
Section 7.4.2.6	Page 46 Line 1348 contact precautions – should be TBPs.	TBP include droplet and airborne, not implied here
Section 7.4.3.2	Page 50 Line 1469 Should read 'SICPs'.	Already there
Table 4	Page 81 Clinical practice – contact precautions should read TBPs, apron and gloves should read TBPs.	TBP include droplet and airborne, not implied here
Table 4	Page 83 Hydrogen peroxide – the recommendation in the literature for this is only weak. National Manual stipulates 'a combined detergent disinfectant solution at a dilution (1000 ppm av.cl.); (Actichlor plus)' there is no evidence within this document that this is not a sufficient method of cleaning. Page 63 Line 1892 notes that there are limited data on whether hydrogen peroxide reduces rates of acquisition. Not all boards will have access to hydrogen peroxide. If this is a method of cleaning that will be recommended, this should be included within the national manual. Otherwise, Actichlor plus is a system all boards are familiar with and currently use effectively to reduce cross-infection and in outbreak situations.	Amended to qualify use of fogging. Disinfectants are discussed in text
Table 4	Page 84 Staff cohorting recommended. Page 49 Line 1459 notes no studies have	Amended

Section		
	Comments	
	evaluated the impact of cohorting staff aside from other interventions. Staff numbers will not be sufficient to allow staff cohort with no evidence this is necessary. Why is this a recommendation? Staff education yes.	

Closing date: Please forward this electronically by 5pm on 31st October 2014 at the very latest to consultations@his.org.uk